

## Research Article

## Role of Katankateryadi Kwatha in Insulin Secretion and Restoration of Biochemical Changes in Streptozotocin-Nicotinamide Induced Diabetes Mellitus Type 2 in Rats

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### Abstract

Diabetes Mellitus Type 2 is a metabolic disorder characterized by hyperglycemia with major and minor complications. The major cause of diabetes mellitus type 2 is lack of insulin secretion or insulin resistance. Experimentally the diabetes mellitus type 2 is induced by several chemicals like Streptozotocin (STZ). The nitrosourea moiety of STZ is responsible for producing beta cell toxicity which leads to development of diabetes mellitus type 2 in rats. The experimental study includes the formulation of Katankateryadi Kwatha, characterization of Kwatha, acute oral toxicity study of Kwatha, OGTT and antidiabetic effect of Katankateryadi Kwatha in STZ-Nicotinamide induced diabetes in rats. Diabetic group treated with 200 mg/kg (189.34 ± 3.89 mg/dL FBG). Post prandial blood glucose (PPBG) level in Diabetic group treated with 200 mg/kg of Katankateryadi Kwatha shows better reduction in PPBG 159.35 ± 6.29 mg/dL in comparison to diabetic rats treated with 100 mg/kg (180.34 ± 2.45 mg/dL). Rats treated with 200 mg/kg Katankateryadi Kwatha significantly reduced the SGOT and SGPT level (133.34±2.25 IU/L and 170.89±3.76 IU/L). The group treated with Katankateryadi Kwatha (200 mg/kg) treated group increased the HDL level (23.56 ± 5.23) and decreases the TC (112.56 ± 3.24), TG (71.65 ± 5.26), LDL (52.41 ± 2.11) and VLDL (21.05 ± 4.62). C-Peptide data result shows that C-Peptide decreased in diabetic rats 0.36 ± 0.034 ng/dl. Katankateryadi Kwatha is useful in management of diabetes mellitus type 2.

**Keywords:** Diabetes Mellitus Type 2; Streptozotocin; Nicotinamide; OGTT; Katankateryadi Kwatha; Glibenclamide; Gliclazide; C-Peptide

## 1. Introduction

Diabetes Mellitus is a metabolic disorder characterized by hyperglycemia followed by polyurea, polyphagia and polydipsia with vascular complications. The diabetes mellitus is broadly categorized in four types – Diabetes Mellitus Type I, Diabetes mellitus Type II, Gestational Diabetes and disease associated Diabetes Mellitus. Majority of people now a day suffered from diabetes mellitus type II. It is a major health issue that has been reached at alarming level [1]. Today nearly half a billion people are living with diabetes mellitus worldwide. The estimated projection of diabetes mellitus is about 700 million people suffer from diabetes mellitus. Two-thirds of people with diabetes mellitus type II are live in urban areas and three out of four are of working age [2]. Over four million people aged 20–79 years are estimated to die from diabetes-related causes in 2019. The countries with the largest numbers of adults with diabetes aged 20–79 years in 2019 are in China, India and the United States of America, and are anticipated to increase till in 2030. India is home to the second largest number (77 million) of adult population with diabetes worldwide. With 1.2 million deaths in 2019 (14.1% of all-cause mortality), the South East Asia Region has the second highest number of deaths attributable to diabetes in adults 20–79 years among the IDF Regions [3]. Many influences that affect the prevalence of disease throughout a country and identification of those factors is necessary to facilitate change when facing health challenges. Today modern medicine develops so many drugs for management of diabetes mellitus type II. The common drawback of modern medicine is drug resistance because the drugs are used for life long period. To avoid such types of problems with modern medicines the world now a day look towards the herbal medicines which are safe in use for long period of time. The medicinal plants play a pivotal role in the health care of ancient and modern cultures [4]. The utility of medicinal plants in the management of diabetes mellitus type 2 are rising in the developing countries worldwide. There is enhancement in scientific evidences that supports the claim towards the traditional healers. The research on herbal medicines and their natural products have significant importance in demands and use in few decades and increasing the demand of herbal drugs worldwide. The available ancient literatures show that there are more than 400 plant species showing anti-diabetic activity. Although some of the plants have great reputation in Ayurveda, the indigenous Indian system of medicine, many remain to be scientifically established [5]. Streptozotocin (STZ) is an antibiotic derived from species *Streptomyces achromogenes* and structurally it is a glucosamine derivative of nitrosoourea. STZ first reported the diabetogenic activity in rats in 1963. Like alloxan [6], it also causes hyperglycaemia by cytotoxic action on beta cells of the pancreas. The nitrosoourea moiety of STZ is responsible for producing beta cell toxicity, while deoxyglucose moiety enhances the transport across the cell membrane. The free radicals generation and alteration in the endogenous scavengers of the free radicals have been reported in STZ diabetogenicity [9]. In comparative to synthetic medicines, Ayurvedic formulations are effective, less side effects, broad range of action and relatively low cost, makes polyherbal formulation as a good choice in management of diabetes type 2 [10]. So many polyherbal formulations are reported in ancient Ayurvedic literatures but due to lack of scientific evidences such formulations are still hidden. Katankateryadi Kwatha is a polyherbal formulation of total five ingredients. The aim of this study is to scientifically prove the Katankateryadi Kwatha as antidiabetic polyherbal formulation for public welfare.

## 2. Material and Methodology

### 2.1 Experimental Animal Requirements

Albino rats (Charles foster strain of weight 150-200 g male and female) purchased from registered animal house of the IMS BHU. The proposal was approved from Central Animal Ethical Committee, Reg. No. 542/GO/ReBi/S/02/CPCSEA. Approval letter no is DEAN/2019/IAEC/1242 dated 26.05.2019. The rats were kept in polypropylene cages containing paddy husk bedding. The cages were stored inside a ventilated room of animal house in Department of Dravyaguna, IMS, BHU Varanasi. Each cage contains 6 rats and standard condition was maintained (temperature 20-30°C, humidity 65-70% and 12 hrs light & dark cycle). Rats were feed with standard pellet diet (Ashirwad trade,) and water.

### 2.2 Chemicals Requirements

Streptozotocin (STZ) Sigma, Nicotinamide (NAD) SRL, SGPT kit SRL, SGOT kit SRL, HDL kit Himedia, Total cholesterol kit Himedia, C-Peptidase kit Himedia, LDL kit, Katankateryadi Kwatha, Glibenclamide Himedia, Gliclazide Himedia.

### 2.3 Herbal crude drug requirement

*Barberis aristata* DC (Stem), *Glycyrrhiza glabra* Linn (Stem), *Terminalia chebula* Tetz(Fruit), *Terminalia bellirica* Roxb (Fruit), *Embllica officinalis* Gaertn (Fruit) and *Plumbago zeylanica* Linn (Root). All crude drugs was authenticated from Department of Dravyaguna, Faculty of Ayurveda IMS, BHU, Varanasi (Reference No: DG/21-22/313-318).

### 2.4 Preparation of decoction of Katankateryadi Kwatha

Katankateryadi Kwatha (decoction) was prepared using six drugs i.e. *Barberis aristata* DC (Stem) [11], *Glycyrrhiza glabra* Linn (Stem) [12], *Terminalia chebula* Tetz (Fruit) [13], *Terminalia bellirica* Roxb (Fruit) [14], *Embllica officinalis* Gaertn (Fruit) [15] and *Plumbago zeylanica* Linn (Root) [16]. All the ingredients of Katankateryadi Kwatha were subjected for size reduction using the pulverizer. Equal amount of all crude drugs was soaked in 4 times water in vessel and kept overnight for 12 hrs. After 12 hrs contents were boiled at 90<sup>0</sup> C – 95<sup>0</sup> C with stirring. Water was evaporated till 1/4<sup>th</sup> amount was remains and galenicals was filtered through cotton cloth. Filtrate was dried with rotatory evaporator and dried powder was used for their quality control standard test and Physico-chemical analysis [17].

### 2.5 Physicochemical evaluation of Katankateryadi Kwatha (Decoction)

Prepared Decoction of Katankateryadi Kwatha and dried powder of decoction was evaluated for their organoleptic properties, microbial contaminations, presence of heavy metals and presence of pesticide residue [18].

### 2.6 Phytochemical analysis of Katankateryadi Kwatha (Decoction)

The Phytochemical screening of different extracts of crude drugs and Kwatha (decoction) were performed according to the procedure mentioned in Ayurvedic Pharmacopoeia of India (API) [19].

## 2.7 Acute Oral Toxicity Study

Acute oral toxicity was performed as per OECD guidelines 423 for toxicity study and estimation of LD<sub>50</sub> dose after treatment with Katankateryadi Kwatha at different doses within 24 hours for dose optimization. Rats were distributed into total four groups and each group contains 5 rats. Group 1 treated with Vidangadi Kwatha 5 mg/kg po, Group 2 treated with Vidangadi Kwatha 50 mg/kg po, Group 3 treated with Vidangadi Kwatha 300 mg/kg po, Group 4 treated with Vidangadi Kwatha 2000 mg/kg po. The rate of mortality and following parameters (Tremor, clonic convulsion, tonic convulsion, straub tail erection, catatonia, ataxia, loss of lighting reflex, sedation, hypnosis, lacrimation, diarrhea, writhing and rate of respiration) were observed for 24 hours [20].

## 2.8 Oral Glucose Tolerance Test (OGTT) in normal and diabetic Rats

The OGTT in normal rats was performed in overnight (18-h) fasted rats. Rats were randomly divided into following groups and each group contains 6 rats: Group 1: received only saline. Group 2 and 3: treated with Katankateryadi Kwatha 100 mg/kg po and 200 mg/kg po. Group 4: treated with Glibenclamide 600 µg/kg po. Similarly the OGTT in diabetic rats was performed and the rats were randomly divided into following groups: Group 1: normal control received only saline. Group 2: diabetic control received only saline. Group 3 and 4: treated with Katankateryadi Kwatha 100 and 200 mg/kg po. Group 5: treated with Glibenclamide 600 µg/kg po. Glucose (2 g/kg) was fed 30 min after the administration of the drugs. Blood sample was withdrawn at each 30, 60, and 120 min after glucose administration and blood glucose was estimated [21].

## 2.9 Effects of Katankateryadi Kwatha in STZ-NAD induced Diabetic Rats

### 2.9.1 Induction of diabetes and treatment

Streptozotocin (STZ) was freshly prepared in citrate buffer (pH 4.5) and Nicotinamide was prepared in normal saline. Hyperglycemia was confirmed by the elevated random blood glucose levels (>200 mg/dL) [22]. Diabetes mellitus type 2 in rats was induced administration of Nicotinamide 120 mg/kg ip followed by administration of 60 mg/kg Streptozotocin (Sigma, Germany) after 15 minute. The rats with fasting plasma glucose >126 mg/dL & post prandial blood glucose level > 200 mg/dL were used in the study. The rats were randomly grouped in to following groups and each group contains 6 rats: Group 1: normal control received normal diet and water. Group 2: per-see study treated with Katankateryadi Kwatha 100 mg/kg po. Group 3: Diabetic control received normal diet and water. Group 4 and 5: Diabetic rats treated with Katankateryadi Kwatha 100 and 200 mg/kg po. Group 6: Diabetic rats treated with gliclazide 4 mg/kg po. **Biochemical analysis:** During 15 days treatment protocol fasting and post prandial blood glucose was estimated and after 15 days blood were collected and allowed to clot formation. The blood serum was isolated by centrifugation technique at 2500 rpm for 15 min and analyzed for various biochemical parameters -SGOT, SGPT, HDL, LDL, VLDL, Total cholesterol, Triglycerides and C-Peptide level were measured [23].

### 3. Results

#### 3.1 Physicochemical and Phytochemical evaluation of Katankateryadi Kwatha

Dried Katankateryadi Kwatha powder is dark brown in color, astringent-sweet taste and pungent smell. Heavy metals (Arsenic, Lead, cadmium and mercury) [24], microbial contamination [25] and pesticide residue [26] was absent in Katankateryadi Kwatha. Phytochemical screening of Katankateryadi Kwatha was performed and found that carbohydrate, flavonoids, alkaloids, proteins, tannins saponins and amino acids was present and volatile oils are absent in Katankateryadi Kwatha.

#### 3.2 Acute oral toxicity study

The Acute oral toxicity shows that there was no mortality found at dose of 5, 50, 300, and 2000 mg/kg. So The LD<sub>50</sub> dose was greater than 2000 mg /kg. Therefore the dose i.e. 100 mg/kg and 200 mg /kg were safe for treatment.

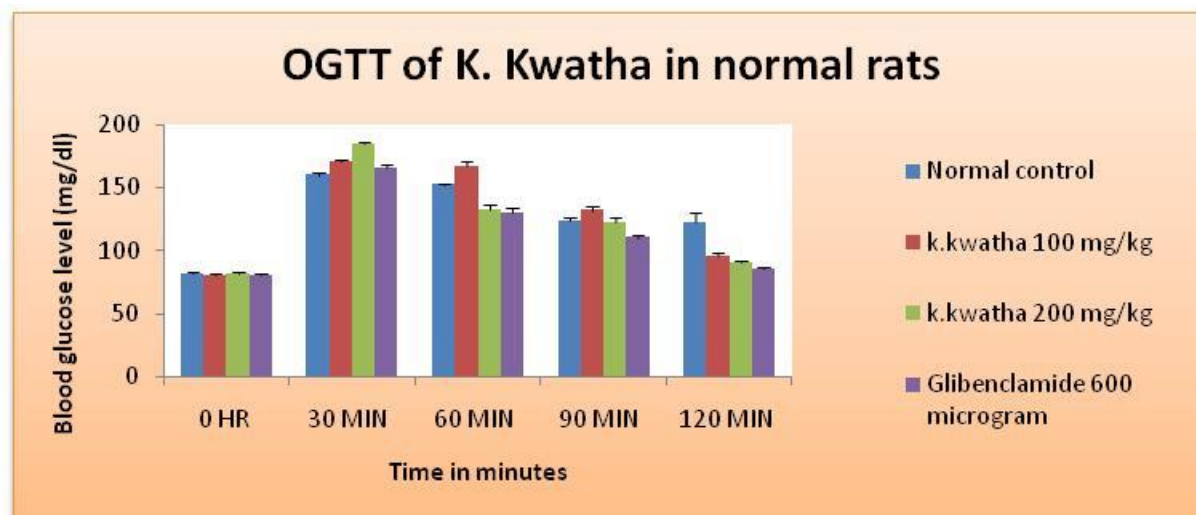
#### 3.3 Blood glucose level during OGTT in normal and diabetic rats

Blood glucose level at different time intervals in OGTT treated with Katankateryadi Kwatha was significantly reduced after treatment compared to the diabetic control group and all treated groups [Table 1, 2 and Fig. 1,2].

**Table 1:** Blood glucose level during OGTT in normal rats.

Groups	Treatment	Blood Glucose Level (mg/dl) in normal rats during OGTT				
		0 Min	30 Min	60 Min	90 Min	120 Min
Group 1	Normal Saline	82.12±1.05	160.12 ± 1.34	152.23 ±1.34	123.54 ± 3.23	122.23 ± 7.63
Group 2	Katankateryadi Kwatha 100 mg/kg po	80.5 ± 2.04	170.32 ± 1.34	167.12 ±3.21	132.13 ± 3.21	95.34 ± 2.65
Group 3	Katankateryadi Kwatha 200 mg/kg po	81.56 ±1.09	184.67 ± 1.89	132.36 ±4.23	122.65 ± 4.21	90.45 ± 1.54
Group 4	Glibenclamide 600 µg/kg po	80.34 ±1.00	165.23 ± 3.21	130.54 ±3.13	110.23 ± 2.36	85.23 ± 1.34

The values represent the mean ± SEM and analyzed by one way ANOVA followed by Dunnett's test. Values are significantly different at the P < 0.05

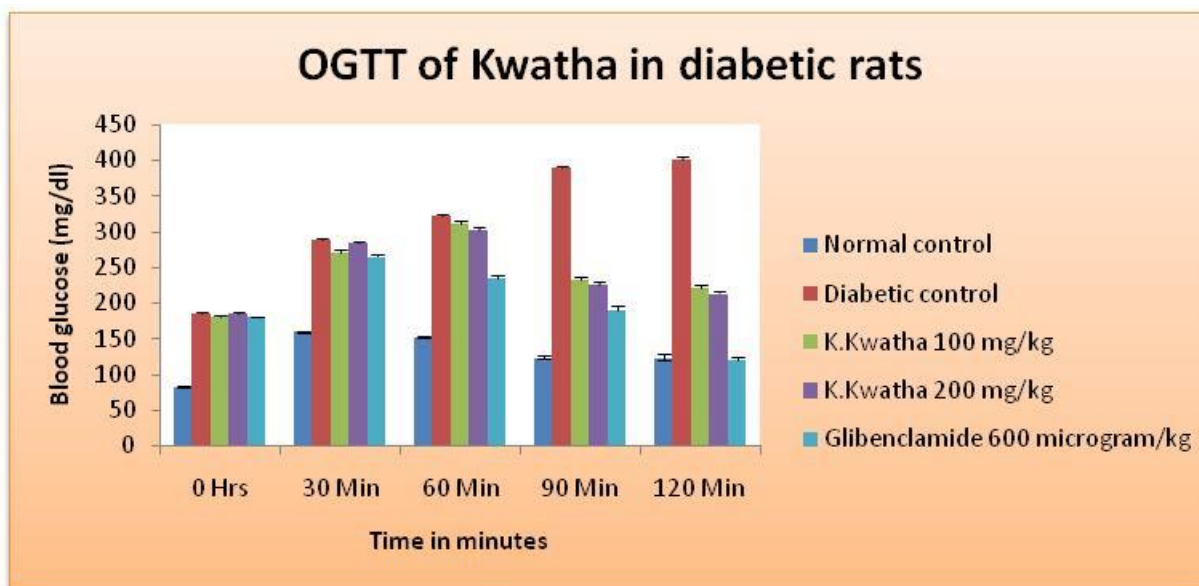


**Figure 1:** Blood glucose level during OGTT in normal rats after treated with Katankateryadi Kwatha and Glibenclamide.

**Table 2:** Blood glucose level during OGTT in diabetic rats.

Groups	Treatment	Blood Glucose Level (mg/dl) in diabetic rats during OGTT				
		0 Min	30 Min	60 Min	90 Min	120 Min
Group 1 NC	Normal Saline	82.12 ± 1.05	160.12 ± 1.34	152.23 ± 1.35	123.54 ± 3.23	122.23 ± 7.63
Group 2 DC	Normal Saline	185.03 ± 2.34	288.12 ± 1.87	322.21 ± 3.12	388.78 ± 3.21	399.98 ± 5.34
Group 3 D	Katankateryadi Kwatha 100 Mg/Kg Po	180.6 ± 3.14	271.34 ± 4.64	310.2 ± 4.26	232.43 ± 3.61	221.54 ± 3.69
Group 4 D	K.. Kwatha 200 Mg/Kg Po	185.58 ± 1.59	284.77 ± 1.83	302.36 ± 5.26	225.67 ± 4.51	213.46 ± 2.44
Group 5 D	Glibenclamide 600 µg/Kg Po	180.44 ± 1.43	265.23 ± 3.21	235.54 ± 3.23	190.24 ± 5.27	120.42 ± 3.14

The values mean ± SEM with in the column and analyzed by one way ANOVA followed by Dunnett's test are significantly different at the  $P < 0.05$ , NC= Normal control, DC = Diabetic control, D = Diabetic Group.



**Figure 2:** OGTT in diabetic rats after treated with Katankateryadi Kwatha and Glibenclamide.

### 3.4 Biochemical analysis after treatment in STZ-NAD induced diabetic rats

#### 3.4.1 Estimation of fasting and post prandial blood glucose

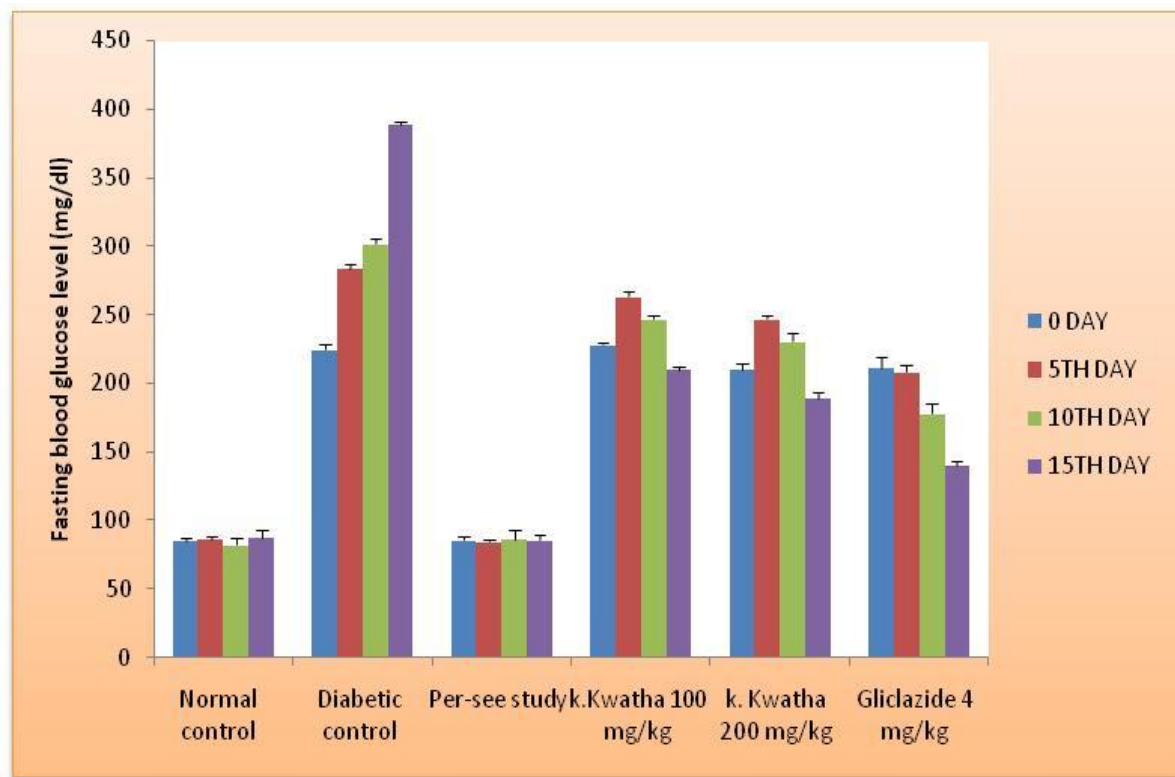
During 15 days treatment protocol diabetic rats shows significant reduction in elevated fasting blood glucose level (FBG) and post prandial blood glucose level (PPBG) in Katankateryadi Kwatha and Gliclazide treated groups. There were no significant changes in normal fasting blood glucose (FBG) level after treatment with Katankateryadi Kwatha in normal rats [28]. Diabetic group treated with 100 mg/kg shows mild reduction in fasting blood glucose level ( $210.34 \pm 1.45$  mg/dL) in comparison to rats treated with 200 mg/kg ( $189.34 \pm 3.89$  mg/dL FBG). Gliclazide group shows FBG level  $140.32 \pm 2.90$  mg/dL after 15 days treatment while diabetic group shows  $388.64 \pm 2.53$  mg/dL FBG level (Table 3, Fig. 3). Post prandial blood glucose (PPBG) level in diabetic group shows  $401.54 \pm 2.13$  mg/dL and per-see group shows  $125.89 \pm 3.34$  mg/dL. Diabetic group treated with 200 mg/kg of Katankateryadi Kwatha shows better reduction in PPBG  $159.35 \pm 6.29$  mg/dL in comparison to diabetic rats treated with 100 mg/kg ( $180.34 \pm 2.45$  mg/dL). The Gliclazide treated group shows  $140.32 \pm 2.90$  mg/dL PPBG (Table 4, Fig. 4).



**Table 3:**Fasting Blood Glucose Level in STZ-NAD Induced Diabetic Rats during treatment.

S.N.	Groups	Fasting Blood Glucose Level (mg/dl)			
		0 Day	5 <sup>th</sup> Day	10 <sup>th</sup> Day	15 <sup>th</sup> Day
1	Normal control	85.12± .31***	86.23±1.74***	82.37±4.76***	87.23 ± 6.35***
2	Diabetic control	225 ± 3.25	284 ± 2.78**	302 ± 3.78**	388.64 ± 2.53**
3	Per-see study	85.88±2.89***	84.67±1.78***	86.34± 6.34***	85.89 ± 3.78***
4	Katankateryadi Kwatha 100 mg/kg po	228.12 ± 54*	263.23 ± 3.54*	247.12 ± 2.12*	210.34 ± 1.45*
5	Katankateryadi Kwatha 200 mg/kg po	210.45± 3.56*	246.76±2.89**	230.78 ± 6.32**	189.34 ± 3.89**
6	Gliclazide 4 mg/kg po	211.56± 7.34*	208.32 ± 4.78**	178.21 ± 7.12**	140.32 ± 2.90**

Values represents in mean ±SEM (n = 6), analyzed by one way ANOVA, \* P<0.05, \*\* P<0.001, \*\*\*P<0.0001 compared with diabetic control.



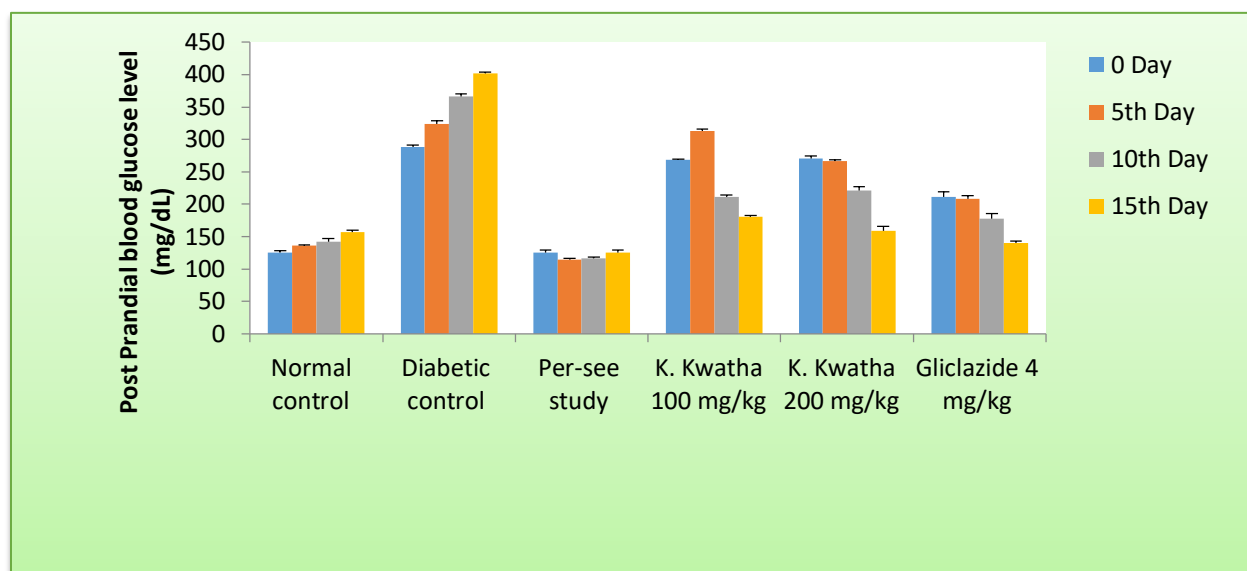
**Figure 3:** Fasting Blood Glucose Level in STZ-NAD induced Diabetic Rats during Treatment.



**Table 4:** Post Prandial Blood Glucose Level after treatment in STZ-NAD Induced Diabetic Rats during treatment.

S.N.	Groups	Post Prandial (PP) Blood Glucose Level (mg/dl)			
		0 Day	5 <sup>th</sup> Day	10 <sup>th</sup> Day	15 <sup>th</sup> Day
1	Normal control	125.12 ± 3.31**	136.23 ± 1.34**	142.37 ± 4.56**	157.23 ± 2.55**
2	Diabetic control	288.44 ± 3.23*	324.11 ± 4.28*	366.12 ± 3.72*	401.54 ± 2.13*
3	Per-see study	125.38 ± .69***	114.67 ± 1.73***	116.44 ± 2.35***	125.89 ± 3.34***
4	Katankateryadi Kwatha 100 mg/kg po	268.17 ± 1.34*	313.23 ± 2.54*	211.22 ± 3.13*	180.34 ± 2.45*
5	Katankateryadi Kwatha 200 mg/kg po	270.42 ± 4.26*	266.46 ± 2.44**	220.78 ± 6.12**	159.35 ± 6.29 ***
6	Gliclazide 4 mg/kg po	211.56 ± 7.34*	208.32 ± 4.78 **	178.21 ± 7.12 *	140.32 ± 2.90 **

Values represents in mean ± SEM (n = 6), analyzed by one way ANOVA, \* P<0.05, \*\* P<0.001, \*\*\*P<0.0001 compared with diabetic control.

**Figure 4:** Post Prandial Blood Glucose (PPBG) Level in STZ-NAD induced diabetic rats

### 3.4.2 Estimation of SGOT and SGPT

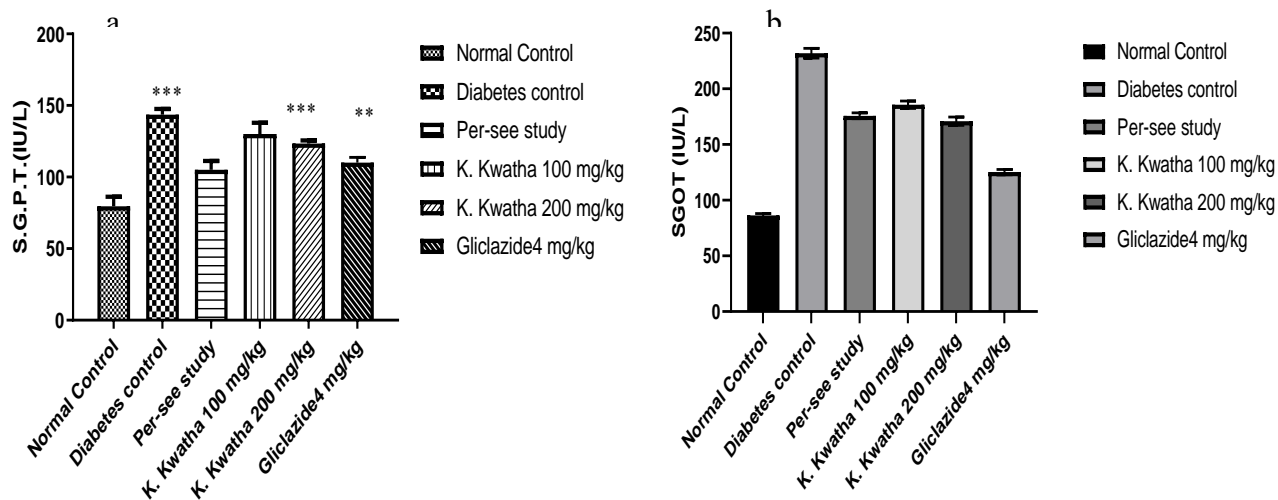
The SGOT (serum glutamate pyruvate tansaminase) and SGPT (serum glutamaic oxaloacetic transaminase) are the liver markers. The increase level of SGOT and SGPT are the indication of hepatotoxicity [29]. The level of SGOT and SGPT reduced to its normal value after treatment with Katankateryadi Kwatha. Diabetic control group shows highest SGOT and SGPT level (143.60 ± 3.94 IU/L and 232.02 ± 4.39 IU/L). Rats treated with 200 mg/kg Katankateryadi Kwatha significantly reduced the SGOT and SGPT level (133.34 ± 2.25 IU/L and 170.89 ± 3.76 IU/L)

when compared with rats treated with 100 mg/kg. Gliclazide treated group shows 110.12±3.65 IU/L SGOT and 125.23±2.36 IU/L SGPT level (Table 5, Fig. 5).

**Table 5:** Liver profile in diabetic rats after treatment with Katankateryadi Kwatha.

Group	Treatment	SGPT (IU/L)	SGOT (IU/L)
1.	Normal control	79.58± 6.66	86.30± 1.72
2.	Diabetic control	143.60±3.94	232.02±4.39
3.	Per-see study	78.06±6.1 ***	85.78±2.6 ***
4.	Katankateryadi Kwatha 100 mg/kg	130.16±7.71***	185.69±3.3***
5.	Katankateryadi Kwatha 200 mg/kg	133.34±2.25**	170.89±3.76**
6.	Gliclazide 4 mg/kg	110.12±3.65	125.23±2.36

Values are Expressed as Mean ± S.E.M.,(n=6) analyzed by one way ANOVA followed by Dunnett’s test \*P value<0.05, \*\*P value <0.01, \*\*\*<P value<0.001 compared with control group.



**Figure 5:** Shows the SGPT and SGOT level in STZ-NAD induced diabetic rats after treatment with Katankateryadi Kwatha and gliclazide

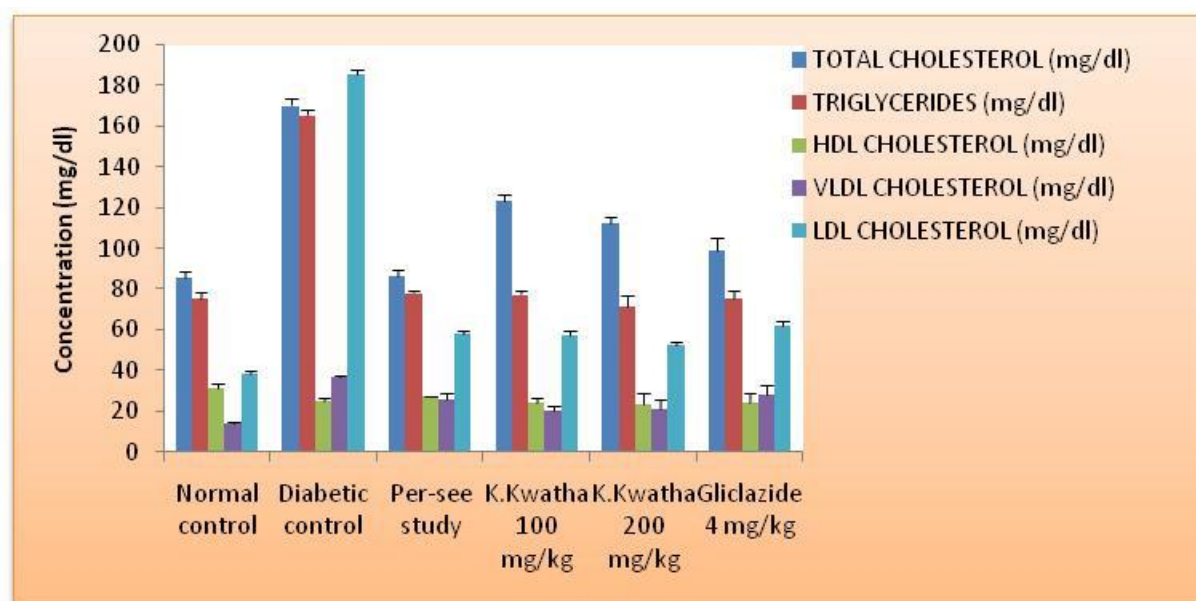
**3.4.3 Estimation of HDL, LDL, VLDL, total cholesterol and triglycerides in STZ-NAD induced diabetic rats**

In lipid profile HDL is good cholesterol and LDL is bad cholesterol [30]. In control group the normal value of TC, TG, HDL, LDL and VLDL are 85.6 ± 3.33, 75.6 ± 2.81, 31.63 ± 2.06, 38.68 ± 1.52 and 14.32 ± 0.39 mg/dL. While in diabetic group rats shows TC, TG, HDL, LDL and VLDL 170.23 ± 3.56, 165.47 ± 2.3, 25.32 ± 0.95, 185.56 ± 2.32 and 36.85 ± 0.65 mg/dL. The group treated with Katankateryadi Kwatha (200 mg/kg) treated group increased the HDL level (23.56 ± 5.23) and decreases the TC (112.56 ± 3.24), TG (71.65 ± 5.26), LDL (52.41 ± 2.11) and VLDL (21.05 ± 4.62) level significantly with Gliclazide treated rats (Table 6, Fig. 6).

**Table 6:** Lipid Profile of diabetic rats after treatment.

Group	Treatment	TC (mg/dl)	TG (mg/dl)	HDL (mg/dl)	VLDL (mg/dl)	LDL (mg/dl)
1.	Normal control	85.6 ± 3.33	75.6 ± 2.81	31.63 ± 2.06	14.32 ± 0.39	38.68 ± 1.52
2.	Diabetic control	170.23 ± 3.56	165.47 ± 2.3	25.32 ± 0.95	36.85 ± 0.65	185.56 ± 2.32
3.	Per-see study	86.74 ± 2.39	78.14 ± 1.14	27.36 ± 0.12	26.12 ± 3.12	58.45 ± 1.45
4.	Katankateryadi Kwatha 100 mg/kg	123.56 ± 2.56	77.36 ± 2.13	24.32 ± 2.13	20.41 ± 2.53	57.58 ± 2.33
5.	Katankateryadi Kwatha 200 mg/kg	112.56 ± 3.24	71.65 ± 5.26	23.56 ± 5.23	21.05 ± 4.62	52.41 ± 2.11
6.	Gliclazide 4 mg/kg	98.89 ± 6.25	75.45 ± 3.56	24.58 ± 4.15	28.14 ± 5.12	62.35 ± 1.62

Values are Expressed as Mean ± S.E.M.,(n=6) analyzed by one way ANOVA followed by Dunnett's test \* P value<0.05, \*\*P value <0.01,\*\*\*<P value<0.001 compared with control group.

**Figure 6:** Lipid profile of treated diabetic rats.

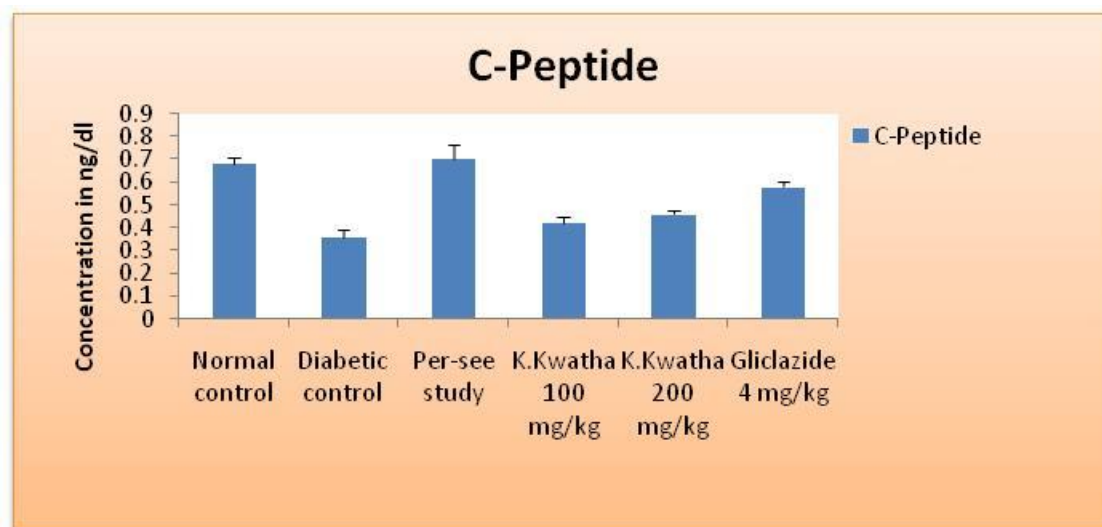
### 3.4.4 Estimation of serum C-Peptide in STZ-NAD induced diabetic rats

After statistical analysis of C-Peptide data result shows that C-Peptide decreased in diabetic rats  $0.36 \pm 0.034$  ng/dl when compared with normal group ( $0.68 \pm 0.025$ ) and treatments with Katankateryadi Kwatha 100 mg/kg, 200 mg/kg and Gliclazide 4 mg/kg significantly increases the C-Peptide level ( $0.42 \pm 0.031$ ,  $0.46 \pm 0.015$ ,  $0.58 \pm 0.025$ ) (Table 7, Fig. 8). The C-Peptide level is the marker of insulin secretion from beta cells of pancreas [31]. The Katankateryadi Kwatha at dose 200 mg/kg increases the C- Peptide level indicates that Katankateryadi Kwatha having the insulin secretory activity likewise Gliclazide.

**Table 7:**C-Peptide level in rats after treatment

Group	Treatment	C-Peptide (ng/dl)
1.	Normal control	0.68 ± 0.025
2.	Diabetic control	0.36 ± 0.034***
3.	Per-see study	0.70 ± 0.063*
4.	K .Kwatha 100 mg/kg	0.42 ± 0.031**
5.	Katankateryadi Kwatha 200 mg/kg	0.46 ± 0.015**
6.	Gliclazide 4 mg/kg	0.58 ± 0.025***

Values are expressed in mean ± SEM, \*P<0.05 significant value when compared with control, \*\*P<0.001 highly significant compared with diabetic control, \*\*\*P<0.0001 highly significant.

**Figure 7:**C-Peptide level in diabetic rats after treatment.

#### 4. Discussion

Type 2 diabetes mellitus is a complex, heterogeneous, and polygenic disease characterized by hyperglycemia [32]. There are many factors that contribute to the high blood glucose levels in these type 2 diabetes patients. An important factor is the body's resistance to insulin, and the second factor is the falling production of insulin by the cells of the pancreas. Lack of insulin plays a primary role in the metabolic derangements linked to diabetes, and hyperglycemia in turn plays a key role in the complications of the diabetes. Several other types of diabetes mellitus are caused by an interaction of genetics and environmental factors [33]. Diabetes type 2 is also refers to as Madhumeha in Ayurveda. Diabetes is associated with vascular and renal dysfunction characterized by hypertension, dyslipidemia, microalbuminuria, macroalbuminuria and glomerular messangial expansion. Diabetes mellitus may present with characteristic symptoms such as polyphagia, polydypsia, polyuria, blurring of vision and weight loss. In its severe forms, keto-acidosis or a non-ketonic hyperosmolar state may develop and lead to stupor, coma and in the

absence of effective treatment to death. Diabetes patients are 25 times more prone to blindness, 2 times more prone to heart attacks, 2-6 times more prone to stroke and 17 times more prone to kidney damage as compared to non-diabetics [34]. The characteristic feature of type 2 diabetes mellitus is hyperglycemia that is reported by elevation in fasting plasma glucose [ $\geq 7.0$  mmol/l ( $\geq 126$  mg/dL)] and 2-hour postprandial glucose [ $\geq 11.1$  mmol/l ( $\geq 200$  mg/dL)] [35].

Diabetes mellitus type 2 is a syndrome of insufficient insulin secretion or insulin resistance or both. However, insulin secretory capacity is not the major determinant in the development of type 2 diabetes, as neither hyperglycemia nor glucose intolerance develops in insulin-resistant patients. Altered insulin secretion caused by either  $\beta$ -cell dysfunction or reduced  $\beta$ -cell mass or both. However, surgical and chemical reductions of  $\beta$ -cell volume induce functional adaptation of the normal  $\beta$  cell to prevent a rise in fasting glucose or reduction in the 1st phase of insulin secretion, suggesting that  $\beta$ -cell dysfunction is closely associated with the pathogenesis and pathophysiology of DM type 2 [36]. Normal dynamics of insulin secretion selective for glucose stimulation, especially the acute phase of the insulin response, is already reduced in the early stage of type 2 diabetes, patients with IGT, and their first-degree relatives. As the early phase of insulin secretion is determined by the RRP (readily releasable pools), a reduction in its size and/or impairment of signaling for exocytosis of the insulin granules from the RRP (readily releasable pools) may well develop in IGT and the early stage of type 2 diabetes. The RRP (readily releasable pools) is completely depleted in completely developed stage of diabetes mellitus type 2 [37]. Type 2 diabetes, or its antecedent impaired glucose tolerance, is often associated with other disorders, particularly central (visceral) obesity, hypertension and dyslipidemia (characterized by elevated low-density lipoprotein (LDL), triglycerides and decreased high density lipoprotein (HDL) [38]. There is no specific recommendation on the percentage of calories that should come from carbohydrate, protein, and fat. The medications for the treatment of type 2 diabetes have the target to increase the insulin secretion (Sulfonylurea) or increase the peripheral utility of glucose or inhibit the metabolism of carbohydrate and their absorption [39]. The primary mechanism of action of the sulfonylurea drugs is to stimulate insulin release from pancreatic B cells. Sulfonylurea are used in patients with type 2 but not type 1 diabetes, since these medications require functioning pancreatic B cells to produce their effect on blood glucose. The Daruharidra has the Hypoglycemic, anti-cancer, gastro-irritant, anti fatigue; anticoagulant, Antipyretic, local-anesthetic, anti- protozoal, anti-tuberculosis, anti-bacterial, anti-tumor, hypotensive, anti-inflammatory, anti-trachoma, CNS-depressant action also [40].

The drug Yastimadhu is a prominent plant drug recommended in cough, asthma, bronchitis and throat affections e.g. hoarseness of throat and voice abnormalities (including laryngitis and pharyngitis) [41]. Haritaki contains Tanninic acid, Chebulic acid, anthraquinones and polyphenolic compounds. The fruit of Haritaki is bitter and astringent in taste. It is useful in dyspepsia, bilious headache and diarrhea; and applied to the eyes, piles and used as brain tonic [42]. Amalaki fruit is a good source of vitamin C, carotene, nicotinic acid, riboflavine, D-glucose, D-fructose, myoinositol and apectin with D-galacturonic acid, D-arabinosyl, D-xylosyl, L-rhamnosyl, D- glucosyl, D-mandosyl and D-galactosyl residues, embicol, mucic. Chitraka root contains Plumbagin, 3-chloroplumbagin, 3, 3'-bipumbagin, Chitranone, zeylinone, isozeylinone, elliptinone, droserone, chitranone, zeylinone, isozeylinone,

isoshinanolone, maritinone, 4-naphthoquinone, plumbagic acid, seselin, 5-methoxyseselin, suberosin, xanthyletin, xanthoxyletin [43]. Chitraka roots and root bark are bitter, dry, stomachic, carminative, and astringent to the bowels, anthelmintic, alterative; they also cure intestinal troubles, dysentery, leucoderma, inflammation, piles, bronchitis, itching, liver disorders, consumption, ascitis, and anaemia [44].

Herbal medicines are popular because of low cost, minimum side effects and the abundant availability in India due to its varied climatic zones. India has around 45,000 species of plants, out of which 15,000–20,000 plants have proven medicinal value. The physicochemical properties of crude drugs including ash value, extractive value, foreign matter, loss on drying and the Phytochemical study was completed for the standardization of crude drugs [45]. The quality control testing of Katankatareyadi Kwatha was performed and shows there was no microbial contamination in the preparation, absence of heavy metals (lead, arsenic, cadmium and mercury) and the pesticide residue was absence. The Kwatha was dark brown in color, astringent taste and pungent smell due to presence of large amount of flavonoids and tannins. The acute oral toxicity was performed for finding of lethal dose ( $LD_{50}$ ) and effective dose ( $ED_{50}$ ) and found that Katankatareyadi Kwatha was safe and effective at maximum dose ( $>2000$  mg/kg b.w.) in rats. Oral glucose tolerance test was performed in normal and diabetic rats and found that the 200 mg/kg of Katankatareyadi Kwatha was more effective than 100 mg/kg. The Katankatareyadi Kwatha reduced the mean value of 40 mg/dL within 2 hrs of treatment. In addition, diabetic rats have impaired glucose tolerance and the additional load of glucose is found to impair the tolerance further. Therefore, the present research investigated both short-term (i.e., FBG and OGTT) during Katankatareyadi Kwatha treatment. From the results obtained, it was clear that fasting blood glucose concentrations and post prandial blood glucose values were dramatically elevated together with oral glucose tolerance in diabetic rats. Katankatareyadi Kwatha at dose of 200 mg/kg produced a statistically significant decrease in FBG beginning from the 1 hrs and progressing till the end of 2 hrs in comparison to the normal and diabetic control group, which indicated Katankatareyadi Kwatha was more beneficial to the long term glucose blood control. From the data obtained in the OGTT, it was clear that Katankatareyadi Kwatha reduces blood glucose levels after 60 min and 120 min of glucose administration. The test suggested that Katankatareyadi Kwatha acts by increasing peripheral utilization of glucose. STZ-NAD induced Diabetes is reported with alterations in plasma blood glucose lipid and lipoprotein profile. In uncontrolled type 2 diabetes mellitus, there will be an increase in TC, LDL, VLDL and TC with decrease in HDL level which contributes to the coronary artery disease. Raised plasma free fatty acid (FFA) level plays a major role in the pathogenesis of insulin resistance and type 2 diabetes. In the present study marked increase in serum lipids and lipoprotein in diabetic rats was observed in diabetic rats and regular administration of the Katankatareyadi Kwatha for 15 days lowered the TC, TG, FFA, and LDL values while elevate the HDL value. The elevation of bad cholesterol and decreased good cholesterol were restored to near normal level during treatment. Results revealed that Katankatareyadi Kwatha was also effective in hyperlipidemia.

The previous studies show that insulin is the important hormone that regulates blood glucose homeostasis by stimulating the utilization of glucose by liver, muscle and adipose tissue, with stimulation of anabolic processes of glucose, protein and lipid synthesis. Insulin is initially synthesized in beta cells in the form of pro-insulin. In this form the alpha and beta chains of active insulin are linked by a third polypeptide chain called the connecting

peptide, or C-peptide, which are assumed to primarily serve as a facilitator for the formation of active insulin by helping insulin to fold and be cleaved as necessary [47]. STZ-Nicotinamide injection caused diabetes mellitus type 2, which may be due to partial destruction of  $\beta$  cells of the islet of langerhans of the pancreas. Over-production (excessive hepatic glycogenolysis and gluconeogenesis) of glucose and decreased utility of glucose by the tissues are the fundamental basis of hyperglycemia in diabetes mellitus type 2 [48].

This study has revealed that Katankatareyadi Kwatha produced a marked decrease in blood glucose at 200 mg/ kg body weight in normal as well as in diabetic rats after 15 days of treatment protocol. Streptozotocin induces diabetes by free radical generation, which causes destruction of insulin secreting beta cells of the islets of langerhans, resulting in a decrease in endogenous insulin level. The free radical scavenging effect of Nicotinamide reduced the effect of STZ. NAD<sup>++</sup> is the principal metabolite of Nicotinamide. NAD<sup>++</sup> level in beta-cells in pre-diabetic and diabetic is significantly reduced and destruction of beta-cells may occur via oxidative stress. Increased levels of reactive oxygen species in beta-cells may result in oxidative damage to DNA resulting in DNA strand breaks. The enzyme poly ADP ribose polymerase or PARP is believed to play a role in DNA repair in beta cells and the PARP uses NAD<sup>++</sup> as its substrate. In the context of a reduced level of NAD<sup>++</sup> result in cellular apoptosis. Nicotinamide is an inhibitor of PARP and reduced the apoptosis. All of these effects of Nicotinamide play some important role in the possible anti-diabetogenic action of Nicotinamide against the STZ. Nicotinamide by the above mechanism minimize the effects of STZ and helps in partial destruction of beta cells leads to development of diabetes mellitus type 2 [49]. In the present study the data obtained clearly indicate that the administration of Katankatareyadi Kwatha lowers the hyperglycemia and increases the C-Peptide level means increases the synthesis and secretion of insulin. On the basis of the current investigation it was noted that the Katankatareyadi Kwathahas the antidiabetic activity and these results provide pharmacological evidence for its folklore claim as an antidiabetic agent.

## 5. Conclusion

The Katankateryadi Kwatha a polyherbal formulation reduces the blood glucose level significantly in diabetic rats. Kwatha reduces the enzymatic activity of alpha amylase, alpha glucosidase which are responsible for hyperglycemia. Kwatha also inhibit the glucose absorption from dialysis membrane, hence we can says that Katankateryadi Kwatha can be used as a potent antidiabetic formulation for the management of hyperglycaemia.

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