













**Table 4:** Pairwise comparisons of area under receiver operating characteristic curves of the imaging methods for detecting various hepatic steatosis grades.

ATI vs. PDFF	S0 vs. S1-S3 (S>0)	S0-S1 vs. S2-S3 (S>1)
Difference between areas	0.0982	0.04
Standard Error <sup>a</sup>	0.0932	0.0422
95% Confidence Interval	-0.0845 to 0.281	-0.0428 to 0.123
z statistic	1.054	0.947
Significance level	p=0.292	p = 0.344
ATI vs. MRS		
Difference between areas	0.0982	0.08
Standard Error <sup>a</sup>	0.0932	0.0951
95% Confidence Interval	-0.0845 to 0.281	-0.106 to 0.266
z statistic	1.054	0.841
Significance level	p=0.292	p = 0.400
PDFF vs. MRS		
Difference between areas	0	0.12
Standard Error <sup>a</sup>	0	0.0938
95% Confidence Interval	-	-0.0639 to 0.304
z statistic	-	1.279
Significance level	p=1.000	p = 0.201

<sup>a</sup>DeLong test. ATI= attenuation imaging, MRS= magnetic resonance spectroscopy, PDFF= proton density fat fraction, S=steatosis grade.

grade 1 (S0-1 vs. S2-3), and 0.93 (0.69-1.00, p<0.001, for grade 2 (S0-2 vs. S3). The corresponding Youden’s index, sensitivity, specificity as well as the AUROCs of MRI-PDFF and proton-MRS are shown in Table 3.

There were no significant differences on pairwise comparison of the AUROCs of the imaging methods (ATI, MRI-PDFF and proton-MRS) for detecting S> grade 0 and S> grade 1, as summarised in Table 4. Due to limited sample size, data were insufficient for a pairwise comparison of the AUROCs for detecting S> grade 2.

## Discussion

In this study, ATI showed a significant correlation with MRI-PDFF, and proton-MRS. ATI also showed a significant correlation with liver steatosis grade as determined by histology, but not with NAS activity score and fibrosis stage. The diagnostic performance of ATI for detecting liver steatosis greater than grades 0,1 and 2 were 0.91, 0.97 and 0.93, respectively. The diagnostic performance of ATI, MRI-PDFF and proton-MRS were shown not to be significantly different in detecting various liver steatosis grades. The strength of this study lies in the use of subjects who had liver biopsy and within a short interval between biopsy and imaging. The success of ATI in this morbid obese cohort with liver steatosis without any other underlying liver pathologies

is another strength. The comparison of ATI with both MRI-PDFF and proton-MRS which has not been assessed in previous studies further adds to the strength of the study.

ATI showed a strong correlation with steatosis grade (r=0.833). Bae et al [25]. in a study involving 108 patients with histology proven steatosis showed the correlation coefficient of ATI with steatosis grade of 0.660. Likewise, Tamaki et al [26] in a study involving 351 patients showed a correlation coefficient of 0.470. Similarly, Huang et al [27] showed a correlation of 0.721 between ATI and steatosis grade. Variations in the correlation coefficients in these studies could be attributed to the varying BMIs, sample sizes, mixed liver aetiologies, and ethnicities used. Moreover, Nazare et al [28] showed that ethnicity significantly affects liver fat distribution, while Fan et al [29] showed that BMI (even in dose dependent manner) was associated with fatty liver risk. Indeed, our study cohort had no known other liver pathologies as was in other studies. It was further shown that ATI did not correlate with NAS activity score and fibrosis stage, in agreement with previous studies [25,26]. These outcomes may imply that ATI may not be affected by inflammation/fibrosis and therefore a poor marker for both conditions, yet it reassures that ATI seems to only determine the fat content in the liver without being affected by the presence of inflammation/fibrosis. However, more studies linking ATI with necroinflammatory activity score, or fibrosis stage are needed to confirm this outcome.

In this study, the diagnostic accuracy (AUROC) of ATI in detecting liver steatosis grades 1, 2 and 3 were 0.91, 0.97 and 0.93 respectively. The associated AC cut-off points for the above steatosis grades were: 0.59cm/dB/MHz, 0.60cm/dB/MHz, and 0.63cm/dB/MHz, respectively. These outcomes are similar to previous studies [16, 25, 26, 30-37]. It was also shown that pairwise comparison of AUROCs of the imaging methods did not show any significant differences. These results suggest that ATI may be a useful non-invasive tool to quantify liver steatosis. They further imply that ATI may be a reliable method to quantify hepatic fat even in subjects with morbid obesity, and its usefulness may enable early detection of liver steatosis at a relatively low cost. This could allow “mass screening” to be possible for purposes of either general population studies on NAFLD or for early detection of liver steatosis. This might particularly be helpful in encouraging lifestyle changes before marked hepatic damage occurs. However, the AC range of steatosis grade 0 was shown to overlap with the AC range of steatosis grade 1, unlike the outcome of both MRI-PDFF and proton-MRS. Like our findings, we also noticed this overlap in previous studies [16,25,26,30-37]. These outcomes suggest that ATI may not be as sensitive as MRI-PDFF/proton-MRS for distinguishing between steatosis grades 0 and 1, and care must be taken when using ATI as some cases may be allocated as being

positive or negative when in fact not. Under such suspicious circumstances, MRI-PDFF can be used to confirm the outcome. Notwithstanding the above argument, ATI seems to be reasonably more sensitive in detecting severe forms of steatosis (grades 2 and 3).

Our study, together with previous studies showed a variation in AC cut-off points for various steatosis grades (i.e., 0.56- 0.63cm/dB/MHz for steatosis > 0; 0.59-0.72cm/dB/MHz for steatosis > 1; and 0.69-0.94cm/dB/MHz for steatosis > 2) [16, 26, 30, 32, 34, 38-40]. This variation in AC cut-off points in these studies could in part be explained by use of transducers of varying frequencies (mostly 3 and 4MHz as reference frequencies), especially that attenuation is directly related to transducer frequency (the higher the frequency, the higher the attenuation). However, this variation further suggests that presently, ATI has no standardised cut-off points as is the case with MRI, hence the need for more validation studies involving large sample sizes, in multiple centres, and using a homogenous transducer reference frequency.

The major limitation of this study is the small sample size, which could account for our findings, and this was because of COVID-19 pandemic as all elective surgical cases were suspended. However, the results obtained in this study are comparable to few available studies on ATI which had larger sample sizes, thus, care must be exercised when interpreting these results. ATI examinations were conducted by a single operator, thus, interobserver agreement could not be assessed. Nonetheless, inter-observer correlation of ATI has been shown to range from 0.91-0.98 [13, 33, 41-44]. Finally, the cohort was of Chinese ethnicity, therefore, caution should be taken in the generalisation of the results.

## Conclusion

ATI is a promising non-invasive tool in quantifying hepatic fat, whose diagnostic performance is not significantly different to MRI-PDFF and proton-MRS. Future multicentre studies with large sample sizes in subjects with histologically proven liver steatosis and a control group may be needed to further validate this modality, especially that currently there is non-availability of standardized attenuation coefficient cut-off values for each corresponding steatosis grade in various ethnic groups and BMIs.

## Author Contributions

The roles of CC in the study were: conceptualization, data curation, formal analysis, investigation, methodology. WC roles were in: methodology, project administration, resources, supervision, writing, review & editing. KHL roles were in: investigation, supervision, writing, review & editing. VW roles were in: validation, writing, review & editing, supervision. CC and WC were involved in investigation and

analysis. WC sourced the project funding. All authors agreed to be accountable for all aspects of the work.

## Acknowledgements

The authors are grateful to the staff at the Chinese University of Hong Kong- Prince of Wales Hospital (CUHK-PWH) Multidisciplinary Clinic of Metabolic & Bariatric Surgery (MCMBS) and Radiology department for their contributions. The authors also thank all the participants in the study.

**Conflicts of interest:** None to report

## References

1. Szczepaniak LS, Nurenberg P, Leonard D, et al. Magnetic resonance spectroscopy to measure hepatic triglyceride content: Prevalence of hepatic steatosis in the general population. *American Journal of Physiology-Endocrinology and Metabolism* 288 (2005): 462-468.
2. Estes C, Anstee QM, Arias-Loste MT, et al. Modeling nafld disease burden in china, france, germany, italy, japan, spain, united kingdom, and united states for the period 2016–2030. *J Hepatol* 69 (2018): 896-904.
3. Kumar R, Priyadarshi RN, Anand U. Non-alcoholic fatty liver disease: Growing burden, adverse outcomes and associations. *Journal of Clinical and Translational Hepatology* 8 (2020): 76.
4. Li J, Zou B, Yeo YH, et al. Prevalence, incidence, and outcome of non-alcoholic fatty liver disease in asia, 1999–2019: A systematic review and meta-analysis. *The lancet Gastroenterology & hepatology* 4 (2019): 389-398.
5. Younossi ZM, Koenig AB, Abdelatif D, et al. Global epidemiology of nonalcoholic fatty liver disease—meta-analytic assessment of prevalence, incidence, and outcomes. *Hepatology* 64 (2016): 73-84.
6. Chalasani N, Younossi Z, Lavine JE, et al. The diagnosis and management of nonalcoholic fatty liver disease: Practice guidance from the american association for the study of liver diseases. *Hepatology* 67 (2018): 328-357.
7. European Association for the Study of the Liver, European Association for the Study of Diabetes, (EASD). EASL-EASD-EASO clinical practice guidelines for the management of non-alcoholic fatty liver disease. *Obesity facts* 9 (2016): 65-90.
8. Angulo P. Nonalcoholic fatty liver disease. *N Engl J Med* 346 (2002): 1221-1231.
9. Bedossa P, Dargère D, Paradis V. Sampling variability of liver fibrosis in chronic hepatitis C. *Hepatology* 38 (2003): 1449-1457.



10. French METAVIR Cooperative Study Group, Bedossa P. Intraobserver and interobserver variations in liver biopsy interpretation in patients with chronic hepatitis C. *Hepatology* 20 (1994): 15-20.
11. Scatarige JC, Scott WW, Donovan PJ, et al. Fatty infiltration of the liver: Ultrasonographic and computed tomographic correlation. *Journal of Ultrasound in Medicine* 3 (1984): 9-14.
12. Pirmoazen AM, Khurana A, El Kaffas A, et al. Quantitative ultrasound approaches for diagnosis and monitoring hepatic steatosis in nonalcoholic fatty liver disease. *Theranostics* 10 (2020): 4277.
13. Tada T, Iijima H, Kobayashi N, et al. Usefulness of attenuation imaging with an ultrasound scanner for the evaluation of hepatic steatosis. *Ultrasound Med Biol* 45 (2019): 2679-2687.
14. Zhang YN, Fowler KJ, Hamilton G, et al. Liver fat imaging—a clinical overview of ultrasound, CT, and MR imaging. *Br J Radiol* 91 (2018):20170959.
15. Wong VW, Adams LA, de Lédizinghen V, et al. Noninvasive biomarkers in NAFLD and NASH—current progress and future promise. *Nature reviews Gastroenterology & hepatology* 15 (2018): 461-478.
16. Tada T, Iijima H, Kobayashi N, et al. Usefulness of attenuation imaging with an ultrasound scanner for the evaluation of hepatic steatosis. *Ultrasound Med Biol* 45 (2019): 2679-2687.
17. Fan J, Kim S, Wong VW. New trends on obesity and NAFLD in asia. *J Hepatol* 67 (2017): 862-873.
18. Consultation WE. Appropriate body-mass index for asian populations and its implications for policy and intervention strategies. *Lancet (London, England)* 363 (2004): 157-163.
19. Matthews DR, Hosker JP, Rudenski AS, et al. Homeostasis model assessment: Insulin resistance and  $\beta$ -cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia* 28 (1985): 412-419.
20. Lee CH, Shih A, Woo YC, et al. Optimal cut-offs of homeostasis model assessment of insulin resistance (HOMA-IR) to identify dysglycemia and type 2 diabetes mellitus: A 15-year prospective study in chinese. *PloS one* 11 (2016): 0163424.
21. World Health Organization. Definition and diagnosis of diabetes mellitus and intermediate hyperglycaemia: Report of a WHO/IDF consultation (2006).
22. Alberti K, Eckel RH, Grundy SM, et al. Harmonizing the metabolic syndrome: A joint interim statement of the international diabetes federation task force on epidemiology and prevention; national heart, lung, and blood institute; american heart association; world heart federation; international atherosclerosis society; and international association for the study of obesity. *Circulation* 120 (2009): 1640-1645.
23. Kleiner DE, Brunt EM, Van Natta M, et al. Design and validation of a histological scoring system for nonalcoholic fatty liver disease. *Hepatology* 41 (2005): 1313-1321.
24. Stefan D, Di Cesare F, Andrasescu A, et al. Quantitation of magnetic resonance spectroscopy signals: The jMRUI software package. *Measurement Science and Technology* 20 (2009): 104035.
25. Bae JS, Lee DH, Lee JY, et al. Assessment of hepatic steatosis by using attenuation imaging: A quantitative, easy-to-perform ultrasound technique. *Eur Radiol* 29 (2019): 6499-6507.
26. Tamaki N, Koizumi Y, Hirooka M, et al. Novel quantitative assessment system of liver steatosis using a newly developed attenuation measurement method. *Hepatology Research* 48 (2018): 821-828.
27. Huang Y, Bian H, Zhu Y, et al. Quantitative diagnosis of nonalcoholic fatty liver disease with ultrasound attenuation imaging in a biopsy-proven cohort. *Acad Radiol* (2023).
28. Nazare J, Smith JD, Borel A, et al. Ethnic influences on the relations between abdominal subcutaneous and visceral adiposity, liver fat, and cardiometabolic risk profile: The international study of prediction of intra-abdominal adiposity and its relationship with cardiometabolic risk/ intra-abdominal adiposity. *Am J Clin Nutr* 96 (2012): 714-726.
29. Fan R, Wang J, Du J. Association between body mass index and fatty liver risk: A dose-response analysis. *Scientific reports* 8 (2018): 1-7.
30. Ferraioli G, Maiocchi L, Raciti MV, et al. Detection of liver steatosis with a novel ultrasound-based technique: A pilot study using MRI-derived proton density fat fraction as the gold standard. *Clinical and translational gastroenterology* 10 (2019): 00081
31. Hsu P, Wu L, Yen H, et al. Attenuation imaging with ultrasound as a novel evaluation method for liver steatosis. *Journal of clinical medicine* 10 (2021): 965.
32. Fujiwara Y, Kuroda H, Abe T, et al. The B-mode image-guided ultrasound attenuation parameter accurately detects hepatic steatosis in chronic liver disease. *Ultrasound Med Biol* 44 (2018): 2223-2232.

33. Yoo J, Lee JM, Joo I, et al. Reproducibility of ultrasound attenuation imaging for the noninvasive evaluation of hepatic steatosis. *Ultrasonography* 39 (2020): 121.
34. Jeon SK, Lee JM, Joo I, et al. Prospective evaluation of hepatic steatosis using ultrasound attenuation imaging in patients with chronic liver disease with magnetic resonance imaging proton density fat fraction as the reference standard. *Ultrasound Med Biol* 45 (2019): 1407-1416.
35. Ferraioli G, Monteiro LBS. Ultrasound-based techniques for the diagnosis of liver steatosis. *World journal of gastroenterology* 25 (2019): 6053.
36. Ferraioli G, Maiocchi L, Savietto G, et al. Performance of the attenuation imaging technology in the detection of liver steatosis. *Journal of Ultrasound in Medicine* 40 (2021): 1325-1332.
37. Welman CJ, Saunders J, Zelesco M, Abbott S, Boardman G, Ayonrinde OT. Hepatic steatosis: Ultrasound assessment using attenuation imaging (ATI) with liver biopsy correlation. *Journal of Medical Imaging and Radiation Oncology* 67 (2023): 45-53.
38. Hsu P, Wu L, Yen H, et al. Attenuation imaging with ultrasound as a novel evaluation method for liver steatosis. *Journal of clinical medicine* 10 (2021): 965.
39. Yoo J, Lee JM, Joo I, et al. Reproducibility of ultrasound attenuation imaging for the noninvasive evaluation of hepatic steatosis. *Ultrasonography* 39 (2020): 121.
40. Bae JS, Lee JM, Park S, Lee KB, Han JK. Magnetic resonance elastography of healthy livers at 3.0 T: Normal liver stiffness measured by SE-EPI and GRE. *Eur J Radiol* 107 (2018): 46-53.
41. Bae JS, Lee DH, Lee JY, et al. Assessment of hepatic steatosis by using attenuation imaging: A quantitative, easy-to-perform ultrasound technique. *Eur Radiol* 29 (2019): 6499-6507.
42. Ferraioli G, Soares Monteiro LB. Ultrasound-based techniques for the diagnosis of liver steatosis. *World J Gastroenterol* 25 (2019): 6053-6062.
43. Jeon SK, Lee JM, Joo I, et al. Prospective evaluation of hepatic steatosis using ultrasound attenuation imaging in patients with chronic liver disease with magnetic resonance imaging proton density fat fraction as the reference standard. *Ultrasound Med Biol* 45 (2019): 1407-1416.
44. Tamaki N, Koizumi Y, Hirooka M, et al. Novel quantitative assessment system of liver steatosis using a newly developed attenuation measurement method. *Hepatology Research* 48 (2018): 821-828.