

Research Article

Colourless Agar for Enhancing Colour Contrast between Microbial Colonies and Agar

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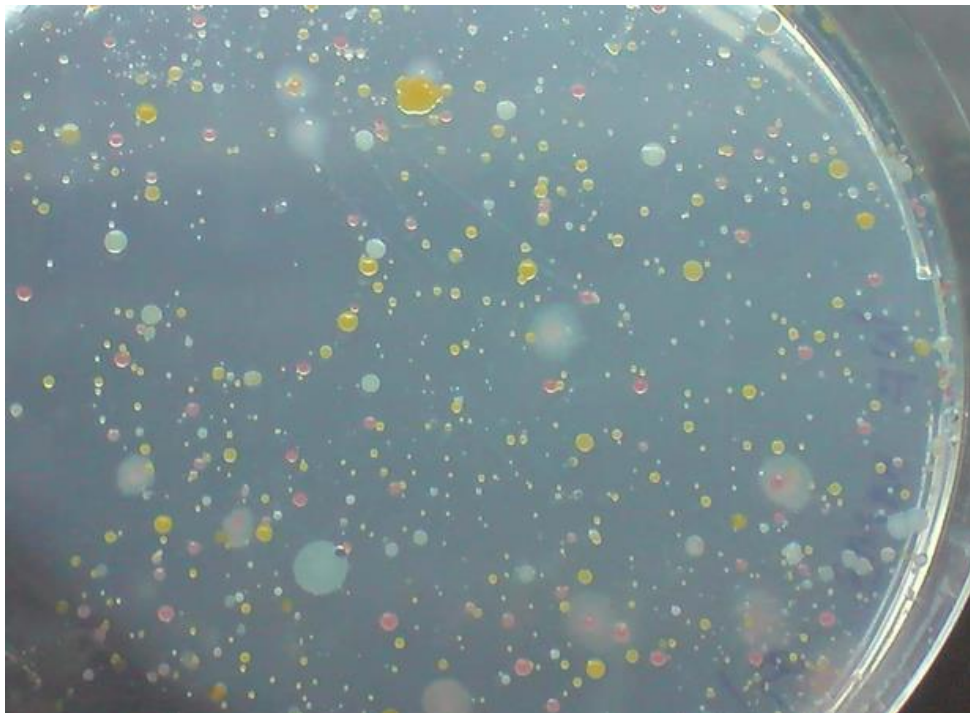
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Graphical Abstract



Short Description

By separately sterilizing glucose and ammonium compounds, a method was developed for creating colourless agar, that helped improve colour contrast between microbial colonies and agar. With no generation of growth inhibitory compounds during agar preparation, colourless agar was able to support the growth of a wide variety of microbial species including common bacteria such as *Escherichia coli* DH5 α , *Bacillus subtilis* NRS-762, and *Pseudomonas protegens* Pf-5. Additionally, the formulated colourless agar was able to support a cell density useful for profiling microbial diversity in different environments. Show here is the microbial flora from deionized water cultivated on formulated colourless agar at 30 $^{\circ}$ c after spread plate inoculation, where small colonies are more easily identified due to high optical transparency of the colourless agar.

Abstract

Lack of colour contrast hampers automated identification of colonies on agar, given that many microbial colonies are of the same colour as the background colour of most agar: beige colour. On the other hand, a colourless agar could increase the colour contrast between the agar background and microbial colonies. But, the challenge in preparing a colourless agar comes from the formation of coloured compounds when sugars and ammonium compounds are sterilized together in an autoclave. Hence, by separating glucose and ammonium compounds into different solutions for autoclave sterilization, a method was developed for preparing colourless agar that remained colourless even with 1 g/L of yeast extract supplementation. Specifically, three separate solutions were used in reconstituting the colourless agar at ~48 $^{\circ}$ c after individual sterilization. Solution A comprised glucose, MgSO $_4$ and agar powder; Solution B contained NH $_4$ Cl, K $_2$ HPO $_4$, KH $_2$ PO $_4$, and

NaCl; and Solution C was yeast extract solution for providing vitamins and trace elements needed by microorganisms unable to grow in minimal medium. Reconstituted colourless agar could be poured into agar plates using standard techniques and had a viscosity similar to many commercial agar. Composition of the colourless agar medium was [g/L]: D-Glucose, 2.0; NH $_4$ Cl, 0.5; K $_2$ HPO $_4$, 0.5; KH $_2$ PO $_4$, 0.1; NaCl, 0.5; MgSO $_4$.7H $_2$ O, 1.0; Yeast extract, 1.0; Agar, 15.0. On observation against varied background images, the formulated colourless agar lacked the yellow tinge present in LB Lennox agar and had greater optical transparency. Good growth of common bacteria such as *Escherichia coli* DH5 α (ATCC 53868), *Bacillus subtilis* NRS-762 (ATCC 8473) and *Pseudomonas protegens* Pf-5 (ATCC BAA-477) was observed in both liquid and solid versions of the formulated colourless agar medium. Specifically, colonies of *E. coli* DH5 α , *B. subtilis* NRS762 and *P. protegens* Pf-5 exhibited similar morphology and characteristics compared to growth on LB Lennox agar. Use of the formulated agar medium in cultivating microbial flora present in deionized water recovered many types of colonies, which suggested that the medium was able to support a variety of microbial species. Collectively, by separately sterilizing sugars and ammonium compounds, a method was developed for preparing colourless agar which helped improve colour contrast for colony identification and counting. The formulated colourless agar could find use in various microbial ecology studies seeking to profile microbes from different environmental niches.

Keywords: Colourless agar; Glucose; Ammonium compounds; Coloured compounds; Autoclave sterilization; *Escherichia coli*; *Bacillus subtilis*; *Pseudomonas protegens*; Yeast extract; Colour contrast

Highlights

- 1) Colourless agar could improve colour contrast between colonies and agar medium. By heat sterilizing glucose and ammonium compounds separately, followed by reconstitution at -48°C , a method was developed for preparing colourless agar.
- 2) Greater optical transparency of colourless agar enables better identification of microbial colonies.
- 3) Good growth of *Escherichia coli* DH5 α , *Bacillus subtilis* NRS-762 and *Pseudomonas protegens* Pf-5 in liquid versions of colourless agar medium revealed that the medium was able to support a reasonable cell density and, more importantly, growth inhibitory compounds were not generated during agar preparation.
- 4) Furthermore, colourless agar medium was able to recover morphologies of *E. coli* DH5 α , *B. subtilis* NRS-762 and *P. protegens* Pf-5 typical of growth on LB Lennox agar. However, white colonies of *E. coli* DH5 α and *B. subtilis* NRS-762 were obtained instead of beige ones.
- 5) Ability to support the growth of a variety of microbial species from deionized water meant that the formulated colourless agar was useful for the cultivation of different species of microbes.

Importance of the work

Lack of colour contrast hampers automated colony identification and counting given the difficulty of discerning small yellowish microbial colonies against beige colour, which is the common colour of agar background. Colourless agar, on the other hand, could enhance the colour contrast between colonies and agar background. However, preparation of colourless agar is hampered by heat induced reaction between sugars and ammonium compounds that generates chromogenic compounds. By separately sterilizing glucose and ammonium compounds, this study reports a simple method for the preparation of colourless agar that could help enhance the colour contrast between agar background and colonies of any colour. Experiment results revealed that colourless agar remained colourless even with addition of yeast extract at 1 g/L. Growth inhibitory compounds are likely not generated during agar preparation given good growth of a variety of common bacteria in both liquid and solid versions of the colourless agar. Overall, a simple method for preparing colourless agar was validated with good growth of a variety of microorganisms from deionized water that indicated growth inhibitory compounds were not generated during agar

preparation. The formulated colourless agar could find use in a wide variety of applications such as in microbial ecology studies.

Introduction

Most commercially available agars are beige in colour, which is also the colour of many microbial colonies. Thus, there is a lack of colour contrast between microbial colonies and agar, which hampers the identification of small colonies by visual observation as well as automated cell counting software able to differentiate a colony from an agar background during cell counting experiments.^{1 2 3} To ameliorate the problem, a colourless agar would be preferable given that it provides good colour contrast between microbial colonies and agar background.

However, reaction between sugars and ammonium compounds during autoclave sterilization generates coloured compounds that lend a reddish colour to the agar; thereby, preventing the formation of a colourless agar. In this study, an approach of separately sterilizing glucose and ammonium compounds, followed by reconstitution after the

solutions have cooled to ~48 °C was used in preparing colourless agar. Additionally, given that many microorganisms could not survive in environments without an input of vitamins and growth factors, the prepared colourless agar must be constituted with small amounts of yeast extract while retaining optical transparency. Experiments revealed that the colourless agar remained colourless even with 1 g/L of yeast extract supplementation. Besides visual observations of the optical transparency of the reconstituted agar, growth experiments were also used in assessing the ability of the formulated agar medium in cultivating a variety of common bacteria (i.e., *Escherichia coli* DH5 α (ATCC 53868), *Bacillus subtilis* NRS-762 (ATCC 8473), and *Pseudomonas protegens* Pf-5 (ATCC BAA477)) in both liquid and solid medium format.

Materials and methods

Materials

LB Lennox medium was purchased from Difco and used as is. Composition of LB Lennox medium is [g/L]: Tryptone, 10.0; Yeast extract, 5.0; NaCl, 5.0. Composition of formulated colourless agar medium is [g/L]: D-Glucose, 2.0; NH₄Cl, 0.5; K₂HPO₄, 0.5; KH₂PO₄, 0.1; NaCl, 0.5; MgSO₄·7H₂O, 1.0; Yeast extract, 1.0; Agar, 15.0. Composition of M9 medium is [g/L]: DGlucose, 4.0; NH₄Cl, 1.0; Na₂HPO₄, 6.78; NaH₂PO₄, 3.0; NaCl, 0.5. Composition of LB Lennox agar is [g/L]: Tryptone, 10.0; Yeast extract, 5.0; NaCl, 5.0; Agar, 15.0. R2A agar was purchased from Merck and used as is. Composition of R2A agar was [g/L]: yeast extract, 0.5; Proteose Peptone, 0.5; Casein hydrolysate, 0.5; Glucose, 0.5; Starch soluble, 0.5; Sodium pyruvate, 0.3; K₂HPO₄, 0.3; MgSO₄, 0.024; Agar, 15.0.

Preparation of colourless agar medium

Solution A comprised 0.4g of D-Glucose, 0.2g of MgSO₄·7H₂O and 3.0g of agar powder in 200 mL of deionized water in a 500 mL glass conical flask. Solution B contained 0.5g of K₂HPO₄, 0.5g of NH₄Cl, 0.1g of KH₂PO₄ and 0.5g of NaCl in 50 mL of deionized water in a 100 mL Schott Duran glass bottle. Solution C contained 1g of yeast extract in 50 mL of deionized water in a 100 mL Schott Duran glass bottle. Solution A, B and C were sterilized separately at 121 °C for 20 minutes.

After cooling to ~48 °C, 10 mL of the sterilized Solution B and C was added to 200 mL of Solution A under aseptic conditions to reconstitute the colourless agar medium, which could be poured into agar plates using standard techniques. The cooling step is important for preventing heat induced reactions between sugars and ammonium compounds after reconstitution of the agar medium.

Growth of bacteria in liquid versions of formulated colourless agar medium

Stock cultures of *Escherichia coli* DH5 α , *Bacillus subtilis* NRS-762 and *Pseudomonas protegens* Pf-5 were prepared in 40% glycerol and stored at -70°C prior to use. *E. coli* DH5 α and *B. subtilis* NRS-762 were activated by inoculating a glycerol stock culture in 100 mL of LB Lennox medium in 250 mL glass shake flask with cotton plugs at 37°C and 30°C, respectively with 230 rpm rotational shaking in a temperature controlled incubator (orbital diameter: 25 mm, Yih Der LM570RD, Taiwan). On the other hand, *P. protegens* Pf-5 was activated similarly, but in M9 medium given its ability to grow in minimal salts medium. After 24 hours of incubation, 1 mL of seed cultures was used in inoculating experiment cultures in the respective media used for the study. Incubation conditions were as follows: *E. coli* DH5 α

(37°C and 230 rpm rotational shaking), *B. subtilis* NRS-762 (30°C and 230 rpm rotational shaking), *P. protegens* Pf-5 (30°C and 230 rpm rotational shaking). Three biological replicates were performed for each experiment.

Measurement of optical density and pH

At appropriate time points, 5 mL of culture was withdrawn from the shake flasks for optical density and pH measurement. Optical density was measured via a UV-Visible spectrophotometer (Shimadzu Biospec Mini) with a quartz cuvette of 10 mm pathlength (volume: 3.5 ml). Dilution with deionized water was performed if the optical density at 600 nm exceeded 1. pH was measured with an Orion 9156 BNWP pH probe outfitted to a Mettler Toledo Delta 320 pH meter.

Growth of bacteria on solid formulated colourless agar medium

E. coli DH5α, *B. subtilis* NRS-762, and *P. protegens* Pf-5 were activated in 100 mL of LB Lennox medium in 250 mL glass shake flask by inoculating

glycerol stock cultures of the bacteria, and incubated at the following conditions: *E. coli* DH5α (37°C and 230 rpm rotational shaking), *B. subtilis* NRS-762 (30°C and 230 rpm rotational shaking), *P. protegens* Pf-5 (30°C and 230 rpm rotational shaking). After 24 hours of incubation, inoculating loops were used aseptically to inoculate experiment cultures on colourless agar medium via the streak plate technique. Incubation conditions for the different bacteria were as follows: *E. coli* DH5α (37°C), *B. subtilis* NRS-762 (30°C), *P. protegens* Pf-5 (30°C) in a temperature controlled incubator.

Profiling the microbial flora of deionized water

Deionized water was collected using a pre-sterilized 20 mL plastic bottle after allowing 5 minutes for flow through of deionized water collected in the deionized water production system. Thus, the deionized water used as inoculum was freshly produced. Spread plate technique was used in inoculating 0.1 mL of deionized water on formulated colourless agar and R2A agar. Agar plates were incubated at 30°C in a temperature controlled incubator and observed daily.

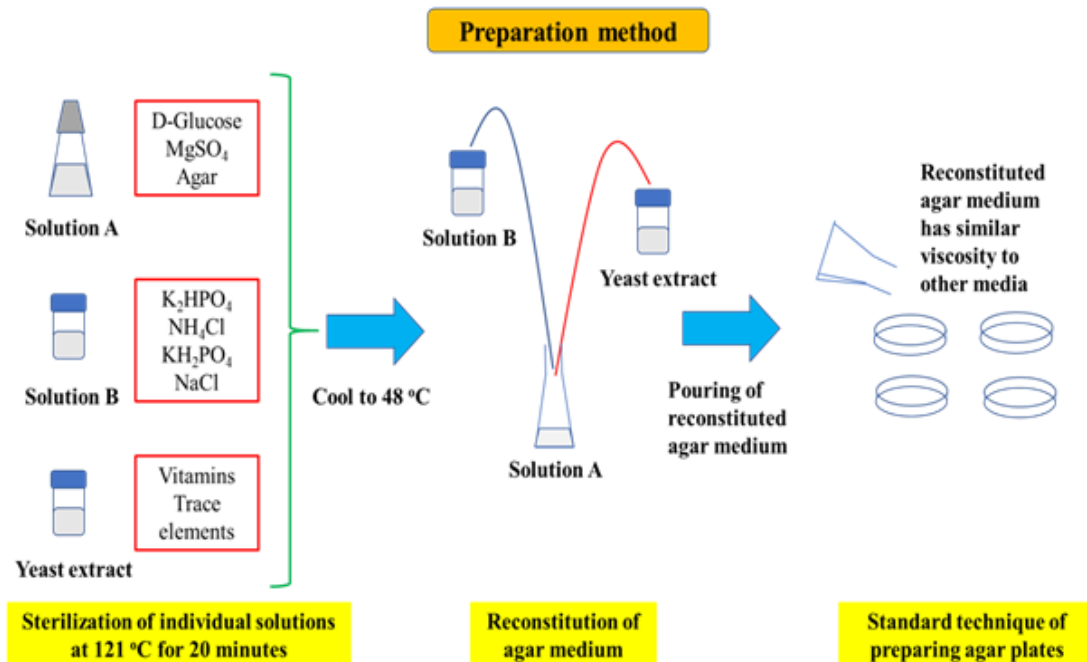


Figure 1: Method for preparing colourless agar. Specifically, the constituents of the agar are separated into three solutions that contain key elements of the agar. Solution A, for example, contains glucose, agar powder and $MgSO_4$, Solution B contains phosphate salts, ammonium chloride and sodium chloride, while Solution C is yeast extract which provides the trace elements and vitamins needed for growth.

Given that heat induced reaction between sugars and ammonium compounds would result in the formation of coloured compounds that lend a colour to the agar, glucose and ammonium compounds must be separately sterilized. Specifically, glucose could not be sterilized with ammonium chloride and yeast extract. However, it could be paired with agar powder in Solution A, which also contains $MgSO_4$ as the sulphur source. Solution B would be constituted by ammonium chloride as the nitrogen source of the colourless agar medium. This was combined with K_2HPO_4 , KH_2PO_4 and sodium chloride. Yeast extract is Solution C as it contains important vitamins and trace elements necessary for growth of microorganisms unable to grow in minimal salts medium. However, since yeast extract contains

sugars and ammonium compounds, it could not be sterilized with glucose or ammonium chloride, given the formation of coloured compounds when sugars react with ammonium compounds.

After separate sterilization, Solutions A, B and C were cooled to approximately $\sim 48^\circ C$ and subsequently mixed to reconstitute the colourless agar medium (Figure 1). The reconstituted solution was swirled repeatedly prior to pouring of agar plates using standard techniques. Viscosity of the colourless agar medium was similar to those of commercial agar media. Finally, yeast extract could be added up to a concentration of 1 g/L in the colourless agar medium formulation without affecting the optical transparency of the medium.

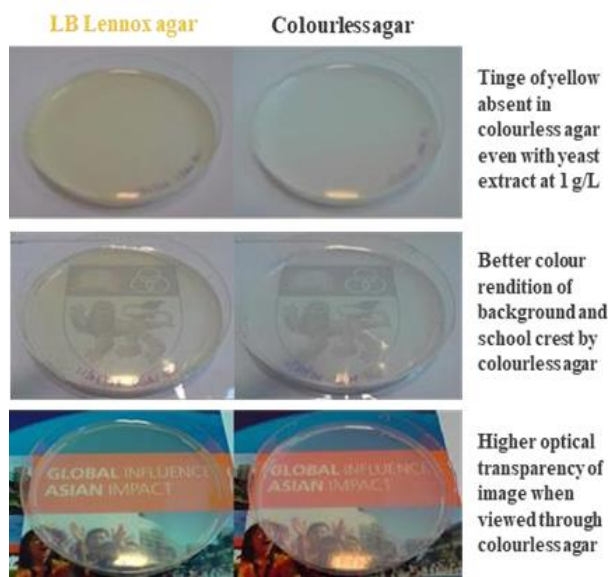


Figure 2: Colourless agar delivers higher optical transparency and colour rendition compared to a commercial LB Lennox agar, which is beige in colour

Compared to beige LB Lennox agar, the formulated colourless agar provided higher optical transparency and colour rendition (Figure 2). Specifically, tinge of yellow was absent in the colourless agar even if it was supplemented with yeast extract to a concentration of 1 g/L. Furthermore, colourless agar

was able to render a background image's colours better than a beige LB Lennox agar. Finally, greater optical transparency was apparent in images observed through the colourless agar compared to a beige LB Lennox agar.

Components	Concentration (g/L)
K ₂ HPO ₄	0.5
KH ₂ PO ₄	0.1
D-Glucose	2
NH ₄ Cl	0.5
Yeast extract	1
NaCl	0.5
MgSO ₄ .7H ₂ O	1
Agar	15

Table 1: Composition of formulated colourless agar

Composition of formulated colourless agar medium is shown in Table 1. To allow sufficient capacity for supporting the growth of myriad microbes, a glucose concentration of 2 g/L was used. Ammonium chloride concentration was designed to maintain a 1:1 concentration ratio with glucose on a molar basis. Concentration of yeast extract, which is important to provide a source of vitamins and growth factors for the growth of microbes unable to synthesize

vitamins, could be increased and decreased for understanding the effect of yeast extract in inducing the growth of different microbes. Buffering capacity of the agar as well as a source of potassium and phosphate comes from K₂HPO₄ and KH₂PO₄. MgSO₄.7H₂O, on the other hand, provides sulphate and magnesium for the microorganisms. Finally, NaCl adjusts the osmolarity of the medium.

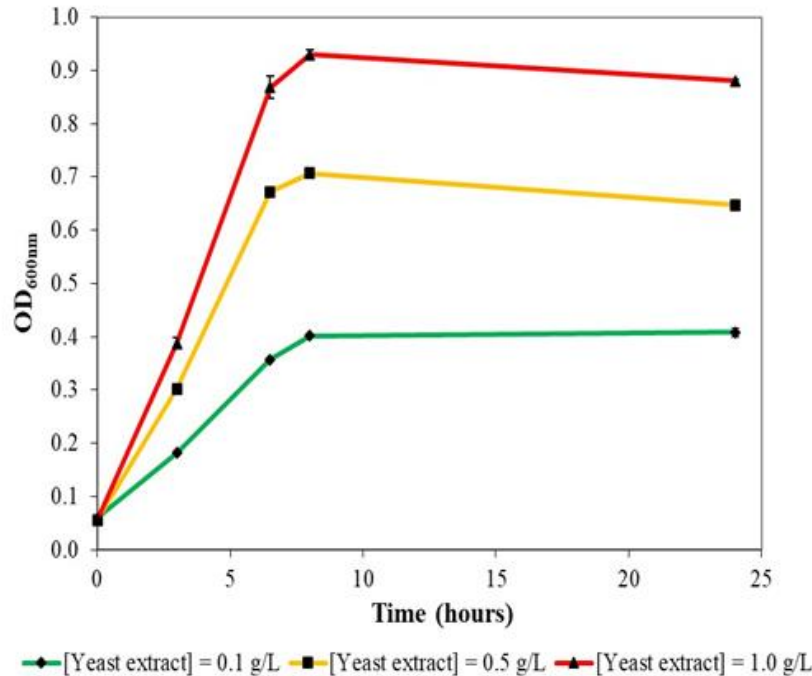


Figure 3a: Growth of *Escherichia coli* DH5 α at 37°C and 230 rpm rotational shaking in liquid version of formulated colourless agar with different concentrations of yeast extract supplementation. *E. coli* DH5 α could not grow in minimal salts medium without yeast extract supplementation.

Escherichia coli DH5 α could not grow in liquid version (without agar powder) of the colourless agar medium without yeast extract supplementation. With increasing concentration of yeast extract supplementation, *E. coli* DH5 α demonstrated better growth and biomass formation, with maximal optical density obtained of 0.4, 0.7 and 0.9 at 0.1 g/L, 0.5

g/L and 1.0 g/L yeast extract supplementation, respectively (Figure 3a). Growth rates of *E. coli* DH5 α also increased with yeast extract concentration in a concentration-dependent manner. In general, *E. coli* DH5 α grew reasonably well in yeast extract supplemented colourless agar medium with no drastic decline in optical density.

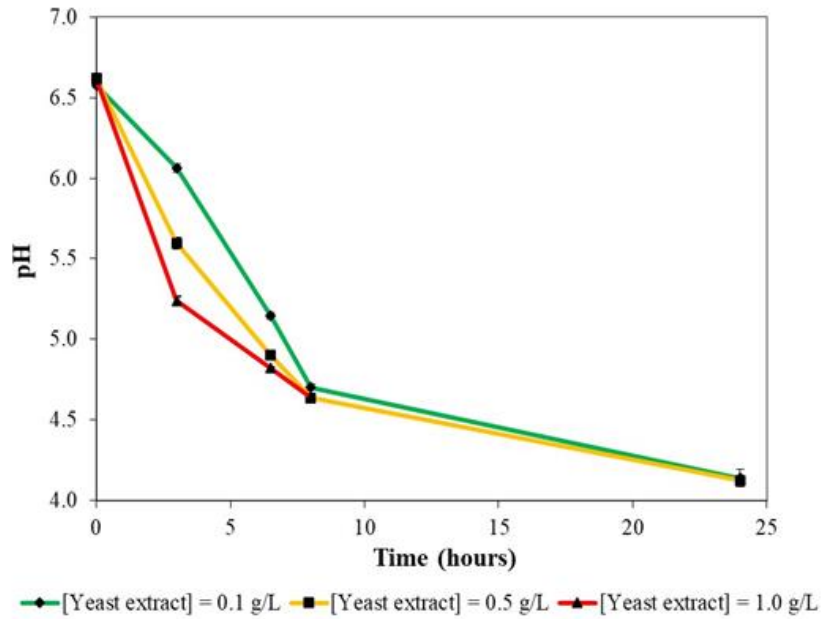


Figure 3b: Variation of pH during growth of *E. coli* DH5 α in liquid version of formulated colourless agar medium at 37°C and 230 rpm rotational shaking.

At the onset of growth in liquid version of colourless agar medium, *E. coli* DH5 α secreted significant amounts of acidic metabolites into the culture broth, which resulted in a decrease of pH from 6.6 to 4.6 at the start of stationary phase in the cultures (Figure 3b). Following which, pH declined further to 4.1 at the end of cultivation. What likely transpired was

rapid secretion of acidic metabolites in a yeast extract concentration-dependent manner where the higher the yeast extract supplementation, the greater would be the amount of acidic metabolites secreted. pH of culture broth continued to decrease during stationary phase.

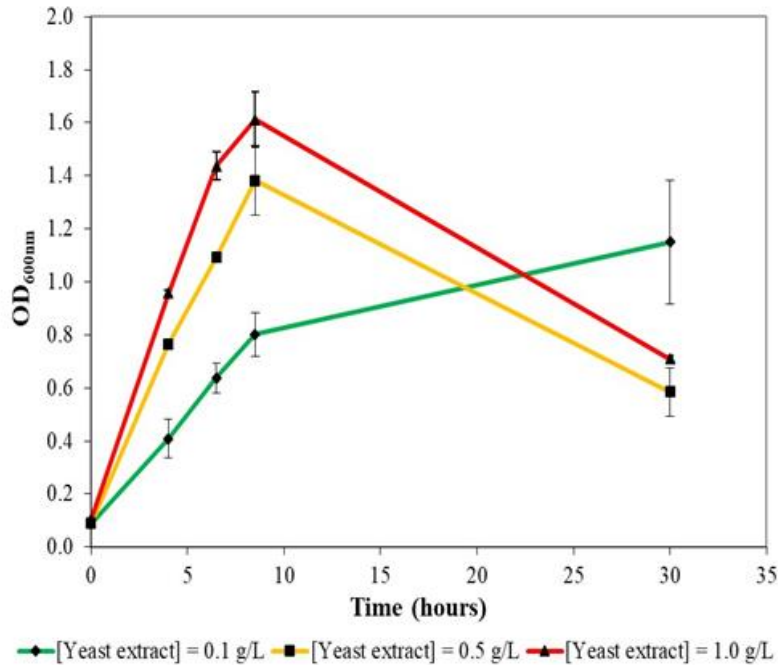


Figure 4a: Growth of *Bacillus subtilis* NRS-762 in liquid version of formulated colourless agar medium at different concentrations of yeast extract supplementation at 30°C and 230 rpm rotational shaking. Drastic decline in optical density after stationary phase likely resulted from cannibalism of non resistant cells. *B. subtilis* NRS-762 could not grow in minimal salts medium without yeast extract supplementation.

Similar to the case with *E. coli* DH5 α , *B. subtilis* NRS-762 exhibited higher growth rates with greater amount of yeast extract supplementation in the formulated colourless agar medium (Figure 4a). Specifically, maximal optical density obtained was 1.4 and 1.6 for yeast extract concentration of 0.5 and 1.0 g/L, respectively. On the other hand, growth of *B. subtilis* NRS-762 at low yeast extract supplementation of 0.1 g/L extended beyond 30 hours given the slow rate of cell growth. Drastic decline in optical density after onset of stationary

phase in cells cultivated in 0.5 and 1.0 g/L yeast extract supplementation was likely due to cannibalism rather than prophage induced cell lysis. Specifically, in delaying entry into the sporulation programme, subpopulations of *B. subtilis* NRS-762 likely secreted cell lysis factors that lysed non resistant cells for releasing cellular content needed for sustaining the remaining population. Prophage induced cell lysis, on the other hand, would have resulted in precipitous decline of the optical density rather than the gradual decline observed.

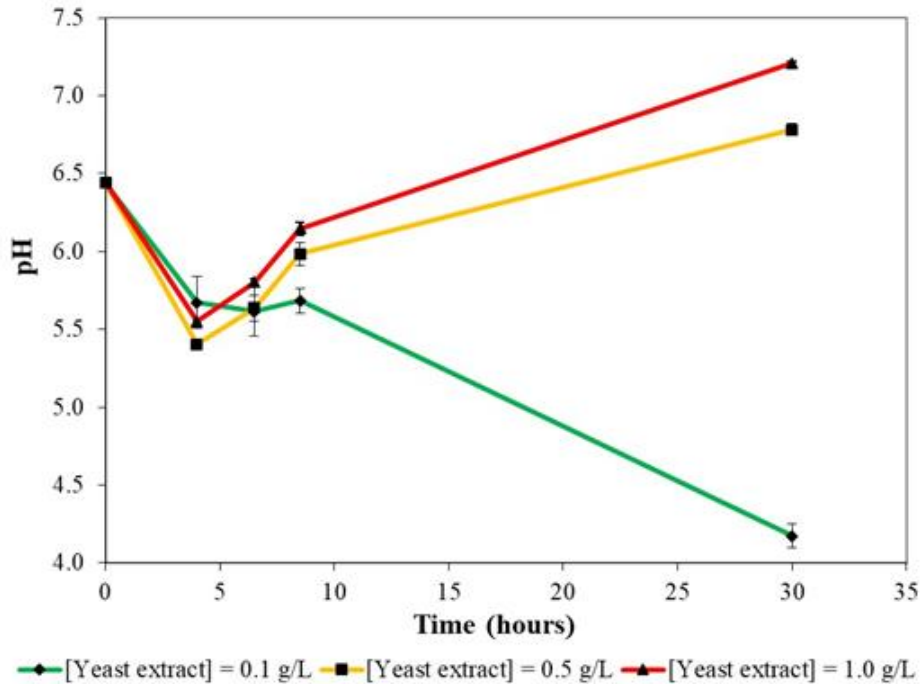


Figure 4b: Variation of pH during growth of *B. subtilis* NRS-762 in liquid version of formulated colourless agar medium with different concentrations of yeast extract supplementation at 30°C and 230 rpm rotational shaking.

Growth of *B. subtilis* NRS-762 in colourless agar medium with 0.1 g/L yeast extract supplementation might be using a different set of metabolic pathways compared to that in colourless agar medium with 0.5 and 1.0 g/L yeast extract, given the distinctly different pH profiles during growth (Figure 4b).

Specifically, in the case of growth in colourless agar medium with 0.5 and 1.0 g/L yeast extract, pH decreased followed by an increase. On the other hand, growth of the bacterium in colourless agar medium with 0.1 g/L of yeast extract exhibited a continuous decrease in pH.

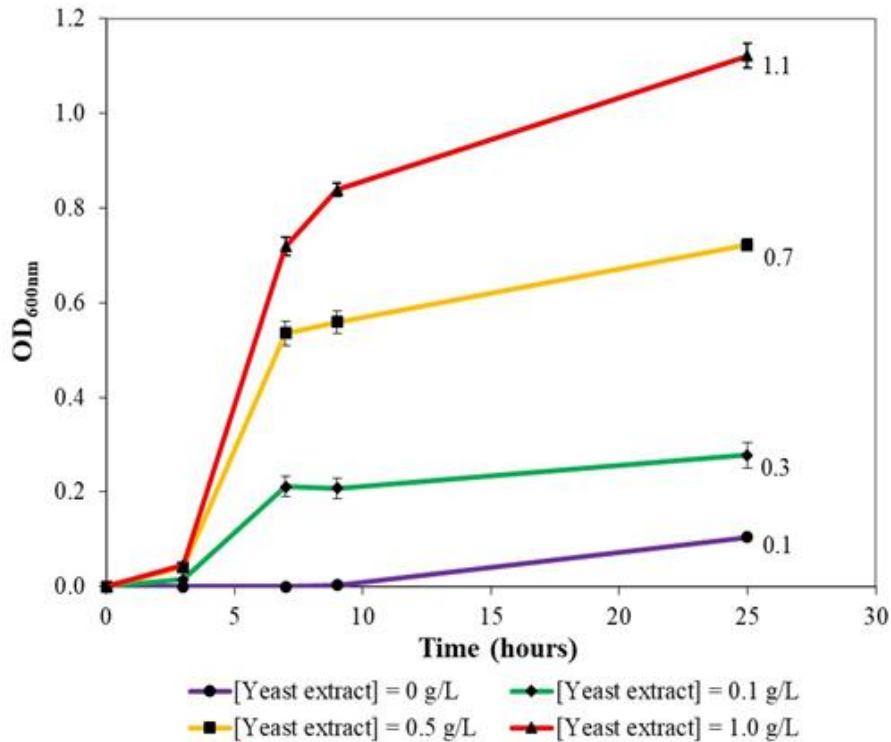


Figure 5a: Growth of *Pseudomonas protegens* Pf-5 in colourless agar medium supplemented with different concentrations of yeast extract at 30°C and 230 rpm rotational shaking. Note that growth rates and maximal optical density increased with yeast extract concentration in a concentration-dependent manner.

Growth rates and maximal optical density of *P. protegens* Pf-5 in formulated colourless agar medium positively correlated with amount of yeast extract supplementation. Specifically, optical density increased from 0.1 to 0.3 and 0.7 and finally 1.1 with increasing yeast extraction supplementation from 0, 0.1, 0.5 and 1.0 g/L, respectively (Figure 5a). Hence, given that yeast extract is a source of vitamins and growth factors, rates of cell division and biomass formation increased with increasing yeast extract supplementation as *P. protegens* Pf-5 cells do not need to expend metabolic energy from nutrients in synthesizing vitamins and other building blocks for growth. On the other hand, the bacterium could grow slowly in liquid version of colourless agar medium

without yeast extract supplementation where an intense green colour broth was observed. With increasing concentrations of yeast extract supplementation, the green colouration of the broth decreased (0.1 g/L yeast extract) and turned to beige coloured broth (at 0.5 and 1.0 g/L yeast extract). Overall, this likely suggested that secretion of a green colour siderophore by *P. protegens* Pf-5 was suppressed with increasing concentration of yeast extract since yeast extract was able to supply the iron necessary for growth, where in the case of no yeast extract supplementation, large amount of siderophore would need to be synthesized for sequestering iron needed for biomass formation.

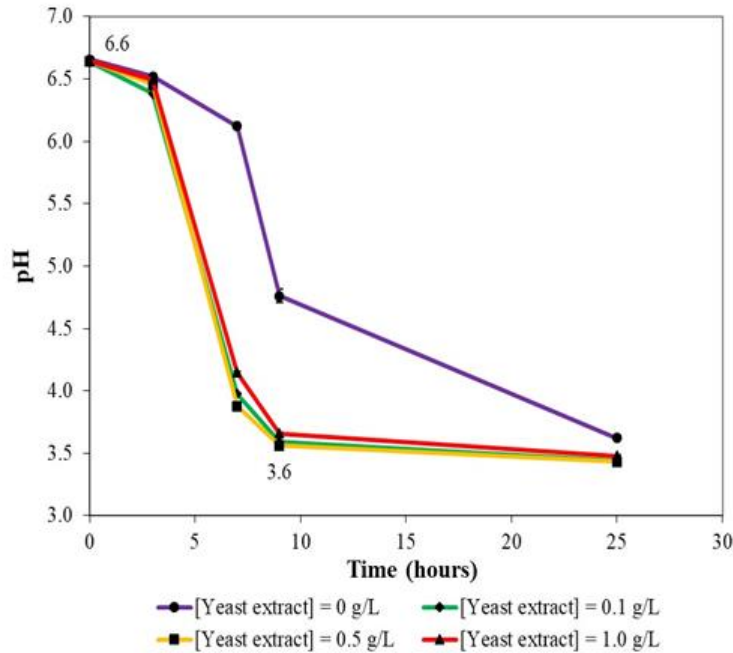


Figure 5b: Variation of pH during growth of *P. protegens* Pf-5 in liquid version of formulated colourless agar medium with different concentrations of yeast extract supplementation. Note that the bacterium likely utilized similar metabolic pathways for growth in colourless agar medium irrespective of amount of yeast extract supplementation (0.1, 0.5, and 1.0 g/L).

pH profile for *P. protegens* Pf-5 growth in liquid version of formulated colourless agar medium under different concentrations of yeast extract supplementation (0.1, 0.5 and 1.0 g/L) revealed that irrespective of the concentrations of yeast extract used, similar pH profile was obtained, which suggested that similar sets of metabolic pathways were used for the growth of *P. protegens* Pf-5 in the medium (Figure 5b). Specifically, pH decreased from 6.6 at the onset of growth to 3.6 after 9 hours of incubation. pH continued to decrease thereafter. On the other hand, a different pH profile was observed

for growth of *P. protegens* Pf-5 in liquid version of colourless agar medium without yeast extract supplementation, which suggested that the bacterium was likely using different sets of metabolic pathways for growth. Finally, pH profile studies revealed that *P. protegens* Pf-5 could grow at a pH of 3.6; for example, Figure 5a clearly indicated that optical density continued to increase for the bacterium after 9 hours of incubation in liquid versions of formulated colourless agar medium with 0.5 and 1.0 g/L yeast extract supplementation.

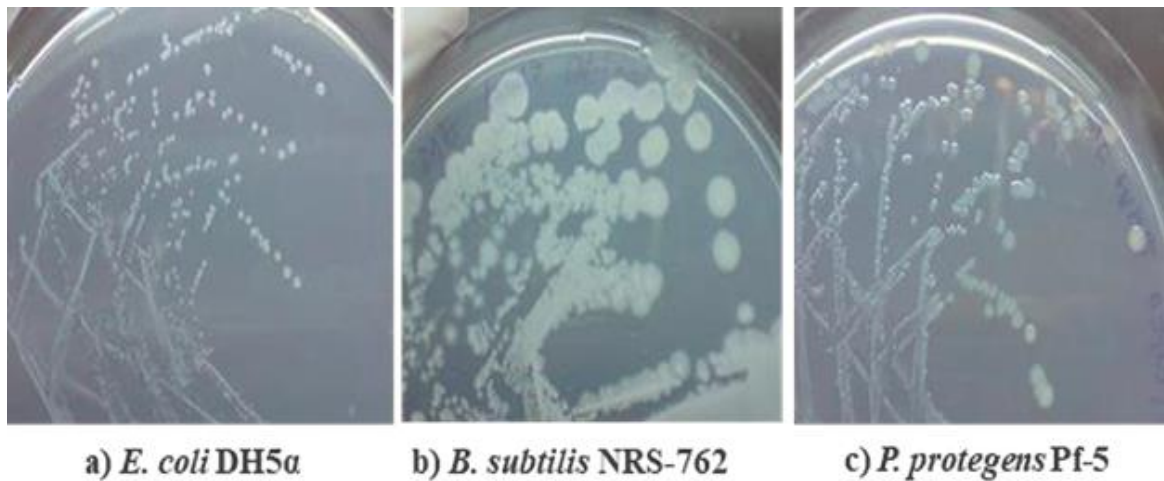


Figure 6: Growth of *E. coli* DH5α, *B. subtilis* NRS-762 and *P. protegens* Pf-5 in solid formulated colourless agar medium after streak plate inoculation.

Good growth of *E. coli* DH5α, *B. subtilis* NRS-762 and *P. protegens* Pf-5 on solid formulated colourless agar medium was observed with colony morphologies similar to those obtained during growth on LB Lennox agar medium, which suggested that growth inhibitory compounds were not produced during agar preparation (Figure 6). Specifically, small, round, white colonies were obtained for *E. coli* DH5α growth on colourless agar medium, which was similar to the morphology obtained during growth on LB Lennox agar. However, colonies of *E. coli* DH5α on colourless agar medium lacked the beige colour typical of the bacterium's colonies on LB Lennox agar. In the case of *B. subtilis* NRS-762, white,

wrinkled, round colonies were obtained during growth on colourless agar medium, which compared well with the beige, round and wrinkled colonies on LB Lennox agar. But, the mechanisms underlying the loss of pigmentation of *E. coli* DH5α and *B. subtilis* NRS-762 colonies during growth on formulated colourless agar remain unknown. Finally, *P. protegens* Pf-5 colonies on colourless agar medium were greenish, small and round, which were similar to those obtained on LB Lennox agar. However, there was no secretion of green siderophore by *P. protegens* Pf-5 during growth on colourless agar medium with yeast extract supplementation at 1 g/L.

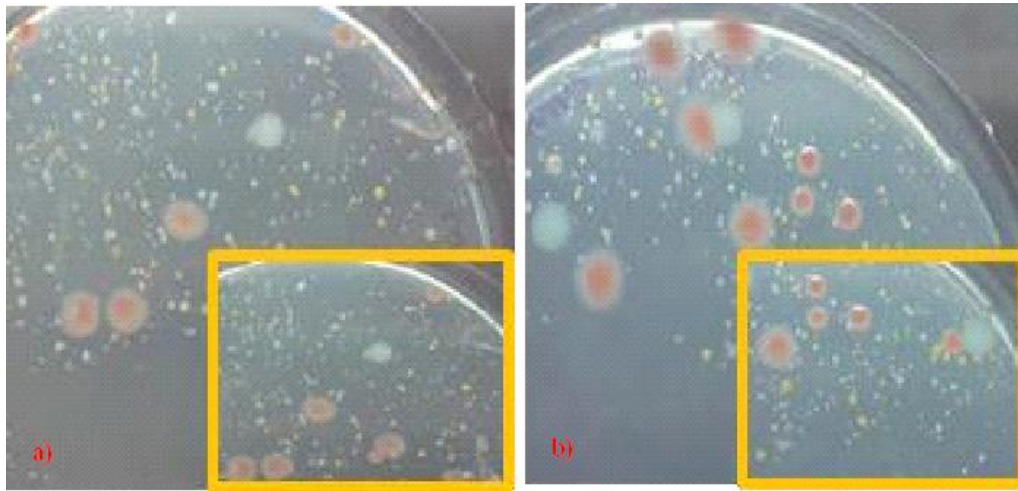


Figure 7: Growth of microbial flora in deionized water on a) R2A agar, and b) formulated colourless agar incubated at 30°C. Insets showed a close-up of the original photograph. Note that many types of microbial species were present in deionized water, and colourless agar was able to support the growth of many of the species; thereby, illustrating that growth inhibitory compounds were not generated during agar preparation.

For assessing whether the formulated colourless agar was able to support the growth of various types of microbes present in the environment as well as the presence of growth inhibitory compounds generated during agar preparation, spread plate culture of deionized water inoculum was performed on both the gold standard R2A agar and the formulated colourless agar medium. Many different types of microbial colonies of different morphologies and pigmentation were recovered on both types of agar, which confirmed the presence of a large variety of microorganisms in deionized water (Figure 7). Specifically, the ability of formulated colourless agar medium in recovering a large variety of different types of microbial colonies attested to the fact that the formulated colourless agar medium was able to support the growth of many microbial species. More importantly, the result also showed that growth inhibitory compounds were not generated during agar preparation. Overall, the high optical transparency of the formulated colourless agar medium enabled greater ease in identifying small colonies recovered from the deionized water inoculum. Great diversity in

microbes present in deionized water pointed to the important role that monochloramine played in suppressing microbial growth in drinking water distribution system and the detrimental impact to microbiological quality of water upon its removal by the ion exchange system that produced deionized water.

Conclusions

By separately sterilizing sugars and ammonium compounds during agar preparation and reconstituting the different solutions at ~48°C, a method for preparing colourless agar was developed. The colourless agar improved colour contrast between microbial colonies and agar background. More importantly, the formulated colourless agar was amenable to supplementation with yeast extract (up to a concentration of 1 g/L) without introduction of colour. Growth experiments with *Escherichia coli* DH5α, *Bacillus subtilis* NRS-762 and *Pseudomonas protegens* Pf-5 in liquid versions of the colourless agar (without agar powder) confirmed that the medium could support the growth of common

bacteria to an optical density of ~1.0 in an otherwise low nutrient content medium. On the other hand, streak plate culture of the same bacteria on solid formulated colourless agar medium recovered growth of the microbes in morphologies similar to those obtained on LB Lennox agar. Loss of pigmentation was an issue with growth of *E. coli* DH5 α and *B. subtilis* NRS-762 on colourless agar but not *P. protegens* Pf-5. Specifically, whitish, round colonies were obtained for *E. coli* DH5 α and *B. subtilis* NRS-762 instead of beige colonies. Culture experiments aimed at assessing the ability of the colourless agar medium in supporting the growth of a variety of microbes confirmed that the medium could recover many types of microbial species from deionized water, which highlighted that the medium was suitable for use in microbial ecology studies aimed at assessing the microbial flora of a habitat. More importantly, the result indicated that no growth inhibitory compounds were generated during agar preparation. Collectively, the formulated colourless agar medium could find use in a wide variety of applications in microbiology ranging from microbial ecology studies to cell counting experiments.

Supplementary materials

Additional information on the growth of different bacteria on formulated colourless agar can be found in the appended supplementary materials.

Conflicts of interest

The author declares no conflicts of interest.

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