

Research Article

Post-Mendelian Genetic Model in COVID-19

Nicola Picchiotti^{1,2#}, Elisa Benetti^{3#}, Chiara Fallerini^{3,4#}, Sergio Daga^{3,4}, Margherita Baldassarri^{3,4}, Francesca Fava^{3,4,5}, Kristina Zguro³, Floriana Valentino^{3,4}, Gabriella Doddato^{3,4}, Annarita Giliberti^{3,4}, Rossella Tita⁵, Sara Amitrano⁵, Mirella Bruttini^{3,4,5}, Laura Di Sarno^{3,4}, Diana Alaverdian^{3,4}, Giada Beligni^{3,4}, Maria Palmieri^{3,4}, Susanna Croci^{3,4}, Mirjam Lista^{3,4}, Ilaria Meloni^{3,4}, Anna Maria Pinto⁵, Chiara Gabbi⁶, Stefano Ceri⁷, Antonio Esposito⁷, Pietro Pinoli⁷, Francis P. Crawley⁸, Elisa Frullanti^{3,4}, Francesca Mari^{3,4,5}, GEN-COVID Multicenter Study, Marco Gori^{1,9}, Alessandra Renieri^{3,4,5*}, Simone Furini^{3*}

¹University of Siena, DIISM-SAILAB, Siena, Italy

²Department of Mathematics, University of Pavia, Pavia, Italy

³Med Biotech Hub and Competence Center, Department of Medical Biotechnologies, University of Siena, Italy

⁴Medical Genetics, University of Siena, Italy

⁵Genetica Medica, Azienda Ospedaliero-Universitaria Senese, Italy

⁶Department of Biosciences and Nutrition, Karolinska Institutet, Stockholm, Sweden

⁷Politecnico di Milano, DEIB, Milano, Italy

⁸Good Clinical Practice Alliance-Europe (GCPA) and Strategic Initiative for Developing Capacity in Ethical Review-Europe (SIDCER), Leuven, Belgium.

⁹Universite Côte d'Azur, Inria, CNRS, I3S, Maasai

***Corresponding author:** Alessandra Renieri, Medical Genetics Unit, University of Siena, Viale Bracci, 253100 Siena, Italy. Phone: +390577233303; Fax: +390577233325.

Simone Furini, Department of Medical Biotechnologies, University of Siena, Le Scotte Polyclinic-Administrative Building, Viale Bracci, 253100 Siena, Italy. Phone: +390577585730; Fax +390577586173.

Received: 02 November 2021; **Accepted:** 10 November 2021; **Published:** 09 December 2021

Citation: Picchiotti N, Benetti E, Fallerini C, Daga S, Baldassarri M, Fava F, Zguro K, Valentino F, Doddato G, Giliberti A, Tita R, Amitrano S, Bruttini M, Sarno LD, Alaverdian D, Beligni G, Palmieri M, Croci S, Lista M, Meloni I, Pinto AM, Gabbi C, Ceri S, Esposito A, Pinoli P, Crawley FP, Frullanti E, Mari F, GEN-COVID Multicenter Study, Gori M, Renieri A, Furini S. Post-Mendelian Genetic Model in COVID-19. Cardiology and Cardiovascular Medicine 5 (2021): 673-694.

Abstract

Host genetics is an emerging theme in COVID-19. A handful of common polymorphisms and some rare variants have been identified, either through GWAS or candidate gene approach, respectively. However, an organic model is still missing. Here, we propose a model that takes into account both common and rare coding variants. This model has been piloted in a cohort of 1,318 Italian SARS-CoV-2 positive individuals. Ordered logistic regression of clinical WHO grading on age, stratified by sex, was used to obtain a binary phenotypic classification useful for subsequent data mining. Genetic variability from WES was synthesized in several Boolean representations differentiated according to allele frequencies and genotype effect. LASSO logistic regression was used for extracting relevant features, part of them being sex specific. We defined a set of common coding polymorphisms relevant for COVID-19 severity. Another set of rare coding variants were found to contribute either to mildness or severity of COVID-19, which in some cases simulate a Mendelian inheritance. The combined effect of common and rare variants can be described as an Integrated PolyGenic Score (IPGS). This score can be computed using the following formula: $(n_{\text{mildness}} - n_{\text{severity}}) + F (m_{\text{mildness}} - m_{\text{severity}})$. Here n is the number of common driver genes, m is the number of driver rare variants, and F is a factor for an appropriate weighing of the more powerful rare variants. We named this model: “post-Mendelian”. The model has been tested with multiple train-test splits and the polygenic score accurately separated patients with severe COVID-19 from those having a mild form. Furthermore, segregation of IPGS can be observed in familial cases in which more than one infected individual is present.

Keywords: COVID-19; Genetic science modelling; Host genetics; Integrative polygenic score

1. Introduction

Coronavirus disease 2019 (COVID-19) presents an important test case for developing new models for studying complex disorders with a background of combined genetic and environmental factors. Unlike other multifactorial disorders, the main environmental factor for COVID-19, SARS-CoV-2, can easily be identified through PCR-based tests on swab. Assuming a relatively low impact of viral genome variability [1], the remaining variability in clinical outcome may likely be associated with age and host genetics, including sex. Genome-Wide Association Studies (GWASs) have identified a certain number of common polymorphisms in relevant genes [2-4]. However, these associations do not fully explain the variability of clinical outcomes. The candidate gene approach has shown that, as with many other complex disorders, a simple Mendelian inheritance is also found in COVID-19, affecting some rare individuals with defective gene variants related to innate immunity [5-7].

In early COVID-19 research, the Italian GEN-COVID Multicenter Study explored COVID-19 host genetics through Whole Exome Sequencing analysis (WES) in a cohort of 35 hospitalized patients. This research led to the preliminary identification of a combination of rare and common variants that appeared to potentially impact patient clinical outcome [8]. Then, within this Italian GEN-COVID Multicenter Study, biospecimens from more than 1,000 SARS-CoV-2 positive individuals were collected in the GEN-COVID Biobank (GCB), with clinical data stored in the related Patient Registry (GCPR), and genetic data in the connected Genetic Data Repository (GCGDR) [9]. SNP genotyping of this cohort contributed to the identification of some loci associated with COVID-19 [3], while WES analysis pinpointed additional common non bi-allelic polymorphisms [10-12] and rare coding variants [13]. However, an organic model explaining how common variants may combine with rare variants is still lacking. In the study presented in this paper, we propose a new model for describing the severity of COVID-19 using both common and rare coding variants. The

model was defined in two steps: in the first step, logistic regression approach was used to identify a set of genes that are predictive for the severe or the mild phenotype of COVID-19. In the second step of the model, these extracted predictive genes were used to define a score that separates the extreme COVID-19 phenotypes, giving a disease severity prediction.

2. Materials and Methods

2.1. Patient’s cohort and clinical classification

Demographic and clinical characteristics of the data set were identified and published [9]. The number of cases used for this study was 1,318. In order to obtain a clinical classification as independent as possible from age and sex, an Ordered Logistic Regression (OLR) model was used. Separately for the male and female cohorts, two OLR models were fitted using age to predict the ordinal grading (0, 1, 2, 3, 4, 5) dependent variable (Figure 1). Then, each patient was assigned a clinical classification: equal to 0 (mild), if the actual patient grading was below the one predicted by the OLR; or equal to 1 (severe), if the grading was above the OLR prediction. The patients with a predicted grading equal to the actual grading were excluded from the following analyses in order to compare patients where the genetic contribution towards the severe/mild phenotype is expected to be more relevant.

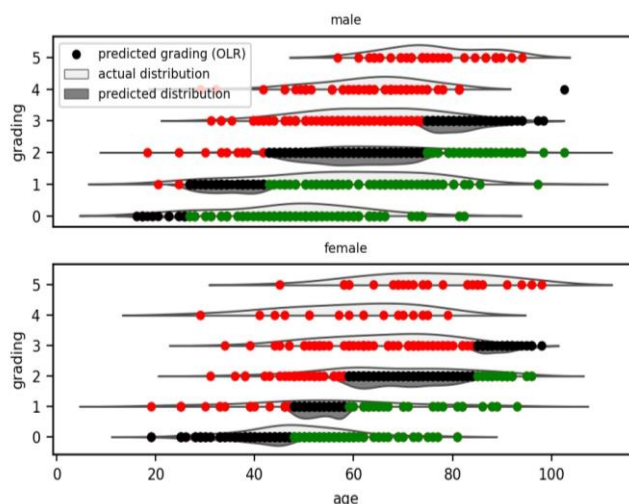


Figure 1: Clinical classification adjusted by age. Two Ordered Logistic Regression (OLR) models, stratified by sex, fitted using age to predict the ordinal grading (0, 1, 2, 3, 4, 5) dependent variable. On the Y axis, the grading according to patients’ treatment is reported (5=deceased; 4=intubated; 3=CPAP/biPAP; 2=oxygen therapy; 1=hospitalized without oxygen support; 0=not hospitalized oligo-asymptomatic patients). On the X axis, age is reported. Red dots represent subjects falling above the expected treatment outcomes according to age (hence considered severe), green dots are subjects falling below the expected treatment outcomes according to age (hence considered mild) and black dots are subjects matching the expected treatment outcomes according to age (hence considered intermediate).

2.2. WES analysis

Whole Exome Sequencing with at least 97% coverage at 20x was performed using the Illumina NovaSeq6000 System (Illumina, San Diego, CA, USA). Library preparation was performed using the Illumina Exome Panel (Illumina) according to the manufacturer's protocol. Library enrichment was tested by qPCR, and the size distribution and

concentration were determined using Agilent Bioanalyzer 2100 (Agilent Technologies, Santa Clara, CA, USA). The Novaseq6000 System (Illumina) was used for DNA sequencing through 150 bp paired-end reads. Variant calling was performed according to the GATK4 best practice guidelines, using BWA for mapping and ANNOVAR for annotating. WES data were represented in a binary mode on a gene-by-gene basis.

2.3. Boolean representation of bi-allelic polymorphisms

In the Boolean representation of bi-allelic polymorphisms, coding variants (missense, non-sense, and ins/del), as well as splicing mutations, with a frequency > 1% as defined in gnomAD for the Non-Finnish European population were considered. Depending on the boolean representation considered (see section 2.6) only homozygous variants, or both homozygous and heterozygous variants were considered. For those genes having more than one common variant, all the unique combinations (mutually exclusive) with a frequency above 5% in the cohort were represented. For instance, *IBSP* gene has 5 coding polymorphisms with 5 combinations and with a frequency above 5% in the cohort. Each combination was represented separately as IBSP_1 to IBSP_5. A further feature, named IBSP_0, was defined to represent the presence of any combination with a frequency lower than 5%. These unique combinations were used to define a matrix of M by N input features (M and N being, respectively, the number of combinations and the number of samples), with the element j,i equal to 1 if the combination of common variants j is present in the sample i .

2.4. Boolean representation of rare variants

Three models were proposed for the binary representation of rare variants: autosomal dominant (AD), autosomal recessive (AR), and X-linked (XL). Only coding variants (missense, non-sense, and ins/del), as well as splicing mutations, were considered, together with any pathogenic mutations (coding and non-coding) having a frequency below 5% (e.g. Phe508del in *CFTR* being 1.1% in the non-Finnish European population). In the AD model, the input feature j,i was 1 if gene j in sample i had at least one variant with a frequency $\leq 1\%$, as defined in gnomAD for the Non-Finnish European population. In the AR model, the input feature j,i was 1 if gene j in sample i had either a variant with a frequency $\leq 1\%$ in a homozygous state or two variants. In the XL model, only genes on chromosomes X were considered. The input feature j,i is 1 if gene j in sample i has a variant with frequency $\leq 1\%$ (hemizygous state in males).

2.5. Sample pre-processing

Among the 1,318 samples, after PCA analysis, 63 outliers were removed. Those corresponded to non-white subjects (32 hispanic, 12 black, 18 asian, 1 arabic). In the Boolean representation of rare variants, the cumulative probability distribution for the number of mutated individuals per gene was estimated, and the elbow method was used to identify the number of mutated individuals per gene that corresponds to a change in the convexity of the cumulative probability. The genes with a number of mutated individuals above this threshold were considered artefacts and not included in further analyses. Among the 18,085 annotated genes, 18 genes were excluded from the analysis due to annotation errors, such as the *ARSD* and the *VCX* family genes.

2.6. LASSO logistic regression

The binary classification problem, i.e., mild/severe cases, was solved by a logistic regression model, one of the most common and successful Machine Learning (ML) algorithms for binary classification tasks with probabilistic interpretation. For each sex, 6 separate logistic models were defined using the following input features: 1) common bi-

allelic coding haplotypes of autosomal genes (hetero plus homo versus wt) extracting common variants acting in a heterozygous state; 2) common bi-allelic coding haplotypes of autosomal genes (homo versus hetero and wt) extracting common variants acting in a homozygous state; 3) common bi-allelic coding haplotypes of X-linked genes extracting common variants acting in a hemizygous state in males or as X-linked dominant in females; 4) rare variants of autosomal genes (hetero plus homo versus wt) extracting rare variants acting in heterozygous state; 5) rare variants of autosomal genes (homo versus hetero and wt) extracting rare variants acting in a homozygous state; and 6) rare variants of X-linked genes extracting rare variants acting in a hemizygous state in males or as X-linked dominant in females. In order to enforce both the sparsity and the interpretability of the results, the model was trained with the additional LASSO (Least Absolute Shrinkage and Selection Operator) regularization term. By denoting with β_k the coefficients of the logistic regression model, and by denoting with λ the strength of the regularization, the LASSO regularization term of the loss, $\lambda \sum_{k=1}^p |\beta_k|$, has the effect of shrinking the estimated coefficients to 0, providing a feature selection method for sparse solutions within the classification task. The weights resulting from the logistic regression algorithm can be interpreted as the feature importances for the task [14].

The fundamental hyper-parameter of the logistic regression algorithm is the strength of the LASSO term, which was tuned with a grid search method on the average accuracy for the 10-fold cross-validation. 50 equally spaced values in logarithmic scale in the range $\lambda \sum_{k=1}^p |\beta_k|$ were tested. The optimal regularization parameter was selected as the one with the best trade-off between the simplicity of the model and the cross-validation score, i.e., as the highest value providing an average score close to the highest score with its half standard deviation. The method is designed to select the most important genes (and not necessarily the entire set of genes contributing to COVID-19 variability). Data preprocessing was coded in Python. For the logistic regression model, the scikit-learn module with the liblinear coordinate descent optimization algorithm was used. The entire cohort was splitted into a training set (90%) and a testing set (10%). The model has been tested with multiple train-test splits.

2.7. The post-Mendelian model

The proposed model aims at combining the genetic knowledge extracted from the individual LASSO logistic regression models by extending the standard “threshold model” of common polymorphisms to a more general framework, including the effect(s) of rare variants. The main hypothesis of the model is that it is possible to define an Integrated PolyGenic core (IPGS) by counting the number of variants in genes promoting the severe or the mild phenotype, assigning a different weight to common and rare variants.

The following formula calculates the IPGS as:

$$\text{IPGS} = (n_{\text{mildness}} - n_{\text{severity}}) + F(m_{\text{mildness}} - m_{\text{severity}})$$

In the equation above, n_{mildness} and n_{severity} are the number of common polymorphic driver genes conferring mildness or severity (respectively) to COVID-19; while m_{mildness} and m_{severity} are the number of driver rare variants conferring severity or mildness (respectively) to COVID-19 (Figure 2).

$$IPGS = (n_{mildness} - n_{severity}) + F (m_{mildness} - m_{severity})$$

Number of common mildness coding variants/haplotypes → $n_{mildness}$
 Number of common severity coding variants/haplotypes → $n_{severity}$
 Number of rare mildness variants → $m_{mildness}$
 Number of rare severity variants → $m_{severity}$
 Integrated Polygenic Score → IPGS
 Multiplicative factor for increasing the weight of rare variants → F

Figure 2: The determinative formula for the post-Mendelian model. This formula determines the integrated polygenic score (IPGS). The first term is the difference between the number of common variants/coding haplotypes conferring mildness ($n_{mildness}$) and severity ($n_{severity}$) to COVID-19; whereas the second term is the difference between the number of rare variants conferring mildness ($m_{mildness}$) and severity ($m_{severity}$) to COVID-19. The multiplicative factor F was included to model the penetrant effect of rare variants in comparison to common variants.

The genes conferring mildness or severity were identified using the LASSO logistic regression models described in section 2.6. Boolean representations 1-3 were used to define $n_{mildness}$ and $n_{severity}$, and boolean representations 4-6 were used to define $m_{mildness}$ and $m_{severity}$. IPGS value calculated for each sex excludes genes specific of the other sex. In order to identify the optimal value of the F factor, for each value of F in the range 0-5, the silhouette coefficient of the clustering (mild vs severe) results was calculated. The optimal value of F was defined as the one that maximizes the silhouette coefficient, i.e., provides the best separation between the two clusters. The entire procedure is schematized in Figure 3.

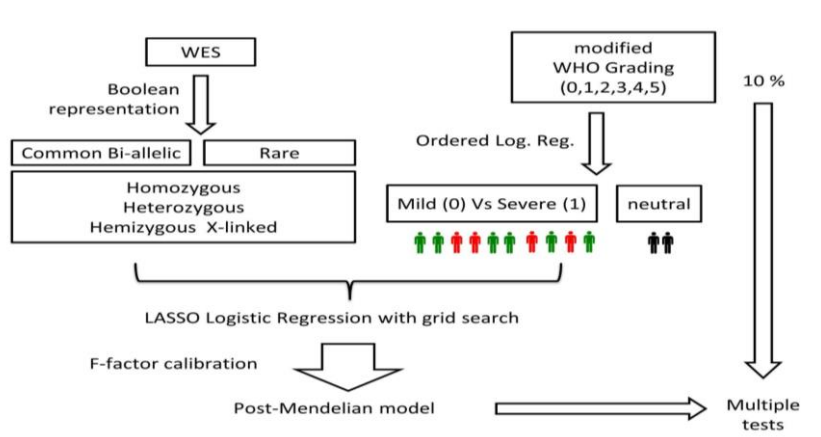


Figure 3: Outline of the method. Separately for the male and female cohorts, two OLR models were fitted using the age to predict the ordinal grading (0, 1, 2, 3, 4, 5) dependent variable. Then, each patient was provided a clinical classification equal to 0 (green), if the actual patient grading was below the one predicted by the OLR, or 1 (red), if the actual patient grading was above the OLR prediction. The patients with a predicted grading equal to the actual grading (black) were excluded from the LASSO analysis. LASSO Logistic Regression was performed on WES data represented in a Boolean manner, including common and rare variants, separately for males and females. Following the analysis

provided on the basis of the WES data analyzed through LASSO Logistic Regression, the post-Mendelian model was applied. For each subject then, an integrated polygenic score (IPGS) was calculated.

2.8. Bootstrap analyses

In order to have more robust feature selection the dataset was divided into a training set and a testing set (90/10), and the feature selection was performed using only samples in the training set. A bootstrap approach with 82 iterations was adopted (Supplementary Tables 1-4). At each bootstrap iteration, 90% of the samples were selected (without replication), to identify the most relevant features for each Boolean representations (Supplementary Tables 1-4).

3. Results

3.1. Assessing clinical classification stratified by sex and adjusted by age

Whole Exome Sequencing (WES) data stored in the Genetic Data Repository of the GEN-COVID Multicenter Study (GCGDR) and coming from biospecimens of 1,318 SARS-CoV-2 PCR positive subjects were used for the analysis [9]. Because age and sex are strong determinants of the clinical outcome, we stratified the cohort by sex and then applied ordered logistic regression to re-classify the patients (Figure 1) as follows: i) severe: subjects falling above the expected treatment outcomes according to age; ii) intermediate: subjects matching the expected treatment outcomes according to age; iii) mild: subjects falling below the expected treatment outcomes according to age. This novel classification is expected to be reliable independent of age. If this is further demonstrated, it should then facilitate the reliable identification of genetic factors responsible for COVID-19 severity as well as potentially identifying (druggable) pathways to treatment.

3.2. The advantage of using several Boolean representations

We reasoned that, in order to identify possible Mendelian-like COVID-19 disorders, different rare variants in the same gene be classified as 1 or 0 (0 being the absence of variants). In order to detect the underlying Mendelian-like mode of inheritance, genes on chr X were considered separately. This representation is able to extract X-linked recessive genes on males (Figure S1q), such as *TLR7* which is now functionally validated by three independent studies [6,7,13]. Likewise, in order to distinguish autosomal dominant from autosomal recessive disorders, two different Boolean representations were considered for the remaining autosomal genes: at least one variant equal for the dominant model and at least 2 variants for the recessive models (Figures S1i-S1p). Furthermore, we reasoned that for the contribution of common polymorphisms, based on the chromosomal location (X versus autosomal) and the genotype (at least one variant versus at least 2 variants), three different Boolean representations were better suited to identify common variants (Figures S1a-S1h). These representations were able to extract autosomal genes acting on specific sex with a specific genotype. For example, using at least one variant in males, LASSO pinpoints the p.Leu412Phe polymorphism in the testosterone-repressed *TLR3* gene (Figure S1b). This is an already known functional polymorphism [15], which is now associated to COVID-19 by two independent studies [11, 16], proved to reduce TNF alpha and autophagy [11].

3.3. Discovery of genes and sex dependent effect

We used severe versus mild cases defined by ordered logistic regression of the training set as inputs for a series of logistic regression models using LASSO regularization. The purpose of these logistic models was to identify which features are better predictors of either severe or mild disease. The six different types of genetic variability identified

above were represented in a Boolean manner and tested separately. The full list of genes identified in both sexes are reported in Supplementary Tables 1-4.

Using the cohort inclusive of both sexes, for common bi-allelic haplotypes (hetero plus homo versus wt), we identified a set of genes ordered by importance (Figure 4). We then repeated the analysis using sex stratification (Figure 4). We found that a part of them are sex-specific, i.e., found in one sex only (Figure S1a, Figure S1b and S1c, Supplementary Tables 1-4). A similar pattern was found for all other Boolean representations.

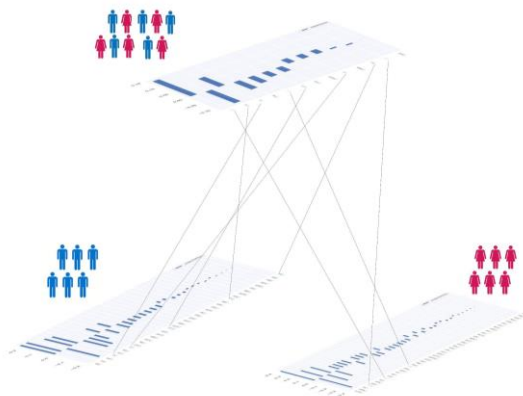


Figure 4: Sex specific effect. Upper panel: genes with common bi-allelic haplotypes (hetero plus homo versus wt), ordered by importance in the cohort that includes both sexes. Down panel: genes with common bi-allelic haplotypes (hetero plus homo versus wt), ordered by importance and stratified by sex (left side: genes in males; right side: genes in females).

3.4. Discovery of relevant genes with common variants

Applying LASSO logistic regression to the Boolean representation of common bi-allelic haplotypes, we identified a set of relevant genes for COVID-19 severity (Figure S1a-S1h and Supplementary Tables 1 and 2). Many genes were found in both sexes (Figure S1a, S1d and Supplementary Tables 1 and 2) and a set of them only affect one sex (Figure S1b, S1c, S1e, S1f, S1g, S1h and Supplementary Tables 1 and 2). Among the relevant common variants associated with severe disease, there are some genes already functionally validated and reported associate with severe COVID-19 disease such as *TLR3* with p.Leu412Phe polymorphism in heterozygous state [11,16] that controls interferon response; and *SELP* with p.Asp603Asn polymorphism in homozygous state [12], a cell adhesion molecule which mediates the activation of platelets and endothelial cells and its soluble isoform is increased in both venous and arterial thrombosis. Both *TLR3* and *SELP* are testosterone regulated genes and in fact impact only on males patients.

3.5. Discovery of relevant genes with rare variants

Using LASSO logistic regression on the Boolean representation of rare variants, we identified a set of relevant genes for COVID-19 severity (Figure S1i-S1r, Supplementary Tables 3 and 4). Many genes were found in common with both sexes (Figure S1i, S1n and Supplementary Tables 3 and 4). Among the relevant genes extracted only in males with rare variants associated with severe disease, there is the already known *TLR7* involved in controlling interferon response

[6,7,13]. Among relevant genes with rare variants associated with severe diseases in females there is *TLR5* (hetero) previously involved in bacterial infection and more recently also in viral infection.

3.6. The post-Mendelian paradigm for COVID-19 modelization

In the previous sections, the protocol used to identify predictive genes for the severe and mild phenotypes were presented and the plausibility of these genes is supported by biological evidence. This agreement leads to plausible biological knowledge based on two independent modelling approaches. The apparent strong confirmation of our hypothesis convinced us to further explore the possibility of combining the derived variants' information regarding these genes into a predictive model of phenotype expression. The proposed model extends the standard “threshold model” of common polymorphisms to a more general framework, including both the effect of common and rare variants. In the case that each common variant carries the same relative risk, subjects with approximately the same number of common variants in genes indicative of mild or severe will behave according to their age (recall the black dots in the ordered logistic regression of Figure 1). In other cases, subjects with unbalanced variants (that is, with a higher number of mild- or severe disease gene variants) will more likely belong to the corresponding phenotype, with a probability that increases as the difference between the number of mild- and severe gene variants increases. By looking at the epidemiological data of the total number of those who are severely ill (intubated or CPAP-BiPAP), one should be able to infer the exact number of genes involved in the model and the exact percentage necessary for the threshold effect. In addition to the cumulative effect of common variants, another group of genes may be affected by a rare mutation. According to an autosomal dominant, autosomal recessive, or X linked mode let us suppose a rare mutation occurring in either a neutral or severe background of common variants. In such cases, a more severe or earlier disease segregating as a Mendelian inheritance may be identifiable. Family members are also likely to have identical or similar combinations of common variants. If a rare variant is identified against a mild disease background, the penetrability of the virus may likely be dependent on the strength of the rare variant in warding off viral replication. If a mildness rare occurs in a neutral or mildness background, the individual will be even more likely to exhibit a mild phenotype (for example, an individual with the persistence of antibodies far away from the vaccine or infection). If rare mildness variants occur in a harmful background, its effect will likely depend on the strength (relevance) of mildness (the type of gene and its mutation as well as the location of the gene in the pathophysiological process. For example, individuals with rare variants in *FACLA* (which prevents the accumulation of lipid microparticles necessary for the development of the virus) could have milder symptoms even in a negative background (because the action is upstream). On the other hand, some individuals are asymptomatic following infection but continue to test positive for SARS-CoV-2 for a significant period without developing COVID-19. These could be individuals with common mildness variants but “severity rare mutations” favoring the spread of the virus on the tissues. However, the downstream process appears milder because they respond well by innate and adaptive immunity. The number of rare variants that individuals in the pilot study were found to have varied from 0 to 10, with an average of 3. Some individuals without rare variants were also found to present with or develop severe disease, indicating that common variants alone could be involved in affecting disease severity. In individuals with rare variants, the correct prediction of disease severity required a consideration of both common variants and rare variants. In cases of individuals having more than one rare variant gene in the same variant category (e.g., mild or severe), the segregation gene variants simulated the oligogenic model (digenic, trigenic, etc.).

3.7. Fitting the model in the cohort and calibration of rare variants contribution

In the cohort of 1,318 SARS-CoV-2 used in the pilot study, we used the post-Mendelian model depicted in the previous section to determine an integrated polygenic score in which rare variants contribute together with common polymorphisms to COVID-19 liability. By counting the mutations of genes extracted by LASSO logistic regression for each patient in the cohort, we were able to obtain the integrated polygenic scores as reported in the frequency distribution shown in Figure 5 separately for severe (red) and mild (green) phenotypes. The calibrated F factor (i.e. the coefficient for which the mean silhouette coefficient is at its highest) is shown in Figure 5 for the male (2) and female (1.2) cohorts. In the overall cohort (male and female cohorts taken together), classical Mendelian inheritance was simulated when rare variants occurred in a medium or severe background (subjects having only 1 susceptibility mutation) and oligogenic inheritance (having 2 susceptibility mutations, having 3 mutations, etc.).

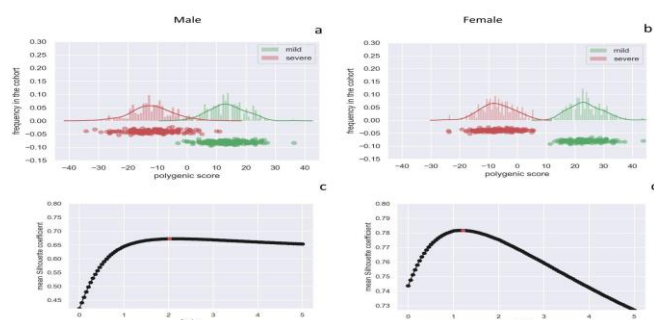


Figure 5: Integrated Polygenic Score, factor calibration, and phenotypes. Frequency distributions of the IPGS score for the male cohort (Panel a) and female cohort (Panel b). The red distribution is related to the severe phenotypes and the green one to the mild phenotypes. Panel c and Panel d report the mean silhouette coefficient of the clustering of mild and severe phenotypes as a function of possible F values of the IPGS formula in the range 0-5. The coefficient is a measure of the goodness of the clustering. The maximum value that provides the best separation between the two clusters is 2 for the male cohort (Panel c) and 1.2 for the female cohort (Panel d).

3.8. Segregation of the post-Mendelian model in families

In the overall cohort 15 familial cases were collected and analysed. Segregation analysis using an integrated polygenic score is in line with the phenotype in each of the families (Figure 6). Through further exploration for rare variants in this individual family members, an extremely rare pathogenic mutation was detected in *IFNAR1*. The frequency, however, of *IFNAR1* variants was too low to identify using logistic regression in a cohort of this size.

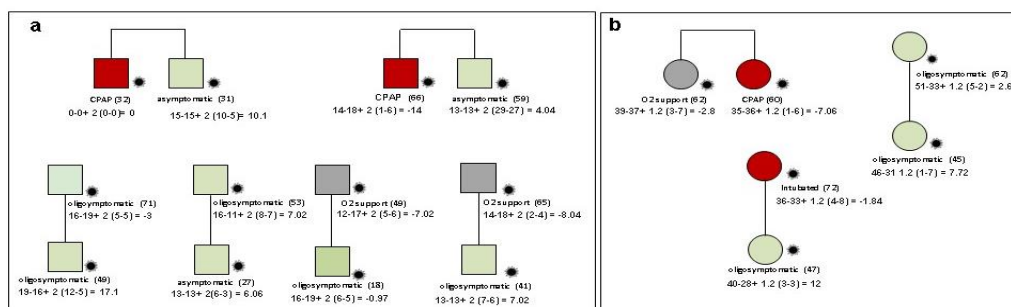


Figure 6: Segregation analysis afforded by the post-Mendelian model. Example of segregation analysis using the post-Mendelian model applying an integrated polygenic score to 9 pedigrees. The squares represent male subjects and the circles represent female subjects. Red = severely affected (category 1 in Figure 1); green = oligo-asymptomatic subjects (category 0 in fig. 1); grey squares represent intermediate subjects (category black in Figure 1). Under each symbol is reported the treatment and (in parenthesis) the age. For each patient the formula for the computation of the integrated polygenic score (IPGS) is reported. **Panel a:** Brothers of 32 and 31 years with discordant phenotypes: hospitalized CPAP treated, and oligosymptomatic, respectively. In agreement with their phenotype, they have IPGS 0 and 10.1 respectively, mainly due to increased common polymorphisms associated with mild disease in the asymptomatic brother (such as p.Ile57Val of *AURKA*, a cell cycle regulator downregulated during SARS-CoV-2 infection; p.Cys357Arg/p.Val335Met of *GBP3*, a strong repressor of the activity of the viral polymerase complex, which results in decreased synthesis of viral proteins; and p.M1? of *TLR8*, a member of the Toll-like receptor family which plays a fundamental role in pathogen recognition and activation of innate immunity). **Panel b:** Sisters of 62 and 60 years of age with partially discordant phenotype, hospitalized with oxygen support only and hospitalized CPAP treated, respectively. In agreement with their phenotype, they have IPGS minus 2.8 and minus 7.05, respectively, mainly due to increased severity of rare variants in the more severely affected sister (including Amyloid Beta Precursor Protein Binding Family A Member 3 *APBA3*, which plays a role in immune response; and the low density lipoprotein receptor family member *LRP8*, which has a role in the suppression of innate response). Treatment with immunosuppressive agents may be an option in this patient.

3.9. Testing the model

In order to test whether the proposed model is able to generalize the predictions to unseen samples, the entire cohort was splitted into a training set and a testing set (10%). The entire training procedure was completely blind to the samples in the testing set, namely, the training set was used to: (i) train the OLR models for predicting the phenotype adjusted by age and sex; (ii) identify the parameters and the regularization strength (by cross-validation on the training set) of the LASSO logistic regression models; (iii) estimate the optimal value of the F score in the IPGS. The predictivity of the model was estimated by using the ROC-AUC. In order to have a robust estimate of the predictivity of the model, the whole procedure of train-test split was repeated 150 times. Then, the distribution of the ROC-AUC scores was compared with the one (null-hypothesis) obtained by the shuffling of the phenotypes. As expected, the average ROC-AUC is centered at 0.5 in the presence of random phenotype. Instead, when the actual phenotypes are adopted the ROC-AUC distribution is significantly shifted towards higher values (p-value of 2.7×10^{-6}) as shown in Figure 7.

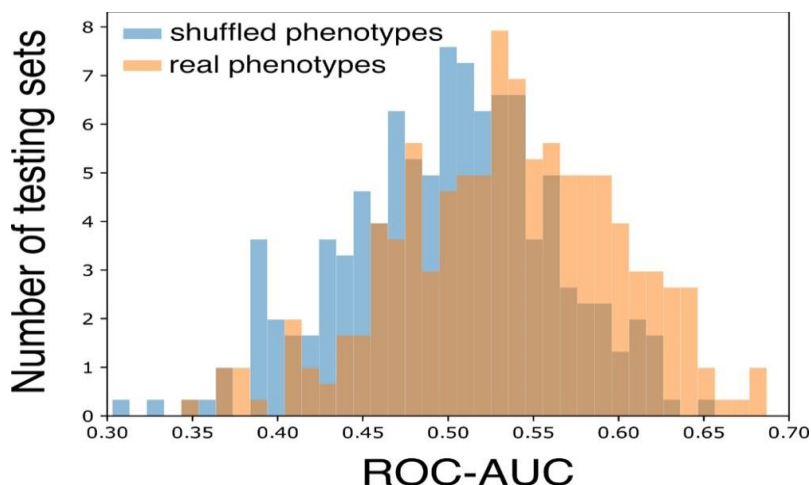


Figure 7: Model performances. The ROC-AUC is compared between models trained using randomly shuffled phenotypes (blue boxes) and the actual COVID-19 severity phenotypes (orange boxes). The two-sided t-test for the null hypothesis that 2 independent samples have equal average is performed with a statistically significant p-value of 2.7×10^{-6} .

4. Discussion

Using data analysis techniques to investigate approximately 20,000 genes, we have identified a subset of a few hundred candidate genes involved in the COVID-19 mild and severe disease. Some of those candidate genes appear to be involved in disease severity when associated with rare variants; while others of them appear to be involved though one or more common polymorphism/s.

Extracted genes bearing rare variants simulate a Mendelian-like disorder in infected members. Among these is the recently identified X-linked recessive disorder associated with the *TLR7* gene [6, 7, 13]. In this case males appear to be severely affected due to a defect in the viral sensor *TLR7*, which is unable to appropriately trigger the downstream events leading to interferon production. The appropriate therapy for these males is interferon as soon as the infection is identified. About ten families with this defect in the cohort of the pilot study were identified. Among females an autosomal dominant-like disorder conditioned by sex is due to *TLR5*, another sensor, previously involved in bacterial infection and more recently also in viral infection [17, 18]. This is an autosomal gene inhibited by estrogen. Those who inherit the mutation are at significant risk of severe COVID-19, especially if they are females in which the amount of the protein is naturally reduced. The recommended treatment, as with *TLR7*, is interferon. Some tens of families with this variant were present in the cohort of the pilot study. There are also inverted Mendelian disorders leading to inheritance of protective rare variants. These include: i) the *XPNPEP2* gene, the *ACE2* co-receptor that is also involved in the metabolism of bradykinin, a potent vasodilator that acts as a protective against vasoconstriction and thrombosis [19]. The gene is found on the X chromosome, thus having a role in protecting males; and ii) *VEGFD* gene, encoding for vascular endothelial growth factor D, predominantly expressed in the lungs, interacting with *ACE2*, with an important role in the pathogenesis of acute lung injury (ALI) and acute respiratory distress syndrome (ARDS) by its properties to increase vascular permeability [20,21]. Extracted genes bearing common variants delineate different groups of “pathogenicity”. For example, *TLR3* p.Leu412Phe (MAF 29,79%) confers reduced inflammatory response (reduced TNF α) and reduced autophagy [11]. The effect is enhanced in males because the *TLR3* is down regulated by

testosterone. In these patients hydroxychloroquine, which inhibits autophagy, is particularly dangerous. Most importantly, we defined a new genetic method for modelling COVID-19 severity (Figure 2). The proposed method has a number of innovative approaches. To arrive at this model based on this formula, first the clinically modified WHO outcome scale was combined with the subjects' age, providing the main "non genetic" factor influencing clinical outcome. Subjects were divided into three categories: those having a clinical outcome as expected solely according to age, those having either a worse than expected according to age, and those having a better clinical outcome than expected according to age. The last two were selected as extreme ends of phenotype and used for selecting relevant genes. Second, the method is gene based and processes the genes in a Boolean manner, i. e., having or not having variants. For both common and rare variants, the Boolean classification was repeated three times in order to correspond with subjects having at least the heterozygous genotype (dominant modes), having the homozygous genotype (recessive model), or having the hemizygous genotype (X-linked model) for genes on chromosome X in males and heterozygous genotype for gene on chromosome X (X-linked dominant model) in females.

Not rarely, genes have more than one coding polymorphism (3413 in the cohort). Various coding changes may move the functioning of a protein either in the same direction (for example lowering or increasing the function) or in the opposite direction, potentially cancelling the effect of juxtaposed genes. We used a "coding haplotype" approach for classifying polymorphic variability in a Boolean manner: we counted the most common combinations (not all theoretical combinations were present due to linkage disequilibrium) and assigned a value of 1 to each gene if the specific combination is present or 0, if the combination was absent. Again, the Boolean classification was repeated three times, corresponding to having at least the heterozygous genotype, having homozygous genotype, or having hemizygous (in males) or heterozygous (in females) genotype for genes on chromosome X. The method treats males and females separately. This represents a first approach to translating into practice sex medicine. As previous studies have borne out, male patients affected by COVID-19 undergo a more severe clinical course. Finally the method we used for gene selection, based on Lasso regularization for a logistic regression classifier, is indirectly confirmed by the association rules produced using a different method.

The proposed method accounts for the role of both common and rare variability, individually and combined. To our knowledge, this is the first method that has been able to synthesize a holistic approach across variabilities, which up until now have been treated separately in host genome research. By using the polygenic score derived from the GWAS approach for the common variability and Mendelian model for rare variability we arrived at a combined approach that significantly outstips the possibility of each of the earlier methodologies approached separately. The introduction of the IPGS formula proved determinative for a significant empowerment of host genome analysis. We call this method the "post-Mendelian model".

This new method now requires further investigation and refinement. For example, the method treats each gene as having the same weight among common as well as among rare variants. Furthermore, a number of genetic variabilities are missed in the model. These include i) non biallelic common polymorphisms, such as polyamino acid repeats, which are known to contribute to COVID-19 [10]; ii) variability due to germline CNV; iii) variability due to somatic mutations, both CNV and SNP [22, 23]; and iv) very rare variants or private variants whose frequency is too low to be detected by LASSO logistic regression. Further studies that take into account the above reported variability and that increase prediction performances are needed to improve the precision of the model.

5. Conclusion

In conclusion, we were able to identify a set of genes in which common polymorphisms confer either severity or mildness in relation to COVID-19 presentation and development. We also identified another set of genes in which rare variants confer either severity or mildness to COVID-19 disease. We have finally defined a new post-Mendelian genetic model, based on a combined analysis of common and rare variants that appears to explain severity in COVID-19 with a high degree of confidence.

The current version of the is robust enough to extract relevant genes from a specific patient and identify the main pathogenic pathways that are accessible to current personalized co-adjuvant treatment flanking the current use of cortisone. Even in its present stage of development, this post-Mendelian model is strongly capable of identifying candidate genes that can explain with confidence the presentation and development of COVID-19 in patients with mild and severe disease. The proposed model should, however, not be employed immediately as a predictive tool, as it requires more testing, preferably in independent cohorts of patients across various population sets. This model allows us the possibility to summarize and critically evaluate the enormous amount of information now available from sequencing experiments. The immediate link demonstrated here between the gene variants extracted by the automatic analyses and their possible biological roles in COVID-19 supports the idea that this approach to the analysis of genetic information can lead to useful conclusions for COVID-19 diseases management. It is also promising for the further development of this post-Mendelian model with reliable predictive capabilities.

By better understanding the role of host genetics in COVID-19 susceptibility and disease severity, we are also in a stronger position to identify public health measures that will may curb the impact of the disease on society as a whole. This should help us to genetically screen already affected patients as well as, eventually, individuals who may potentially be patients in order to predict those who are more or less susceptible to developing COVID-19 post-infection, especially the more severe cases. It should further help us in, not only reassigning therapeutics or developing new interventions (including vaccines), but also in decision-making regarding therapeutics and vaccine allocations. Beyond what this post-Mendelian method can help us understand regarding the role of host genetics in COVID-19 susceptibility and the potential implications for clinical and public health responses, the model also has strong potential for understanding the role of host genetics in other complex disorders. As we move into an era of precision, patient-centric medicine - an era being propelled by the lurking ubiquitous character of COVID-19 - this post-Mendelian method can help us tailor treatments to the specific needs of individual patients.

Acknowledgments

This study is part of the GEN-COVID Multicenter Study, <https://sites.google.com/dbm.unisi.it/gen-covid>, the Italian multicenter study aimed at identifying the COVID-19 host genetic bases. Specimens were provided by the COVID-19 Biobank of Siena, which is part of the Genetic Biobank of Siena, member of BBMRI-IT, of Telethon Network of Genetic Biobanks (project no. GTB18001), of EuroBioBank and of D-Connect. We thank the CINECA consortium for providing computational resources and the Network for Italian Genomes (NIG) <http://www.nig.cineca.it> for its support. We thank private donors for the support provided to A.R. (Department of Medical Biotechnologies, University of Siena) for the COVID-19 host genetics research project (D.L n.18 of March 17, 2020). We also thank the COVID-19 Host Genetics Initiative (<https://www.covid19hg.org/>). This work was funded by MIUR project “Dipartimenti di

Eccellenza 2018-2020” to Department of Medical Biotechnologies University of Siena, Italy (Italian D.L. n.18 March 17, 2020). Private donors for COVID-19 research. “Bando Ricerca COVID-19 Toscana” project to Azienda Ospedaliero-Universitaria Senese (CUP I49C20000280002). Charity fund 2020 from Intesa San Paolo dedicated to the project N. B/2020/0119 “Identificazione delle basi genetiche determinanti la variabilità clinica della risposta a COVID-19 nella popolazione italiana”. the Istituto Buddista Italiano Soka Gakkai for funding the project “PAT-COVID: Host genetics and pathogenetic mechanisms of COVID-19” (ID n. 2020-2016_RIC_3); the Italian Ministry of University and Research for funding within the “Bando FISR 2020” in COVID-19.

Conflicts of interest

All the authors declare no competing financial interests.

Ethics and consent to participate

The GEN-COVID study was consistent with Institutional guidelines and it was approved by the University Hospital of Siena Ethical Review Board (Protocol n. 16929, dated March 16, 2020). As part of GEN-COVID Multicenter Study written informed consent was obtained from all individuals who contributed samples and data. Detailed clinical and laboratory characteristics (data), specifically related to COVID-19, were collected for all subjects.

Consent for publication

The patients were informed of this research and agreed to it through the informed consent process.

Availability of data and materials

The data and samples referenced here are housed in the GEN-COVID Patient Registry and the GEN-COVID Biobank and are available for consultation. You may contact the corresponding author, Prof. Alessandra Renieri (e-mail: alessandra.renieri@unisi.it).

The datasets generated and analysed during the current study are available in the COVID-19 dedicated section (<http://nigdb.cineca.it>) repository, within the Network for Italian Genome (<http://www.nig.cineca.it>). There are no restrictions on data access. Only registration is needed.

Institutional review board statement

The GEN-COVID study was conducted according to the guidelines of the Declaration of Helsinki, and approved by the University Hospital of Siena Ethical Review Board (protocol code 16929, dated March 16, 2020).

Author contributions

Conceptualization, Antonio Esposito, Pietro Pinoli, Francesca Mari and Alessandra Renieri; Data curation, Giada Beligni and Mirjam Lista; Formal analysis, Nicola Picchiotti, Elisa Benetti, Chiara Fallerini, Sergio Daga, Margherita Baldassarri, Francesca Fava, Kristina Zguro, Stefano Ceri, Antonio Esposito, Pietro Pinoli, Francis P. Crawley, Elisa Frullanti, GEN-COVID Multicenter Study, Marco Gori, Alessandra Renieri and Simone Furini; Funding acquisition, Antonio Esposito, Pietro Pinoli and Francesca Mari; Methodology, Nicola Picchiotti, lisa Benetti, Floriana Valentino, Gabriella Doddato, Annarita Giliberti, Sara Amitrano, Mirella Bruttini and Simone Furini; Project administration, Antonio Esposito, Pietro Pinoli, Francesca Mari and Alessandra Renieri; Supervision, Antonio Esposito, Pietro Pinoli, Francesca Mari, GEN-COVID ulticenter Study and Alessandra Renieri; Validation, Simone Furini; Writing – original

draft, Nicola Picchiotti, Elisa Benetti, Chiara Fallerini, Sergio Daga, Margherita Baldassarri, Francesca Fava, Kristina Zguro, Floriana Valentino, Gabriella Doddato, Annarita Giliberti, Rossella Tita, Sara Amitrano, Mirella Bruttini, Laura Di Sarno, Diana Alaverdian, Giada Beligni, Maria Palmieri, Mirjam Lista, Susanna Croci, Ilaria Meloni, Anna Maria Pinto, Chiara Gabbi, Stefano Ceri, Antonio Esposito, Pietro Pinoli, Francis P. Crawley, Elisa Frullanti, Francesca Mari, GEN-COVID Multicenter Study, Marco Gori, Alessandra Renieri and Simone Furini.

References

1. Islam MR, Hoque MN, Rahman MS, et al. Genome-wide analysis of SARS-CoV-2 virus strains circulating worldwide implicates heterogeneity. *Sci Rep* (2020): 14004.
2. Severe Covid-19 GWAS Group, Ellinghaus D, Degenhardt F, et al. Genomewide Association Study of Severe Covid-19 with Respiratory Failure. *N Engl J Med* (2020): 1522-1534.
3. Pairo-Castineira E, Clohisey S, Klaric L, et al. Genetic mechanisms of critical illness in Covid-19. *Nature* 591 (2020): 92-98.
4. COVID-19 Host Genetics Initiative et al. Mapping the human genetic architecture of COVID-19. *Nature* (2021).
5. Zhang X, Tan Y, Ling Y, et al. Viral and host factors related to the clinical outcome of COVID-19. *Nature* 583 (2020): 437-440.
6. van der Made CI, Simons A, Schuurs-Hoeijmakers J, et al. Presence of Genetic Variants Among Young Men With Severe COVID-19. *324* (2020): 663-673.
7. Solanich X, Vargas-Parra G, van der Made CI, et al. Genetic screening for *TLR7* variants in young and previously healthy men with severe COVID-19: a case series. *Front Immunol* (2021): 719115.
8. Benetti E, Giliberti A, Emiliozzi A, et al. Clinical and molecular characterization of COVID-19 hospitalized patients. *PLoS One* 15 (2020): e0242534.
9. Daga S, Fallerini C, Baldassarri M, et al. Employing a systematic approach to biobanking and analyzing clinical and genetic data for advancing COVID-19 research. *Eur J Hum Genet* 29 (2021): 745-759.
10. Baldassarri M, Picchiotti N, Fava F, et al. Shorter androgen receptor polyQ alleles protect against life-threatening COVID-19 disease in European males. *EBioMedicine* (2021): 103246.
11. Croci S, Venneri MA, Mantovani S, et al. The polymorphism L412F in *TLR3* inhibits autophagy and is a marker of severe COVID-19 in males. *In press in Autophagy* 2021.
12. Fallerini C, Daga S, Benetti E, et al. SELP Asp603Asn and severe thrombosis in COVID-19 males: implication for anti P-selectin monoclonal antibodies treatment. *Journal of hematology & oncology* 14 (2021): 123.
13. Fallerini C, Daga S, Mantovani S, et al. Association of Toll-like receptor 7 variants with life-threatening COVID-19 disease in males: findings from a nested case-control study. *Elife* (2021): e67569.
14. Molnar C. Interpretable-machine-learning. A Guide for Making Black Box Models Explainable (2019).
15. Ranjith-Kumar CT, Miller W, Sun J, et al. Effects of single nucleotide polymorphisms on Toll-like receptor 3 activity and expression in cultured cells. *J Biol Chem* 282 (2007):17696-17705.
16. Dhangadamajhi G, Rout R. Association of *TLR3* functional variant (rs3775291) with COVID-19 susceptibility and death: a population-scale study. *Hum Cell* (2021): 1-3.
17. Caballero I, Boyd J, Almiñana C, et al. Understanding the dynamics of Toll-like Receptor 5 response to flagellin and its regulation by estradiol. *Sci Rep* (2017): 40981.

18. Georgel AF, Cayet D, Pizzorno A, et al. Toll-like receptor 5 agonist flagellin reduces influenza A virus replication independently of type I interferon and interleukin 22 and improves antiviral efficacy of oseltamivir. *Antiviral Res* 168 (2019): 28-35.
19. Kumar A. Network Proteins of Angiotensin-converting Enzyme 2 but Not Angiotensin-converting Enzyme 2 itself are Host Cell Receptors for SARS-Coronavirus-2 Attachment. *Biology, Engineering, Medicine and Science Reports* (2021): 01–05.
20. Kong Y, Han J, Wu X, et al. VEGF-D: a novel biomarker for detection of COVID-19 progression. *Crit Care* (2020): 373.
21. Shovlin CL, Vizcaychipi MP. Vascular inflammation and endothelial injury in SARS-CoV-2 infection: The overlooked regulatory cascades implicated by the ACE2 gene cluster. *QJM*. (2020): hcaa241.
22. Zekavat SM, Lin SH, Bick AG, et al. Hematopoietic mosaic chromosomal alterations and risk for infection among 767,891 individuals without blood cancer. *medRxiv* 3 (2020): 100817.
23. Bolton KL, Koh Y, Foote MB, et al. Clonal hematopoiesis is associated with risk of severe Covid-19. *medRxiv* (2020).



This article is an open access article distributed under the terms and conditions of the [Creative Commons Attribution \(CC-BY\) license 4.0](https://creativecommons.org/licenses/by/4.0/)

Supplementary figures and tables

Figure S1 Common polymorphisms and rare variants selected by LASSO Logistic Regression. Comparison of extreme ends of phenotype: subjects of class 1 (red in Fig.1) versus class 0 (green in Fig. 1) as defined by ordered logistic regression.

Common (>1%) bi-allelic polymorphisms (coding haplotypes) considering heterozygous variants plus homozygous variants versus wt genotype and using both males and females (**S1a**) or only males (**S1b**) or only females (**S1c**);

Common (>1%) bi-allelic polymorphisms (coding haplotypes) considering homozygous variants versus heterozygous variants plus wt genotype and using both males and females (**S1d**) or only males (**S1e**) or only in females (**S1f**)

Common (>1%) bi-allelic polymorphisms (coding haplotypes) considering hemizygous variants (only genes on chromosome X) versus wt genotype and using males only (**S1g**) or females only (**S1h**);

Rare variants (<1%) considering heterozygous variants plus homozygous variants versus wt genotype (**dominant model**) and using both males and females (**S1i**) or only males (**S1j**) or only females (**S1m**); Rare variants (<1%) considering homozygous variants versus heterozygous variants plus wt genotype (**recessive model**) and using both males and females (**S1n**) or only females (**S1o**).

For males (**S1p**) the performances are below the random guess; Rare variants (<1%) considering hemizygous variants (only genes on chromosome X) versus wt genotype (**X-linked model**) and using males only (**S1q**) or only females (**S1r**).

Features are gene-based representations of Genotypic Combinations (GC) of common polymorphisms. The histograms (weights) represented by importance of each feature (genes), including age and sex, for the classification task (**Upper Panel**).

The positive weights reflect a susceptible behaviour of the gene to the target COVID-19 disease, whereas the negative weights a mildness action. **Down Panel:** Cross-validation accuracy for the grid of LASSO regularization parameters; the error bar is given by the standard deviation of the average ROC-AUC within the 10 folds; the red point corresponds to the parameter chosen for the fitting procedure.

Supplementary Table 1. Common features extracted in males obtained after 82 bootstraps
Supplementary Table 2. Common features extracted in females obtained after 82 bootstraps
Supplementary Table 3. Rare features extracted in males obtained after 82 bootstraps
Supplementary Table 4. Rare features extracted in females obtained after 82 bootstraps

Supplementary Table 1. Common features extracted in males obtained after 82 bootstraps

Supplementary Table 2. Common features extracted in females obtained after 82 bootstraps

Supplementary Table 3. Rare features extracted in males obtained after 82 bootstraps

Supplementary Table 4. Rare features extracted in females obtained after 82 bootstraps

GEN-COVID Multicenter Study (<https://sites.google.com/dbm.unisi.it/gen-covid>)

Francesca Montagnani^{3,10}, Mario Tumbarello^{3,10}, Ilaria Rancan^{3,10}, Massimiliano Fabbiani¹⁰, Elena Bargagli¹¹, Laura Bergantini¹¹, Miriana D'Alessandro¹¹, Paolo Cameli¹¹, David Bennett¹¹, Federico Anedda¹², Simona Marcantonio¹², Sabino Scolletta¹², Federico Franchi¹², Maria Antonietta Mazzei¹³, Susanna Guerrini¹³, Edoardo Conticini¹⁴, Luca Cantarini¹⁴, Bruno Frediani¹⁴, Danilo Tacconi¹⁵, Chiara Spertilli Raffaelli¹⁵, Marco Feri¹⁶, Alice Donati¹⁶, Raffaele Scala¹⁷, Luca Guidelli¹⁷, Genni Spargi¹⁸, Marta Corridi¹⁸, Cesira Nencioni¹⁹, Leonardo Croci¹⁹, Gian Piero Caldarelli²⁰, Davide Romani²¹, Paolo Piacentini²¹, Maria Bandini²¹, Elena Desantis²¹, Silvia Cappelli²¹, Anna Canaccini²², Agnese Verzuri²², Valentina Anemoli²², Agostino Ognibene²³, Alessandro Pancrazzi²³, Maria Lorubbio²³, Massimo Vaghi²⁴, Antonella D'Arminio Monforte²⁵, Federica Gaia Miraglia²⁵, Mario U. Mondelli^{26,27}, Stefania Mantovani²⁶, Massimo Girardis²⁸, Sophie Venturelli²⁸, Stefano Busani²⁸, Andrea Cossarizza²⁹, Andrea Antinori³⁰, Alessandra Vergori³⁰, Arianna Emiliozzi³⁰, Stefano Rusconi^{31,32}, Matteo Siano³², Arianna Gabrieli³², Agostino Riva^{31,32}, Daniela Francisci^{33,34}, Elisabetta Schiaroli³³, Francesco Paciosi³³, Andrea Tommasi³³, Pier Giorgio Scotton³⁵, Francesca Andretta³⁵, Sandro

Panese³⁶, Stefano Baratti³⁶, Renzo Scaggiante³⁷, Francesca Gatti³⁷, Saverio Giuseppe Parisi³⁸, Francesco Castelli³⁹, Eugenia Quiros-Roldan³⁹, Melania Degli Antoni³⁹, Isabella Zanella^{40,41}, Matteo Della Monica⁴², Carmelo Piscopo⁴², Mario Capasso^{43,44,45}, Roberta Russo^{43,44}, Immacolata Andolfo^{43,44}, Achille Iolascon^{43,44}, Giuseppe Fiorentino⁴⁶, Massimo Carella⁴⁷, Marco Castori⁴⁷, Filippo Aucella⁴⁸, Pamela Raggi⁴⁹, Carmen Marciano⁴⁹, Rita Perna⁴⁹, Matteo Bassetti^{50,51}, Antonio Di Biagio^{50,51}, Maurizio Sanguinetti^{52,53}, Luca Masucci^{52,53}, Alessandra Guarnaccia⁵², Serafina Valente⁵⁴, Oreste De Vivo⁵⁴, Maria Antonietta Mencarelli⁵, Caterina Lo Rizzo⁵, Francesca Ariani^{3,4,5}, Marco Mandalà⁵⁵, Alessia Giorli⁵⁵, Lorenzo Salerni⁵⁵, Patrizia Zucchi⁵⁶, Pierpaolo Parravicini⁵⁶, Elisabetta Menatti⁵⁷, Tullio Trotta⁵⁸, Ferdinando Giannattasio⁵⁸, Gabriella Coiro⁵⁸, Fabio Lena⁵⁹, Leonardo Gianluca Lacerenza⁵⁹, Cristina Mussini⁶⁰, Enrico Martinelli⁶¹, Luisa Tavecchia⁶², Mary Ann Belli⁶², Lia Crotti^{63,64,65,66,67}, Gianfranco Parati^{63,64}, Maurizio Sanarico⁶⁸, Francesco Raimondi⁶⁹, Filippo Biscarini⁷⁰, Alessandra Stella⁷⁰, Tiziana Bachetti⁷¹, Maria Teresa La Rovere⁷², Maurizio Bussotti⁷³, Serena Ludovisi⁷⁴, Katia Capitani^{3,75}, Simona Dei⁷⁶, Sabrina Ravaglia⁷⁷, Rosangela Artuso⁷⁸, Elena Andreucci⁷⁸, Giulia Gori⁷⁸, Angelica Pagliazzi⁷⁸, Erika Fiorentin⁷⁸, Antonio Perrella⁷⁹, Francesco Bianchi^{79,3}, Paola Bergomi⁸⁰, Emanuele Catena⁸⁰, Riccardo Colombo⁸⁰, Sauro Luchi⁸¹, Giovanna Morelli⁸¹, Paola Petrocelli⁸¹, Sarah Iacopini⁸¹, Sara Modica⁸¹, Silvia Baroni⁸², Francesco Vladimiro Segala⁸³, Marco Tanfoni¹, Marco Falcone⁸⁴, Giusy Tiseo⁸⁴, Chiara Barbieri⁸⁴, Tommaso Matucci⁸⁴, Davide Grassi⁸⁵, Claudio Ferri⁸⁵, Franco Marinangeli⁸⁶, Francesco Brancati⁸⁷, Antonella Vincenti⁸⁸, Valentina Borgo⁸⁸, Stefania Lombardi⁸⁸, Mirco Lenzi⁸⁸, Massimo Antonio Di Pietro⁸⁹, Francesca Vichi⁸⁹, Benedetta Romanin⁸⁹, Letizia Attala⁸⁹, Cecilia Costa⁸⁹, Andrea Gabbuti⁸⁹, Roberto Menè^{63,64}, Patrizia Casprini⁹⁰, Giuseppe Merla^{91,92}, Gabriella Maria Squeo⁹¹, Marcello Maffezzoni⁹³, Raffaele Bruno^{94,95}, Marco Vecchia⁹⁴, Marta Colaneri⁹⁴

10. Department of Medical Sciences, Infectious and Tropical Diseases Unit, Azienda Ospedaliera Universitaria Senese, Siena, Italy
11. Unit of Respiratory Diseases and Lung Transplantation, Department of Internal and Specialist Medicine, University of Siena, Italy
12. Dept of Emergency and Urgency, Medicine, Surgery and Neurosciences, Unit of Intensive Care Medicine, Siena University Hospital, Italy
13. Department of Medical, Surgical and Neuro Sciences and Radiological Sciences, Unit of Diagnostic Imaging, University of Siena, Italy
14. Rheumatology Unit, Department of Medicine, Surgery and Neurosciences, University of Siena, Policlinico Le Scotte, Italy
15. Department of Specialized and Internal Medicine, Infectious Diseases Unit, San Donato Hospital Arezzo, Italy
16. Dept of Emergency, Anesthesia Unit, San Donato Hospital, Arezzo, Italy
17. Department of Specialized and Internal Medicine, Pneumology Unit and UTIP, San Donato Hospital, Arezzo, Italy
18. Department of Emergency, Anesthesia Unit, Misericordia Hospital, Grosseto, Italy
19. Department of Specialized and Internal Medicine, Infectious Diseases Unit, Misericordia Hospital, Grosseto, Italy
20. Clinical Chemical Analysis Laboratory, Misericordia Hospital, Grosseto, Italy
21. Department of Preventive Medicine, Azienda USL Toscana Sud Est, Italy
22. Territorial Scientific Technician Department, Azienda USL Toscana Sud Est, Italy

23. Clinical Chemical Analysis Laboratory, San Donato Hospital, Arezzo, Italy
24. Chirurgia Vascolare, Ospedale Maggiore di Crema, Italy
25. Department of Health Sciences, Clinic of Infectious Diseases, ASST Santi Paolo e Carlo, University of Milan, Italy
26. Division of Infectious Diseases and Immunology, Fondazione IRCCS Policlinico San Matteo, Pavia, Italy
27. Department of Internal Medicine and Therapeutics, University of Pavia, Italy
28. Department of Anesthesia and Intensive Care, University of Modena and Reggio Emilia, Modena, Italy
29. Department of Medical and Surgical Sciences for Children and Adults, University of Modena and Reggio Emilia, Modena, Italy
30. HIV/AIDS Department, National Institute for Infectious Diseases, IRCCS, Lazzaro Spallanzani, Rome, Italy
31. III Infectious Diseases Unit, ASST-FBF-Sacco, Milan, Italy
32. Department of Biomedical and Clinical Sciences Luigi Sacco, University of Milan, Milan, Italy
33. Infectious Diseases Clinic, Department of Medicine 2, Azienda Ospedaliera di Perugia and University of Perugia, Santa Maria Hospital, Perugia, Italy
34. Infectious Diseases Clinic, "Santa Maria" Hospital, University of Perugia, Perugia, Italy
35. Department of Infectious Diseases, Treviso Hospital, Local Health Unit 2 Marca Trevigiana, Treviso, Italy
36. Clinical Infectious Diseases, Mestre Hospital, Venezia, Italy.
37. Infectious Diseases Clinic, ULSS1, Belluno, Italy
38. Department of Molecular Medicine, University of Padova, Italy
39. Department of Infectious and Tropical Diseases, University of Brescia and ASST Spedali Civili Hospital, Brescia, Italy
40. Department of Molecular and Translational Medicine, University of Brescia, Italy;
41. Clinical Chemistry Laboratory, Cytogenetics and Molecular Genetics Section, Diagnostic Department, ASST Spedali Civili di Brescia, Italy
42. Medical Genetics and Laboratory of Medical Genetics Unit, A.O.R.N. "Antonio Cardarelli", Naples, Italy
43. Department of Molecular Medicine and Medical Biotechnology, University of Naples Federico II, Naples, Italy
44. CEINGE Biotecnologie Avanzate, Naples, Italy
45. IRCCS SDN, Naples, Italy
46. Unit of Respiratory Physiopathology, AORN dei Colli, Monaldi Hospital, Naples, Italy
47. Division of Medical Genetics, Fondazione IRCCS Casa Sollievo della Sofferenza Hospital, San Giovanni Rotondo, Italy
48. Department of Medical Sciences, Fondazione IRCCS Casa Sollievo della Sofferenza Hospital, San Giovanni Rotondo, Italy
49. Clinical Trial Office, Fondazione IRCCS Casa Sollievo della Sofferenza Hospital, San Giovanni Rotondo, Italy
50. Department of Health Sciences, University of Genova, Genova, Italy
51. Infectious Diseases Clinic, Policlinico San Martino Hospital, IRCCS for Cancer Research Genova, Italy
52. Microbiology, Fondazione Policlinico Universitario Agostino Gemelli IRCCS, Catholic University of Medicine, Rome, Italy
53. Department of Laboratory Sciences and Infectious Diseases, Fondazione Policlinico Universitario A. Gemelli

- IRCCS, Rome, Italy
54. Department of Cardiovascular Diseases, University of Siena, Siena, Italy
 55. Otolaryngology Unit, University of Siena, Italy
 56. Department of Internal Medicine, ASST Valtellina e Alto Lario, Sondrio, Italy
 57. Study Coordinator Oncologia Medica e Ufficio Flussi Sondrio, Italy
 58. First Aid Department, Luigi Curto Hospital, Polla, Salerno, Italy
 59. Department of Pharmaceutical Medicine, Misericordia Hospital, Grosseto, Italy
 60. Infectious Diseases Clinics, University of Modena and Reggio Emilia, Modena, Italy
 61. Department of Respiratory Diseases, Azienda Ospedaliera di Cremona, Cremona, Italy
 62. U.O.C. Medicina, ASST Nord Milano, Ospedale Bassini, Cinisello Balsamo (MI), Italy
 63. Istituto Auxologico Italiano, IRCCS, Department of Cardiovascular, Neural and Metabolic Sciences, San Luca Hospital, Milan, Italy
 64. Department of Medicine and Surgery, University of Milano-Bicocca, Milan, Italy
 65. Istituto Auxologico Italiano, IRCCS, Center for Cardiac Arrhythmias of Genetic Origin, Milan, Italy
 66. Istituto Auxologico Italiano, IRCCS, Laboratory of Cardiovascular Genetics, Milan, Italy
 67. Member of the European Reference Network for Rare, Low Prevalence and Complex Diseases of the Heart-ERN GUARD-Heart
 68. Independent Data Scientist, Milan, Italy
 69. Scuola Normale Superiore, Pisa, Italy
 70. CNR-Consiglio Nazionale delle Ricerche, Istituto di Biologia e Biotecnologia Agraria (IBBA), Milano, Italy
 71. Direzione Scientifica, Istituti Clinici Scientifici Maugeri IRCCS, Pavia, Italy
 72. Istituti Clinici Scientifici Maugeri IRCCS, Department of Cardiology, Institute of Montescano, Pavia, Italy
 73. Istituti Clinici Scientifici Maugeri IRCCS, Department of Cardiology, Institute of Milan, Milan, Italy
 74. Fondazione IRCCS Ca' Granda Ospedale Maggiore Policlinico, Milan, Italy
 75. Core Research Laboratory, ISPRO, Florence, Italy
 76. Health Management, Azienda USL Toscana Sudest, Tuscany, Italy
 77. IRCCS C. Mondino Foundation, Pavia, Italy
 78. Medical Genetics Unit, Meyer Children's University Hospital, Florence, Italy
 79. Department of Medicine, Pneumology Unit, Misericordia Hospital, Grosseto, Italy.
 80. Department of Anesthesia and Intensive Care Unit, ASST Fatebenefratelli Sacco, Luigi Sacco Hospital, Polo Universitario, University of Milan, Milan
 81. Infectious Disease Unit, Hospital of Lucca, Italy
 82. Department of Diagnostic and Laboratory Medicine, Institute of Biochemistry and Clinical Biochemistry, Fondazione Policlinico Universitario A. Gemelli IRCCS, Catholic University of the Sacred Heart, Rome, Italy.
 83. Clinic of Infectious Diseases, Catholic University of the Sacred Heart, Rome, Italy
 84. Department of Clinical and Experimental Medicine, Infectious Diseases Unit, University of Pisa, Pisa, Italy
 85. Department of Clinical Medicine, Public Health, Life and Environment Sciences, University of L'Aquila, Italy
 86. Anesthesiology and Intensive Care, University of L'Aquila, L'Aquila, Italy
 87. Medical Genetics Unit, Department of Life, Health and Environmental Sciences, University of L'Aquila, L'Aquila, Italy
 88. Infectious Disease Unit, Hospital of Massa, Italy

89. Unit of Infectious Diseases, S.M. Annunziata Hospital, Florence, Italy..
90. Laboratory of Clinical Pathology and Immunoallergy, Florence-Prato, Italy
91. Laboratory of Regulatory and Functional Genomics, Fondazione IRCCS Casa Sollievo della Sofferenza, San Giovanni Rotondo (Foggia), Italy
92. Department of Molecular Medicine and Medical Biotechnology, University of Naples Federico II, Naples, Italy.
93. University of Pavia, Pavia, Italy
94. Division of Infectious Diseases I, Fondazione IRCCS Policlinico San Matteo, Pavia, Italy
95. Department of Clinical, Surgical, Diagnostic, and Pediatric Sciences, University of Pavia, Pavia, Italy