

Research Article

Development and Physicochemical Characterization of a Functional Mozzarella Cheese Added with Agavin

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Abstract

In recent years worldwide, the manufacturing and consumption of functional foods have augmented importantly. Dairy products are nutritive and highly consumed while functional foods improve consumer health. A Mozzarella cheese added with agavin, was formulated, prepared, characterized and studied in order to know its properties, influenced by three variables: concentration of agavin as a prebiotic component, pre-acidification as part of the manufacturing process and storage time. The effect of each variable was different, the incorporation of agavin (0.7-3.3%) improved the nutritional value (fiber) and properties such as luminosity, texture and elasticity. The pre-acidification (0.0375%) stage augmented the acidity and improved some functional and physical properties. Both variables did not influence acceptance by the consumer, thus the assessed cheese samples had good sensory scores. Whereas the storage time (1, 11, 21 and 31 days) in which the properties were analyzed, recorded a decreasing trend in moisture content and pH, with low net color changes, and variable changes in texture.

Keywords: Mozzarella cheese; Functional dairy food; Physicochemical properties; Prebiotic; Agavin

1. Introduction

In the world's context, cheese consumption has increased over the years, observing high demand for two groups, fresh and pasta filata cheeses, depending on the socioeconomic conditions and customs of each region. Mozzarella is part of the pasta filata cheeses, it was originally made from buffalo milk, but now it is produced from cow milk in many countries. The properties of this dairy product are mainly consequence of its chemical composition and

structure of the para-casein matrix obtained through the manufacturing process. This cheese has a semisolid and plastic-curd consistency, and through heating, it develops its meltability and stretchability [1-7]. Its industrial production and consumption have augmented through the years.

Both, texture and functional properties of Mozzarella cheese result from its formulation and manufacturing process, therefore the cheese composition and process conditions should be controlled in order to obtain a dairy product with desirable characteristics [8-13]. The meltability as the capacity to melt on heating is directly influenced by factors such as calcium, fat content, moisture content, pH and nitrogen fractions [13, 14] and the stretchability as the ability to elongate and form strings under the effect of the applied forces and temperatures, is also influenced by several factors from the raw milk to post-manufacturing conditions, being the curdling stage used to obtain the linearity of protein chains in its structure [4]. Further, the direct pre-acidification of milk previous to the coagulation stage, with organic acids to substitute the slow acidification by thermophilic microorganisms, is one of the practical variations through the Mozzarella cheese manufacturing, accelerating the process time and offering advantages in the cheddaring stage [1, 7, 14, 15, 16].

On the other part, prebiotics as functional ingredients have proven their beneficial effect in humans, such as a better immune response, possible protection against cancer and the reduction of the cardiovascular diseases. These components selectively stimulate the growth and activity of a limited number of bacteria at the colon; particularly the non-digestible oligosaccharides, that are the most studied and recognized substrate [13, 17-22]. Inulin is a GRAS compound, it has a neutral soft flavor and is a soluble prebiotic polymer of easy incorporation into aqueous systems, like beverages and dairy products. It provides dietary fiber, prebiotic benefits [13, 23] and textural attributes in dairy products [24-26]. Agavin is a variety of inulin, representing a potential source of fructo-oligosaccharides (FOS) with a β -configuration at the second anomeric carbon, which produces the resistance to hydrolysis from human enzymes [27], also FOS take effect on lipid metabolism and reduce risk of colon cancer. Agavin was recognized by Roberfroid and Slavin [28] with a similar behavior than the inulin chicory, to increment the *Bifidobacteria* microorganism growth in humans.

Dairy products are important food matrices in which probiotic bacteria and prebiotics have been supplemented in, generating products with an excellent reputation among consumers [10, 18, 29]. Therefore, the dairy industry has modified several existing items and developed novel ones with healthy properties, such as products with decreased fat, with fortification of minerals and vitamins, with incorporation of omega-3 and fiber [30-32], among other modifications and innovations. Now in our days, foods are judged, selected and purchased, not only in terms of taste, satisfaction and nutritional value, but also in terms of their capacity to improve health and well-being for consumers. It is widely recommendable and convenient to enrich a dairy product of high demand, such the Mozzarella cheese, generating a functional food or prebiotic cheese that contributes to disease prevention, health benefits, and consumer satisfaction, beyond its nutritive value. The manufacture of probiotic cheese favored by prebiotic components should exhibit minimal changes in its properties and important health benefits [10, 26].

Even though, several studies have been completed to analyze and know the effect of different compositions [4, 14, 33], processing changes and post-manufacturing factors [8, 12], the incorporation of a new ingredient and manufacturing process modifications for Mozzarella cheese, will cause different effects that should be studied and known. Thus, the objective of this research was to determine the effect of agavin addition as a prebiotic compound in a Mozzarella cheese type and to know the effect of lactic acid incorporation as part of the manufacture process, studying its physicochemical and physical properties and also assessing the sensory acceptance of the elaborated functional cheese; analyzed as a fresh dairy product and during its storage.

2. Materials and Methods

2.1 Materials

Five brands of commercial pasta filata cheeses: Mozzarella (La Esmeralda, Mexico), Manchego (Nestlé, Mexico), Emmental (Lactalis International, France), Oaxaca (Lala, Mexico) and Provolone (La Clette, France) were characterized to have a commercial frame of comparison. Those materials utilized for cheese making were obtained from a local supermarket and some commercial suppliers. Pasteurized whole milk with 3.2% fat (Alpura, Mexico), sodium chloride (La Fina, Mexico), drinkable water (Ciel, Mexico). Prebiotic powder fiber from the agave, called agavin (Bioagave® CP Ingredients, Mexico), lactic bacteria starter (*Streptococcus thermophilus*) (Danisco Mexicana, Mexico), rennet coagulating liquid (Cuamex, CHR Hansen, Mexico), lactic acid (85%) (Cedrosa, Mexico) and calcium chloride (Omnichem®, Mexico).

2.2 Manufacture of Mozzarella type cheese

By following the process methodology of Zisu and Shah [14] and Jana and Mandal [11] with some small modifications, twelve batches of Mozzarella cheese were manufactured in two stages that were identified as specific systems, utilizing 8 L of pasteurized whole milk (3.2% fat) for each batch. Two of them without agavin, in which one was without lactic acid and the other with it, both were considered as control systems (C1 and C2). The other ten systems or samples (S1 to S10) were prepared, including different levels of agavin (0.70-3.33%), four of them without acid (S1, S2, S5 and S6), and six with the lactic acid addition (S3, S4, S7, S8, S9, S10) as it is shown in Table 1. The concentration levels of agavin were considered as appropriate by other authors and by ourselves, based on our previous formulation tests.

Cheese system	Agavin (%)	Lactic acid (%)
C1	0	0
C2	0	0.0375
S1	0.70	0
S2	1.11	0
S3	0.70	0.0375
S4	1.11	0.0375
S5	1.65	0

S6	3.33	0
S7	1.65	0.0375
S8	3.33	0.0375
S9	0.90	0.0375
S10	1.40	0.0375

Table 1: Mozzarella type cheese systems (with levels of agavin and lactic acid).

Therefore, three variables were considered: i) agavin concentration (0.70, 1.11, 1.65 and 3.33%), ii) lactic acid presence (0.0375%) or none, and iii) storage time (1, 11, 21 and 31 days). Each batch was inoculated with *S. thermophilus* at a concentration of 0.002% (w/w), in which the milk was tempered up to 37°C and kept during 40 min. The agavin was added to a small volume of heated milk until its total dispersion, and lately incorporated to the correspondent batch. All systems included CaCl₂ (0.01% w/w) as part of the formulation. Seven batches (C2, S3, S4, S7, S8, S9 and S10) were pre-acidified by direct addition of lactic acid (0.0375% v/v). Milk of every batch was coagulated with a rennet commercial liquid (0.015% v/v) at 36°C and held 1 h without agitation. Formed curd was cut in cubes of 0.5 cm, and after cubing, the curd was cooked through, 20 min while the temperature gradually was increased up to 40°C. After heating the whey was drained at pH 5.4 and the curd was cheddared into slabs and maintained until reach a pH of 5.2. Then, every system was hand stretched for 3 minutes in hot water at 85°C, with a volume of water 2.5 times the weight of the curd. The slabs were weighed and the cheese blocks were formed, in which salt (1.5% w/w) was added for rubbing action, and held 1 hour while the product temperature decreased. Finally, all systems or samples were stored at refrigeration (6°C ± 1) into plastic bags, and analyzed at days 1, 11, 21 and 31.

2.3 Physicochemical analysis

The Mozzarella cheese systems were analyzed with standard methods from AOAC [34]. Moisture measuring was based on water evaporation, protein was measured by micro-Kjeldahl, while fat content was quantified with the Mojonnier method. pH was determined with a digital potentiometer UB-5 (Denver Instruments, NY, USA). Acidity was quantified with a titratable method using NaOH 0.1 N, and expressed as percentage of lactic acid for milk, in samples of 20 mL, by method 947.05 [34]. Whereas cheese acidity, in which method 920.124 was followed [34], utilized 10 g of cheese mixed with 105 mL of water at 40°C, thus after complete mixing and filtering, 25 mL of filtered liquid was used for titration. For total dietetic fiber (TDF) determination, also an official method (985.29, [35]) was considered; after fat removal and drying of the sample an enzymatic digestion (α -amylase, protease, amylo-glucosidase) was applied to finish with the washing and purification of the insoluble fiber. TDF was determined at first day and only for some cheese systems. All physicochemical analyses were carried out by duplicate.

2.4 Physical determinations

2.4.1 Color: A color measuring instrument (Gardner System 05, Hunter Labs, Reston, VA, USA) was used to obtain the L , a and b Hunter parameters of the cheese systems and to calculate the net change of color in accordance to equation 1.

$$\Delta E = \sqrt{(L_i - L_o)^2 + (a_i - a_o)^2 + (b_i - b_o)^2} \quad (1)$$

Where: L_i , a_i and b_i are the correspondent parameters at different days (11, 21 and 31), whereas L_o , a_o and b_o are the reference or initial data (day 1).

2.5 Texture analysis

Textural parameters of cheeses including compression, penetration and TPA tests [36] were carried out with a Texture Analyzer TA.XT2 (Texture Technologies Corporation, Scardale, NY, USA) at room temperature (22°C). Cheese samples were shaped using a cylindrical probe with 2 cm diameter and 1.5 cm height, each sample was allowed to equilibrate at room temperature during 10 min after its removing from refrigerated storage. For compression test, the samples were compressed up to 50% of their original size at a speed of 0.5 mm/s with a plate of 3.6 cm diameter. The penetration test was carried out with a needle of 5 mm diameter at a speed of 1.7 mm/s, and the samples were penetrated 50% of their original size. Other textural characteristics of the cheeses, including hardness, cohesiveness and springiness were assessed by a TPA test, with the same cylindrical samples and using a speed of 1 mm/s, applying a double compression, with a plate of 3.6 cm diameter, until the 80% of their original size, as recorded by the Texture Analyzer instrument.

2.6 Elongation and meltability

The elongation test was completed by adapting the reported method from Reid and Yan [8]. A designed hook fixture was adapted to the Texture Analyzer TA.XT2 and used to evaluate the stretchability of the Mozzarella samples. Cheese sample was cut as a plate shape of 3.0 cm long and 1.25 cm wide, it was subjected by two sides (central extreme points), fitting a rate of 1 mm/s, with a maximum elongation of 5.0 cm, thus recording the rupture force [8, 37]. This test was applied to the cheese system, before and after-melting of samples heated during 1 min in a microwave and lately held out 1 min to cool down, at the different days of storage. For meltability a test modified from the Schreiber one, was utilized with cheese samples of 2 cm of diameter and 1 cm of thickness. Sample after refrigeration (4°C-30 min) was placed in a glass Petri dish (with filter paper) and then heated in an oven at 200°C for 1 min. After melting, four measures were taken at each direction (45°) [8, 38], computing the meltability capacity in accordance with the equation 2.

$$MC = \frac{D_f - D_i}{D_i} \times 100 \quad (2)$$

Where: MC is the melting capacity (%), D_f is the final average diameter (cm), and D_i is the initial diameter (cm), for each cheese sample.

2.7 Viscoelastic properties

A stress-controlled Universal Dynamic Rheometer (Paar Physica UDS 200, Germany) was used to obtain the viscoelastic properties of Mozzarella samples employing a parallel-plate fixture. Cylindrical samples of the same size (2.0 cm diameter, 1.5 cm height), previously equilibrated at room temperature (21°C), were utilized for viscoelasticity measurements. Once that the linear viscoelastic region was determined by a stress sweep; a frequency sweep over the range of 0.01-10 Hz was performed, the values of the storage modulus (G'), loss modulus (G'') and loss tangent angle (δ), were obtained as a function of frequency and recorded by using the Rheoplus software 32 V2.81.

2.8 Sensory assessment

Sensory tests were carried out with only five systems and only two levels of agavin, the selected samples were C1 as the control, and S1, S2, S3 and S4 as representative of the lower agavin levels, in which two of them included the lactic acid as part of the manufacturing process. Samples with higher levels of agavin were not available. The assessment was completed at room temperature by utilizing a hedonic scale with a magnitude of 5 for maximum like, and 1 for maximum dislike. The test was completed with 25 non trained judges (students and workers), in which the questionnaire was divided in 2 parts: the first one corresponded to a general acceptability of the product, evaluating color, flavor and appearance; whereas the second part corresponded to the textural acceptability of hardness, chewiness, and springiness; terms that were defined to the judges, but without a training process.

2.9 Statistical analysis

ANOVA of the General Linear Model type from the Minitab package (V.16 Minitab Inc., State College, Pennsylvania, USA) with 3 factors was carried out, to study the effects of the agavin addition, pre-acidification and shelf life.

3. Results and Discussion

In order to elaborate and study a functional Mozzarella type cheese, two experimental stages were completed. The first and shorter part, in which a compositional analysis of five commercial cheeses of pasta filata type was carried out, in order to obtain a comparative frame of the Mozzarella cheese characteristics. And a second and long stage, as the most important of this research, included some modifications that were incorporated through the cheese manufacturing process. Measuring the sample properties at different storage times.

3.1 Characterization of commercial cheeses

Five brands of pasta filata cheese were acquired and analyzed, they corresponded to Mozzarella, Manchego, Oaxaca, Emmental and Provolone types, using 2 replicates of each one for physicochemical determinations. Thus, ten samples were utilized for this purpose. The measured mean composition values exhibited a moisture range of 20.2% (± 0.20) for Emmental, to 35.9% (± 1.22) for Oaxaca; a fat range of 17.5% (± 0.84) for Provolone, to 47.4% (± 1.66) for Mozzarella; a protein range of 17.7% (± 0.48) for Provolone, to 21.8% (± 0.28) for Manchego; giving thus a

protein/fat ratio of 0.36-1.29. A range of 0.03 to 0.09% was quantified for acidity and a pH between 4.41 and 5.47 was also determined. In color parameters, a range of 57 to 75 for luminosity, -5.0 to 2.5 for redness and 16 to 22 for yellowness were recorded. As expected, these determinations, although representative of meltable commercial cheeses, were importantly affected by the time passed since they were manufactured, and as expected all of them showed varied moisture and fat contents with tendency to white-cream colors.

Mozzarella cheese showed the next averages: 25.9% for moisture, 47.3% for fat, 18.4% for protein (0.39 protein/fat ratio), 0.09 for acidity, 5.1 for pH and 74 for *L*, -4.3 for *a*, 21.5 for *b*, respectively. Gunasekaran and Ak [39] included a typical composition of a variety of cheeses, such as Cheddar with moisture of 36.7%, Provolone 40.9% and Mozzarella 54.1%, emphasizing the influence of the making process, formulation, degree of ripening and time of retailing on cheese composition. Therefore, it can be assumed that the difference in moisture between commercial and reported cheeses from some studies, are due to the freshness and elapsed time of manufacturing. Cheeses from the supermarket have experimented biochemical and physical changes through the shelf time.

Ismail et al. [4] prepared Mozzarella cheese to improve their properties by adding some ingredients, such as denatured whey protein, quantifying high fat percentages in a range of 26.4 to 34.5. Sulieman et al. [40] also manufactured Mozzarella cheese from cow milk and studied the effect of time (30 days), recording some augments in pH from 4.93 to 5.21, in protein from 17.7 to 26.1% and in fat also, from 15.1 to 24.9%. This work showed an important increase in component contents influenced by the storage period and attributed to the water evaporation happened through it. That most likely was the physical situation with the analyzed commercial samples of our study. Ma et al. [12] also conducted a research, manufacturing Mozzarella cheeses in a wide range of compositions and production processes, to have three groups with different fat content obtaining a protein/fat ratio of 0.71 and a draining pH of 5.9, for the high fat content group. Therefore, and as it has been recognized, the composition and properties are functions of the formulation, manufacturing process and storage time. Then, from the measured and reported parameters, a diversity of Mozzarella cheese characteristics has been and can be found

3.2 Development of the functional Mozzarella type cheese

With respect to the manufacturing process, the modifications incorporated in this work were next:

- i. Cow milk with 3.2% fat.
- ii. Addition of agavin, as a functional ingredient. Once the agavin was diluted in milk, this milk was mixed by agitation with the rest of standardized milk up to complete a batch of eight liters, at the four aforementioned levels.
- iii. Addition of lactic acid at a concentration of 0.375% as a pre-acidification step being previous to the addition of rennet, in seven of the ten analyzed systems.

The rest of the manufacturing process was previously described in the methodology, by following to Zisu and Shah [14].

3.3 Physicochemical characteristics

3.3.1 Composition: The cow milk used for the manufacturing of the functional Mozzarella type cheese exhibited a typical composition, moisture $87.37\% \pm 0.05$, fat $3.2\% \pm 0.06$, acidity $0.318\% \pm 0.06$, pH 6.65 ± 0.01 . With this milk, all the cheeses were manufactured following the different formulations or cheese systems, in which a yield of 11-7 to 13.1% (w/w) was obtained. The proximal composition and physicochemical properties corresponding to the studied cheese systems are included in Table 2.

System	Moisture (%)	Fat (%)	Protein (%)	Acidity (% lactic acid)	pH
C1	49.65 ± 0.00^b	14.01 ± 0.01^b	17.20 ± 0.00^b	0.086 ± 0.00^b	5.54 ± 0.05^a
C2	52.24 ± 0.00^b	9.14 ± 0.01^b	19.75 ± 0.02^b	0.110 ± 0.00^b	5.00 ± 0.03^a
S1	50.19 ± 0.02^b	13.50 ± 0.02^b	13.19 ± 0.01^b	0.068 ± 0.00^b	5.75 ± 0.11^a
S2	49.46 ± 0.01^b	11.77 ± 0.02^b	18.25 ± 0.01^b	0.079 ± 0.00^b	$5.40 \pm 0.01^{a,b}$
S3	50.68 ± 0.01^b	10.40 ± 0.02^b	14.73 ± 0.03^b	0.086 ± 0.00^b	5.31 ± 0.00^a
S4	51.56 ± 0.00^b	10.74 ± 0.01^b	18.36 ± 0.03^b	0.097 ± 0.00^b	$5.06 \pm 0.04^{a,b}$
S5	$50.58 \pm 0.01^{a,b}$	9.24 ± 0.01^b	25.18 ± 0.10^b	0.054 ± 0.00^b	5.25 ± 0.04^b
S6	46.93 ± 0.01^b	9.80 ± 0.01^b	13.88 ± 0.01^b	0.065 ± 0.00^b	5.25 ± 0.04^b
S7	$52.89 \pm 0.00^{a,b}$	8.89 ± 0.00^b	13.34 ± 0.00^b	0.099 ± 0.00^b	4.54 ± 0.01^b
S8	51.50 ± 0.00^b	7.76 ± 0.00^b	15.30 ± 0.02^b	0.077 ± 0.00^b	4.91 ± 0.04^b
S9	$52.16 \pm 0.02^{a,b}$	ND	$21.95 \pm 0.01^{a,b}$	0.068 ± 0.00^a	4.99 ± 0.06^b
S10	52.94 ± 0.01^a	ND	21.56 ± 0.01^b	$0.090 \pm 0.00^{a,b}$	4.81 ± 0.02^b

For each column, values not sharing letters differ significantly ($p < 0.05$). ND-not determined

Table 2: Physicochemical characteristics of the formulated and analyzed Mozzarella cheeses at day 1.

It may be observed that the prepared systems showed a relative low moisture ($>46.9\%$ and $<52.9\%$) and high fat content (7.70-14.0%) in an inverse relationship, in which the fat content of the cheese was conditioned by the standardization of the utilized milk. Similar moisture content (48-57%) was expressed by Guinee et al. [33] for Mozzarella cheese; whereas similar fat content was reported by Ma et al. [12] and Zisu and Shah [14], for those Mozzarella cheeses identified as a low-fat content group, with 11.4% fat and 7.71-9.44%, respectively. While Zisu and Shah [14] reported a moisture range of 52.8 to 60.3% using fat-replacers, and Ismail et al. [4] recorded a moisture content higher to 84% using denatured whey protein, for their studied Mozzarella cheeses. High humidity and low-fat values were correlated by Hernández et al. [41] in their study, in which the high volume of water contributed to a minor fat content retention in the molecular space of the cheese. In other work, Zisu and Shah [14], established that the presence of agavin increased the moisture retention and competed by a molecular space in the protein matrix. Further the pre-acidification also contributed to higher moisture content of our studied low fat Mozzarella cheeses. The protein content was in the range of 13.2-25.2% without a general trend, in which the

system S5 presented the highest value, and giving eight of the ten analyzed systems a fat/protein ratio range of 0.39-1.10.

It may be considered that the manufacturing process and particularly the pre-acidification stage contributed to increase the hydration phenomenon in C2, S4, S7, S8, S9 and S10 cheeses with higher moisture content (>51.5%). The statistical analyses of the composition showed that agavin addition influenced significantly ($p < 0.05$) the moisture, protein, acidity and pH values of the studied cheese systems, but did not affect their fat content. The increase in moisture obtained with respect to the control C1, for those systems including agavin, with the exception of systems S2 and S6, may be attributable to the presence of oligosaccharides from it.

The last comment is in agreement with Leewah et al. [42], who cited that the fiber content has been related to water retention in food systems, due to its capacity for chelation, dissolution and emulsification; being the case of the agavin added to the studied Mozzarella cheese formulations. Franck [24] also cited a good water retention capacity for the chicory inulin, forming a three-dimensional network. Taking account of the analyzed cheese composition, with 46.9-52.9% moisture and 7.7-14% fat, the prepared systems of our study may be considered as low moisture Mozzarella cheeses, cited also as part-skim by Kindstedt and Fox [1], and similar to the classification cited by Muliawan and Hatzikiriakos [43], and by Ayyash and Shah [44]. The obtained yields in this study (11.7-13.1%) are up than those obtained (7.71-9.44%) for low fat Mozzarella cheeses by Zisu and Shah [14].

3.3.2 Fiber determination: Even though all cheese systems (none control) were added with agavin, only five of them (S4, S5, S6, S7 and S8) and one control (C2) were used to the determination of TDF on the first day. The quantity of TDF measured for C2 was the reference, and as it was expected the fiber was higher for the five cheese systems ($> 2.44 \text{ g}/100 \text{ g} \pm 0.35$), that was related to the agavin concentration. Thus, this food item could contribute to the recommended daily fiber consumption by different countries: Colombia with 15-20 g, France 25-30 g, Japan 20-30 g, Mexico 35 g, Netherlands 30-40 g, South Africa 30-40 g, USA 18-21 g, world $> 25 \text{ g}$, just to mention some [45], in addition to the other nutritive components of Mozzarella cheese.

3.3.3 Storage: With respect to the storage, a general and small decreasing in moisture content was observed, consequently an increasing in the other cheese components was recorded, being both changes in combination with biochemical transformations. The quantified composition for the studied systems (data not included) exhibited some changes, mainly detected between day 1 and day 21, with a magnitude lower than 1.2% in moisture. Some systems (six of fourteen) continued with water loss up to day 31 ($< 1\%$), in contrary to the rest of them, that gained a little of water from day 21 ($< 1.8\%$). Therefore, a significant effect ($p < 0.05$) of the storage time on cheese moisture was obtained. As consequence of the moisture loss, proteolysis and lipolysis reactions, the studied cheeses exhibited the next variations after 31 days of storage: proteins suffered a small decreasing (less than 3%), whereas fat recorded 1-5% of decreasing. At the beginning (day 1) the pH ranged from 4.5-5.8 (Table 2), considered as convenient values by Miocinovic et al. [46] for cheeses, prepared by acidification.

Higher pH values corresponded to cheese systems without acid addition (C1, S1, S2, S5 and S6) with exception of S3 system with higher pH than S5 and S6. These pH values are comparable to those reported (5.4-5.9) by Guinee et al. [33] for low moisture Mozzarella cheeses. This parameter decreased through the storage time to 4.3-5.5 (data not shown). The percentage of lactic acid varied from 0.054 to 0.110 in the fresh cheeses, and the relative high acidity (>0.090) for systems C2, S4, S7 and S10, is due to the direct addition of this acid. The lactic acid augmented in most of the systems (eight of twelve) with the time and decreased lightly in the rest of them, as a consequence of the metabolic activity due to bacteria development, proteolysis and lipolysis processes proper of the Mozzarella cheese. Two of the studied variables, acidification and storage time influenced significantly ($p < 0.05$) both parameters, pH and lactic acid. The agavin had a certain effect, but it was not significant.

In general, the physicochemical properties presented variation due to the formulation (agavin adding), manufacturing process (acid addition, Table 1) and biochemical changes during storage. It is clear that the comparison among Mozzarella cheeses from different composition and varied manufacturing process is limited, because the presence of all components and how they were transformed, affecting the physicochemical, rheological, textural and sensory properties of each specific system.

3.3.4 Color determinations: The color is a very important quality and physical property of foods. Then, the Hunter parameters of prepared cheese systems were evaluated to know the effect of the studied factors. Overall and as it was expected, the color of cheese systems was of green (negative a) and yellow tones (positive b), with creamy and luminous external appearances. The evolution of these parameters through the analyzed storage period is presented in Tables 3-6. The luminosity decreased, whereas the other two parameters showed small variations in magnitude.

System	L	a	b
C1	84.835 ± 083	-2.395 ± 0.16	13.185 ± 0.70
C2	78.000 ± 0.91	-2.260 ± 0.06	8.680 ± 0.04
S1	82.400 ± 0.95	-2.225 ± 0.35	12.415 ± 1.10
S2	81.285 ± 0.12	-2.480 ± 0.20	12.520 ± 1.29
S3	80.395 ± 0.05	-2.325 ± 0.33	11.960 ± 0.91
S4	80.765 ± 0.53	-2.170 ± 0.16	11.250 ± 1.39
S5	80.195 ± 2.72	-3.605 ± 0.15	11.370 ± 1.44
S6	78.125 ± 2.10	-3.550 ± 0.03	11.850 ± 1.12
S7	81.575 ± 2.68	-2.840 ± 0.06	11.855 ± 0.62
S8	81.775 ± 1.12	-2.775 ± 0.11	11.800 ± 0.75
S9	86.205 ± 1.12	-2.430 ± 1.95	12.060 ± 1.95
S10	78.830 ± 1.12	-3.760 ± 0.64	11.620 ± 0.06

Table 3: Color parameters for Mozzarella cheeses at day 1.

System	L	a	b	ΔE
C1	81.155 ± 0.25	-3.235 ± 0.11	13.145 ± 0.29	3.77
C2	78.725 ± 0.19	-3.300 ± 0.04	10.685 ± 0.09	1.43
S1	81.140 ± 0.49	-3.560 ± 0.07	12.295 ± 0.08	1.84
S2	77.270 ± 3.75	-3.405 ± 0.12	13.295 ± 0.43	4.19
S3	74.145 ± 2.34	-2.870 ± 0.03	12.010 ± 0.40	6.27
S4	77.160 ± 0.86	-2.975 ± 0.21	11.685 ± 0.33	3.72
S5	76.41 ± 2.48	-3.015 ± 0.06	12.265 ± 0.27	3.93
S6	74.965 ± 1.11	-2.995 ± 0.08	12.895 ± 0.55	3.37
S7	82.67 ± 0.51	-2.785 ± 0.18	13.485 ± 0.29	1.96
S8	81.845 ± 1.92	-2.740 ± 0.18	13.415 ± 0.51	1.62
S9	85.565 ± 0.08	-1.905 ± 0.11	12.425 ± 0.16	12.16
S10	83.640 ± 0.44	-2.680 ± 0.81	12.965 ± 0.11	2.83

Table 4: Color parameters for Mozzarella cheeses at day 11.

System	L	a	b	ΔE
C1	79.845 ± 0.62	-4.240 ± 0.00	12.640 ± 0.03	5.35
C2	59.095 ± 0.07	-2.555 ± 0.11	8.365 ± 0.02	19.05
S1	75.710 ± 0.91	-4.200 ± 0.04	12.330 ± 0.24	6.97
S2	78.625 ± 1.93	-3.535 ± 0.11	12.700 ± 1.94	2.87
S3	75.210 ± 3.13	-2.985 ± 0.08	12.440 ± 0.79	5.25
S4	77.490 ± 1.05	-3.325 ± 0.25	11.960 ± 1.06	3.54
S5	77.910 ± 0.07	-3.080 ± 0.31	12.995 ± 0.87	2.85
S6	78.795 ± 2.13	-3.235 ± 0.11	14.04 ± 0.36	2.31
S7	75.600 ± 1.27	-2.880 ± 0.17	12.25 ± 0.18	5.99
S8	77.61 ± 0.03	-2.770 ± 0.13	13.00 ± 0.02	4.33
S9	52.245 ± 0.98	-2.060 ± 1.79	7.250 ± 0.06	13.014
S10	56.200 ± 0.02	-2.525 ± 1.03	8.845 ± 0.08	26.01

Table 5: Color parameters for Mozzarella cheeses at day 21.

System	L	a	b	ΔE
C1	79.780 ± 0.04	-3.475 ± 0.12	13.850 ± 0.49	5.21
C2	55.575 ± 0.40	-2.445 ± 0.99	8.960 ± 0.28	22.50

S1	77.395 ± 3.42	-3.320 ± 0.10	12.400 ± 0.99	5.12
S2	77.670 ± 0.69	-3.395 ± 0.04	12.645 ± 0.02	3.73
S3	78.315 ± 0.22	-3.205 ± 0.06	12.965 ± 0.77	2.47
S4	81.695 ± 0.36	-3.315 ± 0.09	13.555 ± 0.29	2.74
S5	78.065 ± 0.74	-3.040 ± 0.31	12.545 ± 0.29	2.50
S6	77.55 ± 2.08	-2.690 ± 0.01	12.595 ± 1.49	1.27
S7	74.405 ± 0.05	-2.750 ± 0.07	12.645 ± 0.29	7.21
S8	75.800 ± 0.58	-3.210 ± 0.18	14.095 ± 0.01	6.42
S9	53.155 ± 0.06	-2.005 ± 3.68	6.870 ± 0.18	9.80
S10	56.595 ± 0.56	-2.030 ± 0.32	7.250 ± 0.07	26.27

Table 6: Color parameters for Mozzarella cheeses at day 31.

It may be observed (Table 3) that fresh cheese showed a high luminosity ($L > 78$), in which S5 with -11.4 and S10 with -11.3 presented the highest a (negative) values at day 1, while S1 and S2 had the highest b values (> 12.4). Small variations were recorded at day 11 (Table 4), a decreasing in luminosity ($L > 74$) and an increasing in a and b parameters were recorded, that was more notable with the storage time, $L > 52$ at days 21 and 31. Most of the cheeses exhibited lower values of b with respect to C1 that was the cheese control with higher fat content. These results indicate that agavin influenced the natural color of cheeses, decreasing the light green-yellow appearance typical of dairy products, due the presence of carotenoids and vitamin B12 [47]. Furthermore, the yellowness was the highest for S2 at day 1, for S7 at day 11, for S6 at day 21 and for S8 at day 31, attributed to the formulation, manufacturing process and biochemical changes developed through the storage.

From color data, recorded at days 1, 11, 21 and 31 (Tables 3-6, respectively), it was obtained a significant influence ($p < 0.05$) of agavin on redness and yellowness parameters. The luminosity (> 74 in all systems at days 1 and 11) was not significantly affected by the agavin presence, but it was influenced by the pre-acidification. Alvarez et al. [48] determined that high moisture content was related to an increasing in luminosity of cheeses. Even though they have a high water content, three cheese systems (C2, S9 and S10) showed an important decreasing in L (> 52) at days 21 and 31, expressing the effect of storage time. For the net change of color (Tables 4-6), three cheese systems, S2 (1.11% of agavin and without addition of lactic acid), S3 and S9 (with lower agavin and pre-acidified) exhibited a higher net change of color (4.2, 6.3 and 12.2, respectively) at day 11. Whereas systems C2, S9 and S10, changed more their color (19.0, 13.0 and 26.0, correspondently) at day 21; the same last three cheeses, C2 with 22.5, S9 with 9.8 and S10 with 26.3 for ΔE showed the higher values at day 31. Even though these variations of color, the net change (ΔE) as the overall color change parameter evaluation (1.3-26.3), did not show any significant difference ($p > 0.05$), then it may be considered that minimal color changes were developed during the shelf life of these manufactured functional cheeses.

3.3.5 Textural and rheological parameters: Due to the fact than cheese samples were prepared at different stages. No all the systems were characterized using the same textural tests; then the texture and rheology were only evaluated in some of the cheese systems, the correspondent samples will be specified.

3.3.6 Compression and penetration forces: Both textural tests were recorded the same days of storage for all cheese systems (Figures 1 and 2). The values obtained in the compression test ranged from 4.5 to 30 N, being those systems with agavin (S2, S7, S5 and S6 at the first day) that exhibited the higher magnitudes at the first day. Whereas for penetration measuring the forces were lower, in the range of 0.9 to 4.4 N, corresponding similarly the higher values to cheese systems added with agavin (S2, S7, S5, S1 and S8) at the first day, the specific values are observed in Figures 1 and 2. In agreement with Zisu and Shah [14], the pre-acidification treatment could increase the proteolysis reactions contributing to alter forces. These recorded forces express how the mechanical device took more effort to advance against the whole sample in compression and it was lower to penetrate just the central zone of the cheese sample, respectively. Giving the agavin more physical resistance into the correspondent samples. Casiraghi et al. [49] carried out compression test on different types of cheeses, they recorded a range of 30-50 N for uniaxial compression of Mozzarella cheese under different measurement conditions (lubricated, nonbonded and bonded).

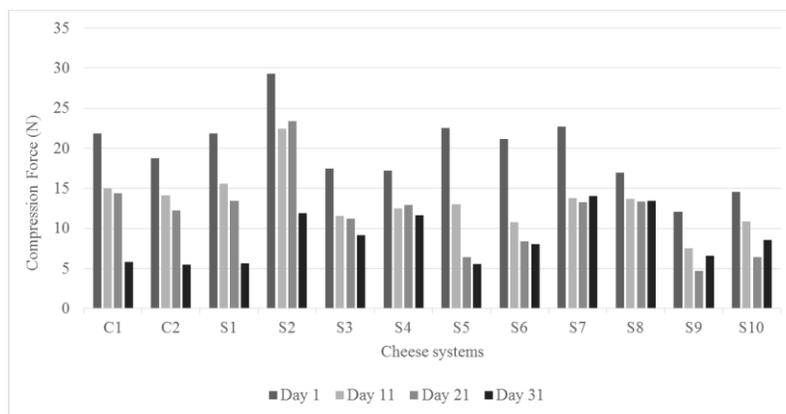


Figure 1: Compression results for all cheese systems through the storage.

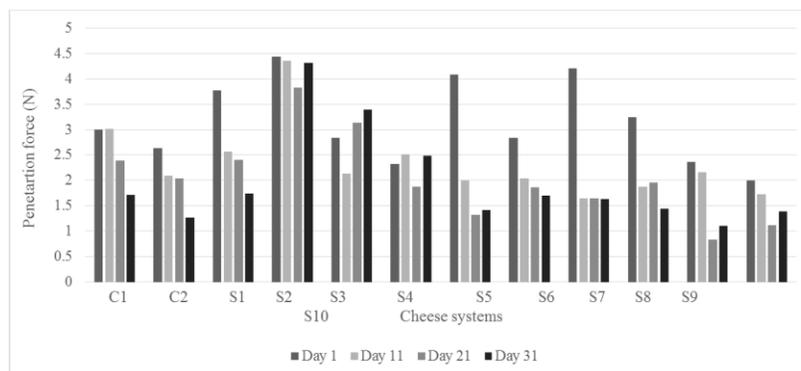


Figure 2: Penetration results for all cheese systems through the storage.

From ANOVA data, it was observed that the three studied variables, agavin addition, pre-acidification and storage time affected significantly ($p < 0.05$) the compression and penetration responses of all cheese systems. Both forces were similar, to cheese controls and all of them showed a decreasing trend as a function of the storage time.

3.3.7 Texture profile analysis: TPA test was applied to four cheese systems (C2, S7, S9, S10) at day 21. From the double compression test, different responses were observed in our study (Figure 3), only some parameters were selected as useful. Cohesiveness (how well the cheese elements hold together, that is evaluated as a ratio of work or energy involved on the second and first bites, 0.26-0.46 dimensionless) and springiness (amount of physical recovery and evaluated as a ratio of deformation length involved with the second and first bites, 0.41-0.73, dimensionless), showed a similar response without a trend. These values of cohesiveness resulted lower than those reported correspondently for low-fat Mozzarella cheeses (0.63-0.74) by Zisu and Shah [14]. Gumminess (as a multiplication of hardness by cohesiveness, 1.38-3.72 N) and chewiness (as a product of hardness, cohesiveness times springiness, 0.56-2.72 N-m) exhibited also a similar response, with a decreasing trend for C2, S7, S9 and S10, respectively. Firmness (data not included in Figure 3) was higher in C2 and S7 systems with 8 N, whereas the other two cheese systems, S9 and S10 recorded a force of 5.48 and 5.24 N. These magnitudes are comparable to hardness recorded for a type of Mozzarella cheese (4-38 N) studied by Zisu and Shah [14], comparable to hardness (8.5-10.1 N) measured for a pasta filata cheese [50], and lower than those hardness values of 37-105 N, reported by Tunick et al. [51] for Mozzarella cheeses after 1-6 weeks of storage.

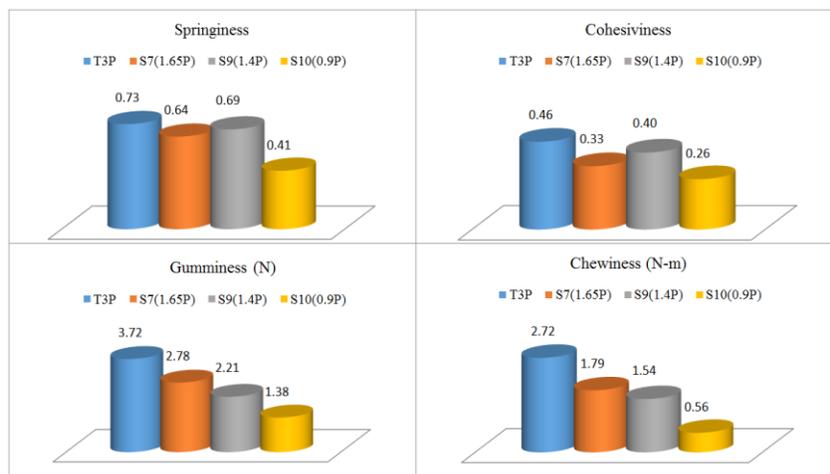


Figure 3: TPA results for cheese systems C2 (T3P), S7, S9 and S10, at day 21.

Our ANOVA results indicated that agavin content affected significantly ($p < 0.05$) the last three texture parameters (gumminess, chewiness and firmness), being the unique factor involved or considered in this TPA test.

3.3.8 Elongation test: The application of this complementary test for texture characterization, exhibited a range of 1.0 to 3.5 N before melting for fresh cheeses, measured as the rupture force between subjected extremes of the cheese sample. The specific magnitudes previous and after melting, that were recorded on different days are

included in Table 7. It is clear and expected that forces needed previous to the melting (<3.5 N) are higher than those needed after melting (<2.0 N) in most of the cheese systems. The effect of temperature and melting contributed to a decreasing of the casein and other components [39, 44]. The recorded values are in accordance with those reported by Gunasekaran and Ak [39] and are also comparable to those strength values at 60 (1.2-2.6 N) and 70°C (0.84-1.8N) reported for Mozzarella by Fife et al. [52]. There was not significant difference between control and cheese with agavin. Whereas the effect of acid addition, is not clear due to the few results obtained for the elongation test, apparently cheeses with lactic acid required lower rupture force, phenomenon that may be an advantage or disadvantage, depending of the final application of this dairy product. It is also observed (Table 7) that rupture force decreased with the storage time.

System	Storage Day							
	Before to melting				After melting			
	1	16	26	31	1	16	26	31
C1	3.48 ± 0.01	0.58 ± 0.26	1.23 ± 0.27	0.90 ± 0.09	0.24 ± 0.01	0.11 ± 0.03	0.28 ± 0.17	0.11 ± 0.01
C2	0.78 ± 0.01	-	-	0.78 ± 0.01	0.78 ± 0.01	-	-	0.78 ± 0.01
S1	-	0.40 ± 0.17	1.36 ± 0.36	0.69 ± 0.10	-	0.26 ± 0.013	0.09 ± 0.04	0.07 ± 0.01
S2	-	2.35 ± 0.25	1.70 ± 0.05	1.00 ± 0.06	-	0.30 ± 0.07	0.08 ± 0.11	0.17 ± 0.09
S3	-	0.98 ± 0.50	1.22 ± 0.57	1.05 ± 0.05	-	0.25 ± 0.11	0.48 ± 0.31	0.05 ± 0.01
S4	-	0.64 ± 0.01	1.25 ± 0.26	0.99 ± 0.06	-	0.17 ± 0.04	0.13 ± 0.03	0.37 ± 0.13
S5	1.97 ± 0.03	-	-	-	0.28 ± 0.03	-	-	-
S6	2.73 ± 0.21	-	-	-	1.96 ± 0.21	-	-	-
S7	0.98 ± 0.39	-	-	0.63 ± 0.03	0.29 ± 0.04	-	-	0.18 ± 0.03
S8	1.25 ± 0.92	-	-	-	1.28 ± 0.09	-	-	-
S9	0.52 ± 0.03	-	-	0.52 ± 0.04	0.30 ± 0.03	-	-	0.18 ± 0.01
S10	0.69 ± 0.02	-	-	0.66 ± 0.02	0.57 ± 0.02	-	-	0.27 ± 0.03

Table 7: Rupture forces (N) in the elongation test for cheese samples through storage.

3.3.9 Melting capacity: For this measurement, in which less objectivity is involved, the cheese samples showed a variety of values (30-157%), as may be appreciated in Table 8. It may be considered than the meltability higher than the recorded for cheese control C1 is a good index; whereas those samples with melt capacity lower than 72.2% as the average value of three determinations in C1 over storage, may be considered as not good pasta filata items, then 23 of the 30 measures showed a good index. Those cheese systems with agavin and lactic acid (S1, S3, S4, S7, S9 and S10) exhibited an average MC higher than 72.2%, being some of them (S4, S7, S9 and S10) very meltable with MC higher that 72.2, as an average value. In general terms these cheeses showed an increasing trend as a function of the storage time.

System	Day			
	1	11	21	31
C1	ND	63.3 ± 0.2	77.8 ± 0.1	75.7 ± 0.2
C2	63 ± 0.6	105.0 ± 0.1	123 ± 0.2	120 ± 1.0
S1	ND	65.9 ± 0.1	157.5 ± 1.5	75.3 ± 0.3
S2	ND	72.4 ± 0.5	41.7 ± 0.5	33.8 ± 0.3
S3	ND	96.8 ± 0.1	38.8 ± 0.5	81.3 ± 0.2
S4	ND	116.1 ± 0.1	91.3 ± 0.3	77.5 ± 0.3
S5	ND	72.0 ± 0.6	42.0 ± 0.3	30.0 ± 0.6
S6	ND	66.0 ± 0.4	87.0 ± 3.0	84.0 ± 2.0
S7	81 ± 0.6	123.0 ± 0.5	144.0 ± 3.6	126.0 ± 2.4
S8	ND	42.0 ± 0.4	48.0 ± 0.3	123.0 ± 2.0
S9	ND	75.0 ± 0.7	114.0 ± 0.6	117.0 ± 1.8
S10	81 ± 0.6	111.0 ± 0.4	120.0 ± 3.0	93.0 ± 2.0

ND-not determined

Table 8: Melting capacity (%), for cheese samples through storage.

And although this parameter expresses the effect of both variables, the agavin that does not favor the melting phenomenon because some of the interaction points are occupied by this compound, while the acidification favors the phenomenon because in contrary, some mineral are lost through this process [16]. It is difficult to know with these limited results what variable was more determinant. However, satisfactory melting capacities were obtained in our cheese samples (>72% for 27 of 38 average values from the cheese systems).

3.4 Viscoelastic and sensory determinations

3.4.1 Viscoelasticity: As expected, all the manufactured cheeses showed an elastic component of higher magnitude than the viscous one, expressing them a firm gel nature. From the moduli magnitudes and $\tan \delta$, it was appreciated that the cheese control 2 was the sample with the highest values for G' and G'' (data not shown) in comparison with those cheeses with added agavin (S8, S7, S9 and S10) and without addition (S6 and S5); being softer gels those corresponding to cheese samples without agavin, in which the difference in magnitude between G' and G'' was lower. The viscoelastic behavior of a cheese has been attributed to the moisture/protein ratio by Rogers et al. [53] that affects the proteinaceous matrix, that obviously was influenced by the agavin presence. Several authors have reported values of the viscoelastic parameters at a frequency of 0.5-1 Hz that represents a value in which a human mouth begins to make structural changes. At that frequency of 1.0 Hz, the viscoelastic parameters for the tested cheese samples are included in Table 9. Observing the magnitudes for both moduli, the most consistent systems (with lower changes between days 1 and 16) corresponded to C2, S7, S9 and S10, they exhibited the highest G' and G'' moduli, three of them with agavin. Higher values of G' (40-50 kPa) and G'' (~ 20 kPa) were obtained by Yun et al. [54] for Mozzarella cheese as a function of the cooking temperature.

System	At day 1		At day 16	
	G' (Pa)	G'' (Pa)	G' (Pa)	G'' (Pa)
C1	29500	9190	6510	2350
C2	15200	4970	16200	5290
S5	846	338	7670	2620
S6	20700	6660	9800	3790
S7	16100	5360	12700	4220
S8	19400	6650	3890	1460
S9	14600	4720	11000	3620
S10	13100	4130	16200	4500

Table 9: Viscoelastic parameters (G' and G'') for some cheese systems at 1 Hz.

In general, all the viscoelastic responses decreased through storage and were more notable in the cheese control than in the cheese with agavin, due to the proteolysis of α and β caseins among other biochemical changes. Mozzarella cheese softens through storage and becomes less elastic. From the ANOVA results, a significant decrease in both moduli as a function of the storage time was detected. The functional changes that take place during the storage and aging have been related directly to the proteolysis phenomenon [5, 14, 44, 55].

3.4.2 Sensory assessment: Only five selected systems (C1, S1, S2, S3, S4) as representative of the whole set, were sensory assessed at day 11. The sensory assessment indicated qualifications of good like in flavor and aroma, a range of 3.04-3.60 of 5 points was recorded for the four cheeses with agavin, in which the S7 (including acid and agavin) presented the lowest values for both attributes. The color was qualified in a range of 2.65-3.21, the texture had 3.08-3.52 and the general acceptance was in a range of 3.13 to 3.73; without any significant difference between the cheese control and the other four cheese systems, two of them (S1 and S2) with lactic acid and the other two (S3 and S4) added with agavin. Comparative sensory results were obtained by Machuca et al. [13], although they used a scale of 7 points, in cheeses made with inulin and oligofructose, used as prebiotic.

The statistical results showed up that consumer could not identify any additional flavor or texture due to agavin, not even the slightly sweet flavor that this ingredient presents by itself. It is assumed that in cheese, the flavor was not modified or perceived by the additions of prebiotic and acid. And the conclusion was that the four cheeses liked with moderation. Other sensory parameters (data non shown) were monitored with a technique of Analysis of Principal Components and Ascendent Classification, and they corroborated the relationship of the agavin with moisture and protein and between lactic acid with acidity and pH, besides the other physicochemical and physical properties.

Conclusions

A functional Mozzarella cheese of low moisture was formulated and manufactured with a prebiotic compound, the agavin. Twelve cheese systems were prepared, characterized and studied fresh and stored. The Mozzarella manufacturing process was modified to incorporate the agavin and the lactic acid with overall good results and yields. The presence of prebiotic and lactic acid influenced some properties of the Mozzarella cheese, but the essential appearance, properties and characteristics of this pasta filata item were conserved. Therefore, combining both factors, it is possible to improve the low-fat Mozzarella cheese, properties such as the flavor, functional and texture, as well as yield may be improved.

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