

JOURNAL OF BIOTECHNOLOGY AND BIOMEDICINE

Research Article

ISSN: 2642-9128



Hoangvi Le, Vikrant Rai, Devendra K Agrawal*

Abstract

Since cholesterol is not routinely measured in astronauts before and after their return from space, there is no data on the role of blood cholesterol level in muscle atrophy and microgravity. Since the first moon landing, aerospace medicine became outdated and has not pushed boundaries like its rocket engineering counterpart. Since the 2019 astronaut twin study, there has yet to be another scientific breakthrough for aerospace medicine. Microgravity-induced muscle atrophy is the most known consequence of spaceflight. Yet, so far, there is no therapeutic solution to prevent it or any real efforts in understanding it on a cellular or molecular level. The most obvious reason to this unprecedented level of research is due to the small cohort of astronauts. With the establishment of private space industries and exponential recruitment of astronauts, there is more reason to push forward spaceflight-related health guidelines and ensure the safety of the brave humans who risk their lives for the progression of mankind. Spaceflight is considered the most challenging job and the failure to prevent injury or harm should be considered reckless negligence by the institutions that actively prevented sophistication of aerospace medicine. In this critical review, role of cholesterol is analyzed across the NASAestablished parameters of microgravity-induced muscle atrophy with a focus on potential therapeutic targets for research.

Keywords: Astronauts; Cholesterol; Hypercholesterolemia; Microgravity; Muscle atrophy; Space flight

Introduction

Microgravity induced skeletal muscle atrophy is well established in aerospace medicine. In 1999, it was first demonstrated that skeletal muscle cells were directly affected by space travel [1, 2]. However, the mechanism of muscle atrophy in astronauts lacks foundation other than microgravity-induced unloading. Skeletal muscle atrophy is characterized by reduced muscle fiber size and wasting. Skeletal muscle wasting is a predominate indicator of terminal illness because skeletal muscle is a homeostatic-sensitive organ. Any slight shift in balance causes skeletal muscle to dramatically reduce itself to maintain homeostasis. Currently, medical experiments on the International Space Station (ISS) have not revealed any substantial leads in how to prevent muscle atrophy or have investigated other reasons for muscle atrophy other than microgravity. This is described in one of NASA's human research gaps M23: "Determine if factors other than unloading contribute to muscle atrophy during space flight" and lists potential risk factors as "inflammation, redox balance, energy balance, hydration, etc." [3]. The potential risk factors in the NASA list are all associated with cholesterol and will be the focus of discussion in this article. Cholesterol is a fundamental and modifiable biochemical component in phospholipid bilayer. Hypercholesterolemia



Affiliation:

Department of Translational Research, Western University of Health Sciences, Pomona, California 91766, USA

*Corresponding author:

Devendra K Agrawal, Department of Translational Research, Western University of Health Sciences, Pomona, California 91766, USA.

Citation: Hoangvi Le, Vikrant Rai, Devendra K. Agrawal. Cholesterol: An Important Determinant of Muscle Atrophy in Astronauts. Journal of Biotechnology and Biomedicine 6 (2023): 67-79.

Received: February 08, 2023 **Accepted:** February 24, 2023 **Published:** March 02, 2023



(HC) is characterized by high cholesterol in the blood with increased levels of low-density lipoproteins (LDLs) and low levels of high-density lipoproteins (HDLs). One study has demonstrated significant skeletal muscle fiber wasting as a result of ApoE inactivation and increased non-HDL cholesterol in LGMD2B mice. It was postulated that damaged vascular barriers allow the chronic leakage of plasma lipids into muscle tissue, resulting in muscle inflammation [4]. Microgravity shortened the body length and increased fat accumulation in microgravity-cultured worms compared to normal gravity cultured worms [5]. Impaired lipolysis in adipose tissues and skeletal muscles due to reduced mRNA expression levels of lipoprotein lipase (LPL) in adipose tissue and reduced LPL activity in skeletal muscle [6]. Impaired function of LPL in skeletal muscle mediates HC which ultimately leads to increased LDLs and muscle atrophy due to decreased blood supply caused by lumen obstruction [7]. This notion is supported by the fact that lower skeletal muscle mass index is associated with dyslipidemia and increased muscle mass is a must to prevent hypercholesterolemia [8]. These results suggest an association of skeletal muscle weakness with hypercholesterolemia and increased muscle mass and lowering LDLs as a therapeutic strategy to prevent muscle atrophy. Cholesterol (in the context of space) has become a significant health marker in heart disease, which NASA also aims to mitigate in astronauts [9]. However, the current collection of space-related medical research on cholesterol is insufficient and does not reveal much about the effect of microgravity on cholesterol. In fact, cholesterol is not a routine measurement for astronauts when landing [10]. According to NASA, hypercholesterolemia, obesity, diabetes, and hypertension could develop in astronauts [11]. Growing interest in hypercholesterolemia-induced muscle atrophy may provide novel insight to protect astronauts from such health risks [4, 12]. This review aims to critically evaluate the cholesterol-related microgravity research and emphasizes the detrimental consequences of undervalued cholesterol as a target in treating muscle atrophy in astronauts.

NASA-defined Spaceflight Health Risks

It is important to recognize that astronauts perform tasks inside and outside the International Space Station (ISS), and few researchers have considered that not all astronauts are tasked with spacewalks. Astronauts who participate in spacewalks could be potential outliers in medical experiments because of the physical impact from the severely outdated spacesuit. According to the audit of April 2017, only 11 of the original extravehicular mobility unit (EMU) spacesuits are still in use, 4 of which are available on the ISS and the rest are on Earth for maintenance [13]. This is prompted by the need to develop a next-generation spacesuit to address the limitations of the current spacesuit. In August 2021, NASA Office of Inspector General performed another audit to examine the current progress of the next-

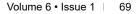
generation spacesuits for the ISS and Artemis missions [14]. This audit also revealed the health-related risks associated with the current spacesuit and examples of negative health outcome from specific missions. Spacesuit-related risks include decompression sickness, thermal regulation, shoulder injuries, hand injuries, malnutrition, and dehydration. In addition to this, astronauts are not completely protected by the current spacesuit from cosmic radiation during spacewalks or on the Moon. Compared to activities done safely inside the ISS, spacewalks are considered one of the most dangerous jobs in mankind and can certainly contribute to the physical and psychological stress of astronauts. However, similar health risks are associated within the ISS as described in Appendix B of NASA's November 2021 audit [IG-22-005] [15]. Some notable mentions are injury due to the operations of extravehicular activity (EVA), altered immune response, inadequate food and nutrition, spaceflight-induced cardiovascular disease (CVD), and space radiation exposure. The same audit highlights how mitigation of human health risks requiring ISS microgravity testing will not be complete by 2030, the retirement date of the ISS, and 8 out the 12 critical human health risks will not be mitigated at an acceptable level for long-term spaceflight [15]. Although this schedule is based on an approximation, it can be delayed even further if research is not productive. This warrants critical evaluation of the current aerospace research efforts and what shortcomings they share to pinpoint the reason behind the slow progress.

Role of Cholesterol in Space Health Risks

Microgravity is undoubtedly a main contender for muscle atrophy, but research in microgravity has an astronomical amount of confounding variables, leaving inconsistent and often uninterpretable results. The main concerns NASA described in their audits have one common variable: cholesterol. According to the history, cholesterol was first described as a perpetrator of heart disease in 1968, only 7 years after the first manned spaceflight in 1961 and around the same time as the Apollo moon landing in 1969. Whether or not the first biomedical researchers of NASA have considered these factors and how far to investigate them is up to debate. Since the discovery on the role of cholesterol role in heart disease, the recommended consumption of cholesterol is less than 300mg/d for astronauts, which has not changed since. The behavior of cholesterol in microgravity was demonstrated in a basic study where simulated microgravity enhanced lipid accumulation along imitation-vessels due to random particle direction [16]. In an environment of potent solar radiation, oxidation of cholesterol may exacerbate the health of astronauts. In each relevant health risk, cholesterol will be described in context of clinical scenarios that mimic health outcomes in microgravity (Figure 1).

Cholesterol and Decompression Sickness

Decompression sickness occurs when an astronaut's





Hoangvi Le, et al., J Biotechnol Biomed 2023 DOI:10.26502/jbb.2642-91280072

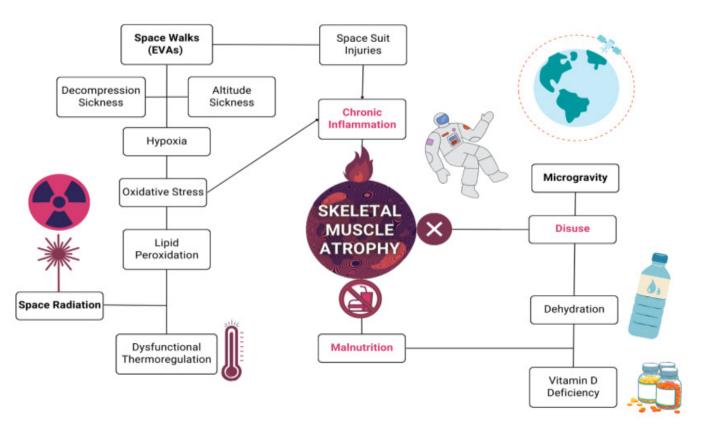


Figure 1: Spaceflight health risks that lead to skeletal muscle atrophy in astronauts. The terms highlighted in red are the known direct causes of muscle atrophy, which are disuse, malnutrition, and chronic inflammation. The bolded text represents the unique circumstances of spaceflight that astronauts are inevitably exposed. These include space walks, aka extravehicular activities (EVAs), space radiation, and microgravity. As illustrated, there are many intersecting confounding variables that may contribute to spaceflight-induced skeletal muscle atrophy. However, the one confounding variable that has yet to be considered is cholesterol, as it plays a role in almost all these clinical scenarios.

body experiences sudden drop in surrounding pressure, such as their spacesuit. Decompression sickness is associated with terrestrial activities such as deep sea diving and nonpressurized aerial flying [17]. Hypercholesterolemia after high-fat meal increases risk for decompression sickness in divers [18]. Astronauts tend to eat before spacewalk to avoid eating in spacesuit. If preparing for an 8-hour spacewalk, it can be assumed that they are eating enough to sustain themselves for that period. Binge eating beforehand can spike serum cholesterol concentrations of an astronaut right before preparing for a spacewalk, as seen in diving. Decompression sickness is associated with oxygen toxicity because deep-sea divers and astronauts both breathe pure oxygen. However, it has long been known that concentrations of oxygen greater than normal breathing air has slow but detrimental effects. One of these effects include oxidative stress-induced inflammation and lipid peroxidation [19]. On the other hand, divers or astronauts returning from breathing pure oxygen results in decreased blood oxygen levels and results in hypoxia. This is also associated with high altitude sickness where there is a drop in oxygen pressure [20]. Acclimatization to high altitudes show significant positive correlation with increasing serum cholesterol [21]. High-altitude induced hypoxia are

directly associated with increased non-HDL cholesterol [22]. Hypoxia can lead to chronic inflammation and oxidative stress, another common characteristic in astronauts, and will be discussed further in the following sections [23, 24].

Cholesterol and Thermoregulation

Astronauts have a water-cooling system in their spacesuits to help regulate their increased temperature during ISS spacewalks. Thermal regulation is challenging because astronauts may spend up to 8 hours in their spacesuits in direct solar radiation while also performing tasks. Cholesterol is a fundamental component of animal cell membrane homeostasis that can act as a buffer molecule to stabilize the phospholipid bilayer over a range of temperatures. Cholesterol lowers membrane fluidity in high temperatures and increases membrane fluidity in lower temperatures [25]. Astronauts experience increased core body temperature and are at risk for heat stress and hyperthermia [26]. On earth, increased ambient temperature is associated with increased levels of LDL and decreased levels in HDL [27]. Heat stress from a heat wave has shown to increase plasma cholesterol in a British population [28]. Hyperthermia is correlated with increased cellular cholesterol contents studied in mammalian



cell lines [29]. Long-term heat exposure influenced cholesterol metabolism in pigs and temporarily increased serum and LDL cholesterol levels. However, inflammation or tissue damage was not present [30]. In hibernating brown bears, dyslipidemia and muscles were protected against lipid-specific oxidative damage due to higher plasma-antioxidants reserves [31]. Increased temperature may only be relevant to current space activities, whereas decreased temperatures on the Moon or Mars may pose a new threat in space exploration.

Cholesterol and Spacesuit Injuries

Extravehicular activity (EVA) injuries of hands and shoulders are usually sustained with the EMU spacesuit during spacewalks and EVA training. In zero gravity, astronauts tend to use their upper extremities for movement and physical tasks, versus their lower extremities, resulting in the overuse of shoulders and hands. The spacesuit hardware of the upper torso was the main cause of injury during active duty [32]. Although, shoulder injury has only increased recently in NASA perhaps due to changes in the space suit design or changes in the spaceflight requirements, such as all astronauts must be EVA certified in order to fly starting in 2000 [33]. Astronauts selected in the 1990's have higher incidences in shoulder surgery. Astronauts who have performed more than five spacewalks were twice as likely to sustain shoulder injuries than astronauts who performed one spacewalk [34]. Shoulder injury is a broad term that encompasses many different types of myopathies or tendinopathies relating to the shoulder rotator cuff. Rotator cuff injury could manifest from hypercholesterolemia due to higher levels of total cholesterol, triglycerides, and LDL cholesterol [35]. Dyslipidemia and lipid deposition is associated with the failure of rotator cuff repairs and increased risk of retear [36]. NASA is fully aware of the detrimental effects of the spacesuit to cause shoulder injury in astronauts and have opened this issue for public collaboration. Texas Women's University and Wichita State University are currently in the works for a mechanical sensor detection system to avoid shoulder injury [37, 38]. On the other hand, astronauts also experience handgrip fatigue due to pressurized gloves and limited mobility [39]. Those with smaller hands are more likely to sustain hand injuries, such as blisters, cuts, or joint pain [13]. From 1993 to 2010, NASA reports that 76 percent of astronauts sustained injuries of the fingernail, finger crotch, metacarpophalangeal joint, or fingertip [40]. Cholesterol deposits were found in extensor tendons of the hands in hypercholesterolemia patients [41, 42]. This could possibly increase the risk of astronauts developing hypercholesterolemia-induced arthritis [43]. Efforts are underway to improve pressurized gloves to mitigate hand injury, such as the next-generation high performance EVA gloves and the second-generation Space Suit Robot Glove (SSRG) [44, 45].

Cholesterol and Spaceflight Malnutrition

As mentioned before, astronauts may spend several hours performing an EVA without eating or drinking fluids. Single food bars were previously installed in EMU spacesuits, but were discontinued due to inconvenience (low calorie, extra weight, smearing on visor, limited storage space, etc.) [13, 46]. Astronauts prefer to eat a solid nutrient-dense meal before an EVA to remain satiated for as long as possible [47]. It is not clear if all astronauts are eating enough calories for their spacewalk or if astronauts over-eat to mitigate potential fatigue and hunger caused by the physical demands of an EVA. The food prepared for astronauts are heavily processed (freeze-dried) to meet sanitary and package requirements for long-term storage. Although fresh foods are more nutritional, they are prohibited due to shorter shelf-life and potential microbial contamination. Freeze-dried food and menu fatigue contribute to decreased appetite in astronauts [48]. Underconsumption of required nutritional intake may increase the risk of malnutrition in astronaut and subsequently increase oxidative stress in muscles. One study demonstrated that depriving nutrients in C2C12 muscle cells significantly increased the levels of reactive oxygen species (ROS) [49]. Food prepared in the ISS are higher in cholesterol (300 mg as of year 2020) than the standard recommendation of 200 mg [50]. If astronauts eat big meals, this may suddenly cause serum cholesterol to spike, such as consuming high amounts of sugar or eggs [51, 52]. It would be beneficial to study the body metabolism of astronauts before and after an EVA to investigate metabolic efficiency. Because most seasoned astronauts are middled aged, they are at higher risk for cardiovascular disease. Age-related metabolic alterations may affect how nutritional intake impacts endocrine functions, muscle mass homeostasis, and lipid profile [53]. Aging is correlated with dyslipidemia and cardiovascular disease [54, 55]. Aging also impacts muscle mass through sarcopenia and chronic inflammation [56, 57]. Striated muscle atrophy has been observed in cardiovascular disease [58]. Cardiovascular disease risk may be due to hypercholesterolemia. High LDLcholesterol has been associated with cardiovascular mortality and lowering LDL-C levels are beneficial for preventing cardiovascular disease in men [59, 60]. Moreover, high levels of oxidized-low density lipoprotein (ox-LDL) exacerbate the progression of atherosclerosis [61]. Although NASA aims to mitigate cardiovascular disease in astronauts, it is concerning that cholesterol is not routinely measured, which leads to the lack of cholesterol-specific research [10].

On the other spectrum, decreased vitamin D was the most striking nutritional changes during spaceflight. Vitamin D shares the same precursor as cholesterol, 7-dehydrocholesterol (7-DHC) [62]. UV light is completely shielded on the ISS and spacesuits have built in UV-blocking material. Therefore astronauts are unable to endogenously synthesize vitamin D for long period of time. Chronic vitamin D deficiency is a



critical concern for long-term space exploration because of the decreased calcium absorption and subsequent bone wasting [63]. Dyslipidemia, increased total cholesterol (TC), low-density lipoprotein cholesterol (LDL-C), decreased in high-density lipoprotein cholesterol (HDL-C) levels can manifest from vitamin D deficiency as well [64]. Astronauts also experience prolonged states of dehydration both in the ISS and during EVA spacewalks. Dehydration occurs after microgravity-induced fluid shift during spaceflight [65]. The recommended daily intake of water for astronauts is eight ounces per hour, but the disposable in-suit drink bag only contains 32 ounces of water, only sufficient for 4 hour spacewalks [66]. The average period of an EVA spacewalk in 2021-2022 is about 7-8 hours, which suggests that one drink bag is not enough for the entire mission [67]. Dehydration can lead to hypertension, muscle fatigue, and dizziness [68-70]. Cholesterol and hydration play a critical role in cell membrane fluidity and are directly correlated with each other [71]. This is demonstrated by increased serum cholesterol concentration in blood tests in dehydrated patients [72]. Therefore, it begs the question of whether the reported cholesterol levels in all previous human astronaut research is consistent or not.

Cholesterol and Space Radiation

In addition to UV radiation, astronauts experience high exposure to space radiation. Space radiation, or cosmic radiation, consists of high-charged energy particles, X-rays, and gamma rays. It is known that lipid peroxidation produced from space radiation [73]. LDL-cholesterol in particular is a vulnerable target of radiation-mediated oxidation [74]. Six hour exposure to UV radiation causes cholesterol to increase by 21.6-fold and direct cell membrane modification [75]. The effects of long-term exposure to dangerous levels of gamma radiation were significantly associated with hypercholesterolemia in Japanese atomic bomb survivors [76, 77]. For these reasons, cardiovascular disease and cancer are negative outcomes of space radiation that need to be mitigated before considering deep-space exploration [78].

Cholesterol and Space-induced Altered Immune Response

Astronauts experienced allergy-like symptoms, such as prolonged congestion, rhinitis, sneezing, and skin rashes, while traveling in space for long periods of time, which prompted investigation of how microgravity affects immune response [79]. Chronic inflammation was observed in astronauts due to many confounding factors, such as oxidative stress, but the mechanism behind microgravity-induced immune hyperactivity is not clear [80]. Since oxidative stress is a major contender of space risk, oxidation of LDL-cholesterol should not be ignored as a predictor of chronic inflammation. Hypercholesterolemia is often characterized by chronic inflammation response to LDL-cholesterol dominated macrophages, or foam cells [81]. Macrophages studies in microgravity are very diverse, making it more difficult to come to a significant conclusion [82]. Hyperlipidemia enhances neutrophil activity and are also elevated during microgravity as a response to oxidative stress [83, 84]. Response to dendritic cells diminished in microgravity as demonstrated by the suppression of T-cell reactivity [85, 86]. Microgravity studies have also shown hypoxia-inducible factor 1α (HIF- 1α) as a potential therapeutic target for its role in macrophage and T-cell activation [87].

Cholesterol and Muscle Atrophy in Astronauts

As mentioned above, NASA lists potentially causative factors for space-induced skeletal muscle atrophy, including inflammation, redox balance, energy balance, hydration, and others [3]. Evidence provided so far support the role of cholesterol in all these potential causes/factors of muscle atrophy. Understanding cholesterol on its very basic level is extremely important to its homeostatic functions in cell anatomy. Too much cholesterol can cause negative effects on important skeletal muscle membrane-protein species, such as those found in the transverse tubule (t-tubule). The t-tubule is considerably more cholesterol-rich than the sarcolemma and increasing this level further impedes the trafficking of intracellular glucose transporter, GLUT4, to the t-tubule and other surface membranes [88, 89]. Increased t-tubule cholesterol concentration may also may negatively alter voltage-gated Ca²⁺ channels [90]. In microgravity (where cholesterol may increase through many variables), astronauts may experience diabetic symptoms and dysfunctional metabolism, rendering carbohydrate/glucose intake as potential waste. Depletion of this cholesterol impairs excitation and contraction coupling. [91] In addition, high cholesterol causes inhibition of ATPases by overloading cholesterol in the striated muscle sarcolemma [92, 93]. Cholesterol depletion induced by statin drugs, used by some astronauts, may exacerbate muscle atrophy [94]. Astronauts who need rotator cuff surgery may be at risk for poor repair outcomes due to fatty infiltration and rotator cuff muscle atrophy [95]. Re-analysis of the twin study landing data show that sharp increase of cytokines and chemokines, and other inflammatory markers, such as IL-6, IL-10, IL-1β, IL-1Ra, CRP, CCL2, and TNF- α , suggest regenerative response to atrophy rather than inflammatory response [96]. In fact, chronic inflammation can disrupt proteins in the skeletal muscle fiber triggering atrophy [97]. It was demonstrated in rats flown in space for 12.5 days that muscle atrophy occurred and it was characterized by the dysfunctional microcirculation, denervation, infiltration and phagocytosis of cellular debris by macrophages and neutrophils in necrotized skeletal muscle fibers [98]. Chronic inflammation in skeletal muscle is dependent on macrophage kinetics and disturbance in cell signaling can lead to muscle fiber degeneration [99]. In chronic inflammation, both M1 and M2 macrophages



Volume 6 • Issue 1 | 72

increase and compete for arginine metabolism as it is a shared substrate for iNOS and arginase. However, M2c macrophages reduce the activity of M1 macrophages, leading to a shift in metabolism from iNOS to arginase, resulting in a pro-fibrotic environment. M2 macrophages also increase myogenic factors, such as MYOG and MYOD [100].

Inflammation-induced striated muscle atrophy has been observed in the dysregulation of muscle fibers such as actin, myosin, and titin [101-103]. Actin fragments were generated by caspase-3 and further degraded by a ubiquitin-proteasome [104]. Muscle-specific E3 ubiquitin-ligase, TRIM63 (or MuRF1), has been shown to degrade myosin light chains 1 and 2, myosin heavy chains, and myosin-binding protein C [105]. TRIM63 has also been observed in titin degradation in cardiomyopathies [106]. In hindlimb unloading studies, proteolytic titin fragments increased significantly after 7-days of gravitational unloading [107]. Current spaceflightrelated muscle research suggests that studies should focus on catabolic state markers, such as FOXO1 and muscle-specific TRIM63 (MuRF1) [108]. However, a recent transcriptome study done in 2021 has shown that deletion of MuRF1 in mice did not prevent muscle atrophy during spaceflight and suggested Cacng1 as a new target for microgravity-induced atrophy. The study also demonstrated that transfecting myotubes with active mutant of FOXO3, FOXO3a, decreased average myotube diameter by 27.5% [109, 110]. Mitochondrial function and mitochondrial DNA (mtDNA) can also be another relevant target for microgravitymediated muscle atrophy caused by oxidative stress, DNA damage, and inflammation [111, 112]. Reactive oxygen species (ROS) can activate NF-kB pathways and subsequent release of cytokines like TNF-a, as well as damaging the sarcolemma and contractile proteins, exacerbating muscle dysfunction in dystrophic muscle cells[49]. The plasticity of skeletal muscle is often challenged by hypercholesterolemia and dyslipidemia. Skeletal muscle can fluctuate between hypertrophy, as seen in bodybuilders, and atrophy, as seen with disuse. Lipotoxicity can trigger inflammation and tissue damage as pro-inflammatory cytokines and adipokines are released from resident macrophages and adipocytes [113]. Increased cholesterol levels have been shown to intensify muscle wasting in the Duchenne muscular dystrophy (DMD) mouse model [114]. This suggests that hypercholesterolemia, through inflammation and fatty infiltration, can promote skeletal muscle atrophy. Age-related muscle atrophy, sarcopenia, reveal how low muscle mass and muscle strength are risk factors for disability and mortality [57]. Research on sarcopenia has suggested OPA1 as a potential therapeutic target for age-related muscle loss and chronic inflammation. Skeletal muscle sacrifices its own reserves to regulate metabolic dysfunction in lipid and glucose homeostasis in any state of health, which is why skeletal muscle atrophy is often observed in a range of diseases, such as infections to cancer [115]. Altered lipid metabolism plays a role in skeletal

muscle weakness, as shown in obesity-like disease models. In rats, short-term high fat diet impaired function in oxidativetype skeletal muscles [116]. For patients with spinal muscular atrophy (SMA), malnutrition is a major concern because of defects in fatty acid transport and mitochondrial β-oxidation [117, 118]. In addition to altered lipid metabolism, glucose metabolism alteration may cause skeletal muscle atrophy, as observed in diabetes [119]. Glucose is essential for skeletal muscle function because it is responsible for maintaining blood glucose homeostasis by metabolizing 80% of consumed glucose. It has been shown in mice, knock out for skeletal-muscle-specific endogenous circadian clock, Bmall, leads to glucose metabolism and systemic glucose homeostasis disruption [120]. A metabolic organ such as skeletal muscle is sensitive to homeostatic disturbances and may lead to imbalances of protein synthesis and degradation. Therefore, insulin-resistance may trigger skeletal muscle protein degradation through mTOR complex 1 signaling pathway (mTORC1) [121]. Hyperglycemia may also trigger muscle atrophy by WWP1/KLF15 pathway in diabetes [122]. Metabolic reprogramming may induce muscle mass wasting through impaired skeletal muscle specific mitochondrial pyruvate carrier (MPC), and upregulating the hepatic gluconeogenesis in the Cory cycle and fatty acid oxidation [123]. It has been hypothesized that microgravity-mediated chronic inflammation can develop into dysmetabolic conditions that include imbalance of lipid and glucose metabolism [124].

Future Prospective and Potential Targets

The main factors precipitating space-induced skeletal muscle atrophy are cholesterol and chronic inflammation (Figure 2). Based on the scientific literature available about inflammatory mediators, oxidative stress, hypercholesterolemia, altered metabolism, and muscular protein degradation, there is an obvious link between muscle atrophy and cholesterol. However, it is gravely important to point out the potential contraindications of implementing statins as a treatment for hypercholesterolemia because statins may also cause myopathies, such as myalgia and rhabdomyolysis [125, 126]. Additionally, glucocorticoids should not be considered as a potential therapy for chronic-inflammation-induced muscle atrophy because it increases the rate of the ubiquitin-proteasome system [127]. Hypercholesterolemic LGMD2B mice with muscle atrophy have developed fatty infiltration and inflammation in limb and girdle muscles histology, which suggests chronic cytokine release from adipocytes and macrophages [4]. Macrophage cytokine overexpression of IL-6, TNF-a, and IL-1 have been demonstrated to increase myogenic factors and FOXO transcription factors leading to the subsequent increase titin degradation through MuRF1 (TRIM63) [99, 128, 129]. With the acknowledgment of microgravity-induced atrophy occurring even with MuRF1 deletion in mice, the mechanism



Hoangvi Le, et al., J Biotechnol Biomed 2023 DOI:10.26502/jbb.2642-91280072

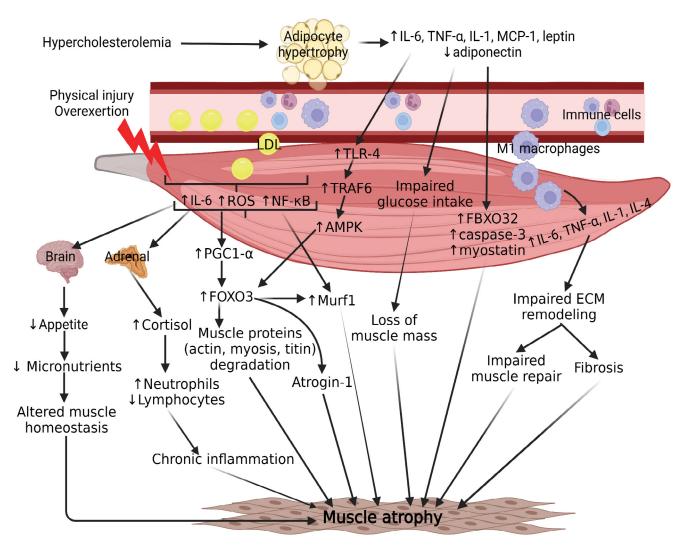


Figure 2: Chronic inflammation is strongly implicated in molecular mechanisms contributing to hypercholesterolemia-mediated skeletal muscle atrophy. The result of high cholesterol can increase adipocyte size and recruitment of immune cells, notably pro-inflammatory macrophages, into the peripheral skeletal muscle tissue. Both adipocytes and infiltrating macrophages contribute to a chronic signaling cascade of cytokines that ultimately lead to upregulation of fibrosis and atrogenes, such as MuRF1 and Atrogin-1. Physical injury or overexertion may further exacerbate chronic inflammation by inducing the release of muscle-specific cytokines, resulting in elevated cortisol levels from the adrenal gland and decreased appetite from the hypothalamus. This persistent crosstalk disrupts skeletal muscle homeostasis and contributes to muscle atrophy. Skeletal muscle is a metabolically sensitive organ and relies on the efficiency of circulating nutrients to maintain muscle mass. High concentrations of cholesterol in the blood have been demonstrated to cause muscle atrophy in peripheral artery disease due to poor circulation from plaque formation in the femoral arteries. Further investigation is needed to thoroughly understand the extent to which cholesterol plays a role in muscle atrophy.

between hypercholesterolemia and chronic inflammation in skeletal muscle must be investigated further to elucidate potential therapeutic targets. Studies investigating the interaction between skeletal muscle cytokines, also known as myokines, and other key organs, such as adipose tissue and the brain, have highlighted the multifaceted role that upregulated levels of IL-6 play in chronic inflammation and the decrease in appetite after exercise. Depending on the signaling origin, such as canonical myeloid cells or noncanonical adipocytes and muscle, IL-6 could play an proinflammatory or anti-inflammatory role in skeletal muscle, respectively [130]. However, IL-6 is a gravity-sensitive

myokine and may contribute to homeostatic dysregulation of muscle [131]. Elucidating the mechanisms of IL-6 and other myokines in microgravity would provide potential targets for developing anti-inflammatory therapies for astronauts. Space travel leads to an increase in production of reactive oxygen species (ROS), which causes cellular stress and damage to astronauts, including muscle atrophy.

The development of an antioxidant cocktail was proposed to maintain the health of astronauts and requires consideration of factors such as the physiological effects of ROS on the body, genetic predisposition of astronauts to damage,



and the efficacy of antioxidants. [132]. For ROS-related muscle atrophy, it was demonstrated that two antioxidants, N-acetyl-L-cysteine (NAC) and pyrroloquinoline quinone (PQQ), significantly decreased the development of skeletal muscle atrophy induced by fasting [49]. Exposure to the microgravity and radiation leads to destroyed red blood cells and excessive iron stores, which results in an imbalance in redox homeostasis, leading to oxidative damage of cells and injuries in the musculoskeletal system. Therefore, antioxidants and exogenous iron chelators were suggested as potential therapies [133]. An article reviewed the significance of certain nutrients in combating the harm brought about by microgravity during space missions. To mitigate oxidative stress, it was suggested to increase the consumption of antioxidants such as vitamins A, C, and E, omega-3 fatty acids, and minerals like copper, zinc, manganese, selenium, and iron through the diet. The combination of dietary defenses and the production of endogenous antioxidants could potentially play a significant role in guarding against oxidative damage. To address cardiovascular problems, it was advised to follow low-glycemic index diets. Vitamin D₃ was deemed important for preventing bone damage, but high doses of it could lead to hypercalcemia, kidney stones, and the calcification of soft tissues [134]. The TCA cycle could be another target for microgravity-induced atrophy as all participating enzymes were reported to have low gene and protein expression, such as citrate synthase, aconitase, isocitrate dehydrogenase, succinate dehydrogenase, fumarase, and malate dehydrogenase [5]. In muscle atrophy, there is a reduction in energy production, which leads to decreased muscle function. By targeting the TCA cycle, it may be possible to increase energy production in the muscle and thus, counteract or prevent muscle wasting. Metabolomics and epigenomics are promising bioinformatic studies that focus on the shifts in environments and how that impacts gene expression. Fundamental areas of space biology research include oxidative stress, DNA damage, mitochondrial dysregulation, epigenetics, telomere length alterations, and microbiome shifts [135]. The Gene Lab database of NASA is useful for researchers interested in bioinformatic analyses of experiments performed in space. However, it limits human data due to privacy concerns and access must be authorized by the NASA Human Research Program. Processing of the bioinformatic data also takes time and requires interdisciplinary understanding of multiple statistical methods.

Conclusions

The area of hyperlipidemia-associated muscle atrophy in astronauts is important but is poorly investigated. Skeletal muscle and cholesterol share an evolutionary-conserved function in energy metabolism and storage. Overall, this review critically evaluated the efforts of NASA, or lack thereof, in the coordination of their life sciences departments. This was also highlighted in the October 2015 audit of NASA [136]. A critical description is also presented as to how the twin study is an example of the small sample size that does not represent the population of astronauts and could lead to vague conclusions about the effects of spaceflight. The basis of scientific research requires larger population and decrease the workload off the few astronauts who are in high demand. Despite the recent renewal of TRISH (Translational Research Institute of Space Health) by the NASA Human Research Program in December 2020, more biomedical researchers and physicians with aerospace background are needed to facilitate meaningful investigations. Aerospace medical programs need to be marketed as much as aerospace engineering programs. Besides military medical residencies, only one civilian residency program for aerospace medicine exists at the University of Texas Medical Branch (UTMB) and one civilian fellowship program at the Mayo Clinic [137]. Recently, UCLA medical school opened their new aerospace medicine fellowship program starting July 2022 [138]. Whether cholesterol levels increase or decrease due to microgravity, better protocols and experimental methods must be established to thoroughly investigate the role of cholesterol in astronaut health. Because cholesterol is an important biomarker for many co-morbidities of cardiovascular disease, it cannot be ignored in aerospace biomedical research.

Author Contributions

Concept and design: HL, VR, DKA; Literature Search: HL; Critical review and interpretation of the findings: HL, VR; Drafting the article: HL, VR, DKA; Revising and editing the manuscript: VR, DKA; Final approval of the article: HL, VR, DKA.

Funding

This work was supported by the research grants R01 HL144125 and R01HL147662 to DKA from the National Heart, Lung, and Blood Institute, National Institutes of Health, USA. The contents of this article are solely the responsibility of the authors and do not necessarily represent the official views of the National Institutes of Health.

Institutional Review Board Statement

Not applicable.

Informed Consent Statement

Not applicable

Data Availability Statement

Not applicable since the information is gathered from published articles.

Acknowledgments

None

Citation: Hoangvi Le, Vikrant Rai, Devendra K. Agrawal. Cholesterol: An Important Determinant of Muscle Atrophy in Astronauts. Journal of Biotechnology and Biomedicine. 6 (2023): 67-79.



Conflicts of Interest

The authors declare no conflict of interest.

References

- 1. Droppert PM. A review of muscle atrophy in microgravity and during prolonged bed rest. J Br Interplanet Soc 46(1993): 83-86.
- Vandenburgh H, J Chromiak, J Shansky, et al. Space travel directly induces skeletal muscle atrophy. Faseb j 13 (1999): 1031-1038.
- 3. NASA. Gap M23: Determine if factors other than unloading contribute to muscle atrophy during space flight (2022).
- 4. Sellers SL, Milad N, White Z, et al. Increased nonHDL cholesterol levels cause muscle wasting and ambulatory dysfunction in the mouse model of LGMD2B. J Lipid Res 59 (2018): 261-272.
- Higashibata A, et al. Microgravity elicits reproducible alterations in cytoskeletal and metabolic gene and protein expression in space-flown Caenorhabditis elegans. npj Microgravity, 2016. 2(1): p. 15022.
- Ward ZJ, et al. Projected U.S. State-Level Prevalence of Adult Obesity and Severe Obesity. New England Journal of Medicine 381(2019): 2440-2450.
- Sellers SL, et al. Increased nonHDL cholesterol levels cause muscle wasting and ambulatory dysfunction in the mouse model of LGMD2B. Journal of Lipid Research 59 (2018): 261-272.
- Lee JH, et al. Relationship between muscle mass index and LDL cholesterol target levels: Analysis of two studies of the Korean population. Atherosclerosis 325 (2021): 1-7.
- 9. NASA. Risk of Cardiovascular Adaptations Contributing to Adverse Mission Performance and Health Outcomes (2022).
- Smith SM, SR Zwart. Nutritional biochemistry of spaceflight. Adv Clin Chem 46 (2008): 87-130.
- 11. Rainey K. Biological rhythms in space and on Earth. NASA (2015).
- 12. Lee JH, et al, Relationship between muscle mass index and LDL cholesterol target levels: Analysis of two studies of the Korean population. Atherosclerosis 325 (2021): 1-7.
- 13. General, N.O.o.I., Final Report IG-17-018 NASA's Management and Development of Spacesuits (2017).
- General, N.O.o.I., Final Report IG-21-025 NASA's Development of Next-Generation Spacesuits (2021).

- 15. General, N.O.o.I., IG-22-005-NASA's Management of the International Space Station and Efforts to Commercialize Low Earth Orbit (2021).
- 16. Hanpanich O, et al. Lipid Accumulation in Vesselimitating Tubes under Microgravity Condition (2012).
- 17. Savioli G, et al. Dysbarism: An Overview of an Unusual Medical Emergency. Medicina 58 (2022): 104.
- Kaczerska D, et al. The influence of high-fat diets on the occurrence of decompression stress after air dives. Undersea Hyperb Med 40 (2013): 487-497.
- 19. Halliwell B, JM Gutteridge. Oxygen toxicity, oxygen radicals, transition metals and disease. Biochem J 219 (1984): 1-14.
- Luks AM, ER Swenson, P Bärtsch. Acute high-altitude sickness. European Respiratory Review 26 (2017): 160096.
- 21. Temte JL. Elevation of serum cholesterol at high altitude and its relationship to hematocrit. Wilderness Environ Med 7 (1996): 216-224.
- 22. Gonzales GF, V Tapia. Association of high altitudeinduced hypoxemia to lipid profile and glycemia in men and women living at 4,100m in the Peruvian Central Andes. Endocrinol Nutr 60 (2013): 79-86.
- 23. Møller P, et al. Acute hypoxia and hypoxic exercise induce DNA strand breaks and oxidative DNA damage in humans. Faseb j 15 (2001): 1181-1186.
- 24. Pena E, et al. Oxidative Stress and Diseases Associated with High-Altitude Exposure. Antioxidants (Basel) 11 (2022).
- 25. Harvard University Department of M. and B. Cellular, 5. Cholesterol Modulates Membrane Fluidity (2020).
- 26. Stahn AC, et al. Increased core body temperature in astronauts during long-duration space missions. Scientific Reports 7 (2017).
- 27. Halonen JI, et al. Outdoor temperature is associated with serum HDL and LDL. Environ Res 111 (2011): 281-287.
- 28. Keatinge WR, et al. Increased platelet and red cell counts, blood viscosity, and plasma cholesterol levels during heat stress, and mortality from coronary and cerebral thrombosis. Am J Med 81 (1986): 795-800.
- 29. Soejima S, et al. Hyperthermic sensitivity and cholesterol levels of mammalian cell lines in culture. Cancer Lett 60 (1991): 159-167.
- Wen X, et al. Effects of long-term heat exposure on cholesterol metabolism and immune responses in growing pigs. Livestock Science 230 (2019): 103857.
- 31. Giroud S, et al. Hibernating brown bears are protected



against atherogenic dyslipidemia. Scientific Reports 11 (2021).

- 32. Anderson AP, DJ Newman, RE Welsch. Statistical Evaluation of Causal Factors Associated with Astronaut Shoulder Injury in Space Suits. Aerosp Med Hum Perform 86 (2015): 606-613.
- Laughlin MS, et al. Shoulder Injury Incidence Rates In Nasa Astronauts, in ntrs.nasa.gov (2014).
- 34. Murray JD, et al. RATE OF SHOULDER SURGERY AMONG NASA ASTRONAUTS (2014).
- Abboud JA, JS Kim. The effect of hypercholesterolemia on rotator cuff disease. Clin Orthop Relat Res 468 (2010): 1493-1497.
- 36. Gatto AP, et al. Dyslipidemia is associated with risk for rotator cuff repair failure: a systematic review and metaanalysis. JSES Reviews, Reports, and Techniques 2 (2022): 302-309.
- 37. Boutwell M, et al. A Wearable Sensor to Mitigate Shoulder Injury in Astronauts. International Journal of Exercise Science: Conference Proceedings 2 (2020).
- 38. Loflin B, et al. Identification of shoulder joint clearance in space suit using electromagnetic resonant spiral proximity sensor for injury prevention. Acta Astronautica 170 (2020): 46-54.
- 39. Rajulu SL, GK Klute. A Comparison Of Hand Grasp Breakaway Strengths And Bare-Handed Grip Strengths Of The Astronauts, SML 3 Test Subjects, And The Subjects From The General Population. NASA Technical Paper 3286 (1993).
- 40. McFarland S. Analysis Of Potential Glove-Induced Hand Injury Metrics During Typical Neutral Buoyancy Training Operations. NASA Technical Reports Server (2016).
- 41. Kruth HS. Lipid deposition in human tendon xanthoma. Am J Pathol 121 (1985): 311-315.
- 42. Yang Y, H Lu, J Qu. Tendon pathology in hypercholesterolaemia patients: Epidemiology, pathogenesis and management. J Orthop Translat 16 (2019): 14-22.
- 43. Li D, et al. Xanthomatosis in bilateral hands mimicking rheumatoid arthritis: A case report. Medicine (Baltimore) 96 (2017): e9399.
- 44. NASA. Space Suit RoboGlove (SSRG) | T2 Portal. technology.nasa.gov (2018).
- 45. Walsh SK. Next Generation Life Support: High Performance Eva Glove (2015).
- 46. Dunbar B. NASA Food for Space Flight. nasa.gov (2004).

- 47. Quora. How Do Astronauts Eat During A Spacewalk? Forbes (2017).
- 48. Tang H, et al. Long-Term Space Nutrition: A Scoping Review. Nutrients 14 (2021).
- 49. Qiu J, et al. Mechanistic Role of Reactive Oxygen Species and Therapeutic Potential of Antioxidants in Denervationor Fasting-Induced Skeletal Muscle Atrophy. Frontiers in Physiology 9 (2018).
- 50. Smith S, et al. Human Adaptation to Spaceflight: The Role of Food and Nutrition Second Edition (NP-2021-03-003-JSC) (2021).
- 51. Silbernagel G, et al. Cholesterol synthesis is associated with hepatic lipid content and dependent on fructose/ glucose intake in healthy humans. Exp Diabetes Res 2012 (2012): 361863.
- 52. Spence JD, DJ Jenkins, J Davignon. Dietary cholesterol and egg yolks: not for patients at risk of vascular disease. Can J Cardiol 26 (2010): e336-339.
- 53. Volpi E, R Nazemi, S Fujita. Muscle tissue changes with aging. Curr Opin Clin Nutr Metab Care 7 (2004): 405-410.
- Liu HH, JJ Li. Aging and dyslipidemia: a review of potential mechanisms. Ageing Res Rev 19 (2015): 43-52.
- 55. Rosada A, et al. Hyperlipidemias in elderly patients: results from the Berlin Aging Study II (BASEII), a cross-sectional study. Lipids Health Dis 19 (2020): 92.
- 56. Tezze C, et al. Age-Associated Loss of OPA1 in Muscle Impacts Muscle Mass, Metabolic Homeostasis, Systemic Inflammation, and Epithelial Senescence. Cell Metabolism 25 (2017): 1374-1389.
- 57. Kalyani RR, M Corriere, L Ferrucci. Age-related and disease-related muscle loss: the effect of diabetes, obesity, and other diseases. Lancet Diabetes Endocrinol 2 (2014): 819-829.
- Sisto IR, M Hauck, RDM Plentz. Muscular Atrophy in Cardiovascular Disease. Adv Exp Med Biol 1088 (2018): 369-391.
- 59. Abdullah SM, et al. Long-Term Association of Low-Density Lipoprotein Cholesterol With Cardiovascular Mortality in Individuals at Low 10-Year Risk of Atherosclerotic Cardiovascular Disease. Circulation 138 (2018): 2315-2325.
- 60. Vallejo-VazAJ, etal. Low-Density Lipoprotein Cholesterol Lowering for the Primary Prevention of Cardiovascular Disease Among Men With Primary Elevations of Low-Density Lipoprotein Cholesterol Levels of 190 mg/dL or Above: Analyses From the WOSCOPS (West of Scotland Coronary Prevention Study) 5-Year Randomized Trial
- Citation: Hoangvi Le, Vikrant Rai, Devendra K. Agrawal. Cholesterol: An Important Determinant of Muscle Atrophy in Astronauts. Journal of Biotechnology and Biomedicine. 6 (2023): 67-79.



and 20-Year Observational Follow-Up. Circulation 136 (2017): 1878-1891.

- 61. Gao S, J Liu. Association between circulating oxidized low-density lipoprotein and atherosclerotic cardiovascular disease. Chronic Diseases and Translational Medicine 3 (2017): 89-94.
- 62. Prabhu AV, et al. Cholesterol-mediated Degradation of 7-Dehydrocholesterol Reductase Switches the Balance from Cholesterol to Vitamin D Synthesis. Journal of Biological Chemistry 291 (2016): 8363-8373.
- 63. Smith SM, SR Zwart, M Heer. Risk Factor of Inadequate Nutrition Human Research Program Human Health Countermeasures Element (2015).
- 64. Kim MR, SJ Jeong. Relationship between Vitamin D Level and Lipid Profile in Non-Obese Children. Metabolites 9 (2019).
- 65. Iwase S, T Mano. [Microgravity and autonomic nervous system]. Nihon Rinsho 58 (2000): 1604-1612.
- 66. NASA. NASA SPACEFLIGHT HUMAN-SYSTEM STANDARD VOLUME 2: HUMAN FACTORS, HABITABILITY, AND ENVIRONMENTAL HEALTH (2019).
- 67. NASA. Space Station Spacewalks (2022).
- 68. Watso JC, WB Farquhar. Hydration Status and Cardiovascular Function. Nutrients 11 (2019).
- 69. Cleary MA, MR Sitler, ZV Kendrick. Dehydration and symptoms of delayed-onset muscle soreness in normothermic men. J Athl Train 41 (2006): 36-45.
- 70. Shaheen NA, et al. Public knowledge of dehydration and fluid intake practices: variation by participants' characteristics. BMC Public Health 18 (2018): 1346.
- 71. Maiti A, S Daschakraborty. How Do Urea and Trimethylamine N-Oxide Influence the Dehydration-Induced Phase Transition of a Lipid Membrane? The Journal of Physical Chemistry B 125 (2021): 10149-10165.
- Campbell NR, et al. Dehydration during fasting increases serum lipids and lipoproteins. Clin Invest Med 17 (1994): 570-576.
- 73. Huff J, et al. Risk of Radiation Carcinogenesis Human Research Program Space Radiation Element, in Human Research Roadmap (2016).
- 74. Khouw AS, S Parthasarathy, JL Witztum. Radioiodination of low density lipoprotein initiates lipid peroxidation: protection by use of antioxidants. Journal of Lipid Research 34 (1993): 1483-1496.
- 75. Kuebodeaux RE, P Bernazzani, TTM Nguyen. Cytotoxic

and Membrane Cholesterol Effects of Ultraviolet Irradiation and Zinc Oxide Nanoparticles on Chinese Hamster Ovary Cells. Molecules 23 (2018).

- 76. Wong FL, et al. Effects of Radiation on the Longitudinal Trends of Total Serum Cholesterol Levels in the Atomic Bomb Survivors. Radiation Research 151 (1999): 736-746.
- 77. Multhoff G, J Radons. Radiation, Inflammation, and Immune Responses in Cancer. Frontiers in Oncology 2 (2012).
- 78. Boerma M, et al. Space radiation and cardiovascular disease risk. World J Cardiol 7 (2015): 882-888.
- Crucian B, et al. Incidence of clinical symptoms during long-duration orbital spaceflight. Int J Gen Med 9 (2016): 383-391.
- Crucian BE, et al. Immune System Dysregulation During Spaceflight: Potential Countermeasures for Deep Space Exploration Missions. Front Immunol 9 (2018): 1437.
- Tall AR, L Yvan-Charvet. Cholesterol, inflammation and innate immunity. Nat Rev Immunol 15 (2015): 104-116.
- 82. Ludtka C, et al. Macrophages in microgravity: the impact of space on immune cells. NPJ Microgravity 7 (2021): 13.
- Brechsler M, et al. Hyperlipidemia-triggered neutrophilia promotes early atherosclerosis. Circulation 122 (2010): 1837-1845.
- 84. Paul AM, et al. Neutrophil-to-Lymphocyte Ratio: A Biomarker to Monitor the Immune Status of Astronauts. Front Immunol 11 (2020): 564950.
- Tackett N, et al. Prolonged exposure to simulated microgravity diminishes dendritic cell immunogenicity. Scientific Reports 9 (2019).
- 86. Kalinski P, et al. Exhaustion of Human Dendritic Cells Results in a Switch from the IL-15R/IL-15- to IL-2R/IL-2-driven Expansion of Antigen-specific CD8<sup>+</ sup> T Cells. The Journal of Immunology 196 (2016): 196.16-196.16.
- 87. Vogel J, et al. Expression of Hypoxia-Inducible Factor 1α (HIF-1α) and Genes of Related Pathways in Altered Gravity. Int J Mol Sci 20 (2019).
- Czech MP, S Corvera. Signaling mechanisms that regulate glucose transport. J Biol Chem 274 (1999): 1865-1868.
- 89. Grice BA, et al. Excess membrane cholesterol is an early contributing reversible aspect of skeletal muscle insulin resistance in C57BL/6NJ mice fed a Western-style highfat diet. American Journal of Physiology-Endocrinology and Metabolism 317 (2019): E362-E373.
- 90. Barrientos G, et al. Membrane Cholesterol in Skeletal





Muscle: A Novel Player in Excitation-Contraction Coupling and Insulin Resistance. Journal of Diabetes Research 2017 (2017): 3941898.

- 91. Barrientos G, et al. Cholesterol removal from adult skeletal muscle impairs excitation–contraction coupling and aging reduces caveolin-3 and alters the expression of other triadic proteins. Frontiers in Physiology 6 (2015).
- 92. Ortega, A. and J. Mas-Oliva, Cholesterol effect on enzyme activity of the sarcolemmal (Ca2+ + Mg2+)-ATPase from cardiac muscle. Biochim Biophys Acta 773 (1984): 231-236.
- 93. Bastiaanse EML, KM Höld, A Van der Laarse. The effect of membrane cholesterol content on ion transport processes in plasma membranes. Cardiovascular Research 33 (1997): 272-283.
- 94. NASA. HRR Gap CV-203: Test countermeasures on the ISS against the spaceflight-induced changes in the cardiovascular system of importance for development of disease, in humanresearchroadmap.nasa.gov. (2022).
- 95. Gladstone JN, et al. Fatty Infiltration and Atrophy of the Rotator Cuff do not Improve after Rotator Cuff Repair and Correlate with Poor Functional Outcome. The American Journal of Sports Medicine 35 (2007): 719-728.
- 96. Gertz ML, et al. Multi-omic, Single-Cell, and Biochemical Profiles of Astronauts Guide Pharmacological Strategies for Returning to Gravity. Cell Rep 33 (2020): 108429.
- 97. Tuttle CSL, LAN Thang, AB Maier. Markers of inflammation and their association with muscle strength and mass: A systematic review and meta-analysis. Ageing Research Reviews 64 (2020): 101185.
- 98. Riley DA, et al. Skeletal muscle fiber, nerve, and blood vessel breakdown in space-flown rats. Faseb j 4 (1990): 84-91.
- 99. Perandini LA, et al. Chronic inflammation in skeletal muscle impairs satellite cells function during regeneration: can physical exercise restore the satellite cell niche? The FEBS Journal 285 (2018): 1973-1984.
- 100. Satriano J. Arginine pathways and the inflammatory response: Interregulation of nitric oxide and polyamines: Review article. Amino Acids 26 (2004): 321-329.
- 101. Nakanishi N, et al. Urinary Titin N-Fragment as a Biomarker of Muscle Atrophy, Intensive Care Unit-Acquired Weakness, and Possible Application for Post-Intensive Care Syndrome. Journal of Clinical Medicine 10 (2021): 614.
- 102. Reid MB, JS Moylan. Beyond atrophy: redox mechanisms of muscle dysfunction in chronic inflammatory disease. J Physiol 589 (2011): 2171-2179.

- 103. Zhang YG, et al. Actin-Binding Proteins as Potential Biomarkers for Chronic Inflammation-Induced Cancer Diagnosis and Therapy. Anal Cell Pathol (Amst) 2021 (2021): 6692811.
- 104. Du J, et al. Activation of caspase-3 is an initial step triggering accelerated muscle proteolysis in catabolic conditions. Journal of Clinical Investigation 113 (2004): 115-123.
- 105. Huang Z, et al. Effect of mammalian target of rapamycin signaling pathway on nerve regeneration. Biotarget 2 (2018): 18-18.
- 106. Peng J, et al. Muscle atrophy in Titin M-line deficient mice. Journal of Muscle Research and Cell Motility 26 (2006): 381-388.
- 107. Ulanova A, et al. Effect of L-Arginine on Titin Expression in Rat Soleus Muscle After Hindlimb Unloading. Frontiers in Physiology 10 (2019).
- 108. Ploutz-Snyder L, et al. HRP-47072 Evidence Report: Risk of Impaired Performance Due to Reduced Muscle Mass, Strength, and Endurance Human Research Program Human Health Countermeasures Element (2015).
- 109. Okada R, et al. Transcriptome analysis of gravitational effects on mouse skeletal muscles under microgravity and artificial 1g onboard environment. Scientific Reports 11 (2021): 9168.
- 110. Cadena SM, et al. Skeletal muscle in MuRF1 null mice is not spared in low-gravity conditions, indicating atrophy proceeds by unique mechanisms in space. Scientific Reports 9 (2019): 9397.
- 111. Bisserier M, et al. Cell-Free Mitochondrial DNA as a Potential Biomarker for Astronauts' Health. J Am Heart Assoc 10 (2021): e022055.
- 112. Nguyen HP, et al. The effects of real and simulated microgravity on cellular mitochondrial function. npj Microgravity 7 (2021).
- 113. Ferraro E, et al. Exercise-induced skeletal muscle remodeling and metabolic adaptation: redox signaling and role of autophagy. Antioxid Redox Signal 21 (2014): 154-176.
- 114. Milad N, et al. Increased plasma lipid levels exacerbate muscle pathology in the mdx mouse model of Duchenne muscular dystrophy. Skeletal Muscle 7 (2017).
- 115. Sartori R, V Romanello, M Sandri. Mechanisms of muscle atrophy and hypertrophy: implications in health and disease. Nature Communications 12 (2021).
- 116. Andrich DE, et al. Altered Lipid Metabolism Impairs Skeletal Muscle Force in Young Rats Submitted to a Short-Term High-Fat Diet. Front Physiol 9 (2018): 1327.



- 117. Li YJ. et al. Metabolic and Nutritional Issues Associated with Spinal Muscular Atrophy. Nutrients 12 (2020).
- 118. Deguise MO, et al. Abnormal fatty acid metabolism is a core component of spinal muscular atrophy. Ann Clin Transl Neurol 6 (2019): 1519-1532.
- 119. Rudrappa SS, et al. Human Skeletal Muscle Disuse Atrophy: Effects on Muscle Protein Synthesis, Breakdown, and Insulin Resistance—A Qualitative Review. Frontiers in Physiology 7 (2016).
- 120. Harfmann BD, et al. Muscle-specific loss of Bmal1 leads to disrupted tissue glucose metabolism and systemic glucose homeostasis. Skeletal Muscle 6 (2016).
- 121. Koopman R, CH Ly, JG Ryall. A metabolic link to skeletal muscle wasting and regeneration. Front Physiol 5 (2014): 32.
- 122. Hirata Y, et al. Hyperglycemia induces skeletal muscle atrophy via a WWP1/KLF15 axis. JCI Insight 4 (2019).
- 123. Sharma A, et al. Impaired skeletal muscle mitochondrial pyruvate uptake rewires glucose metabolism to drive whole-body leanness. eLife 8 (2019).
- 124. Strollo F, et al. Space Flight-Promoted Insulin Resistance as a Possible Disruptor of Wound Healing. Frontiers in Bioengineering and Biotechnology 10 (2022).
- 125. Tournadre A. Statins, myalgia, and rhabdomyolysis. Joint Bone Spine 87 (2020): 37-42.
- 126. Hansen KE, et al. Outcomes in 45 Patients With Statin-Associated Myopathy. Archives of Internal Medicine 165 (2005): 2671-2676.
- 127. Braun TP, DL Marks. The regulation of muscle mass by endogenous glucocorticoids. Frontiers in Physiology 6 (2015).
- 128. Peris-Moreno D, D Taillandier, C Polge. MuRF1/ TRIM63, Master Regulator of Muscle Mass. Int J Mol Sci 21 (2020).

- 129. Zanders L, et al. Sepsis induces interleukin 6, gp130/ JAK2/STAT3, and muscle wasting. J Cachexia Sarcopenia Muscle 13 (2022): 713-727.
- 130. Severinsen MCK, BK Pedersen. Muscle-Organ Crosstalk: The Emerging Roles of Myokines. Endocr Rev 41 (2020): 594-609.
- 131. Smith JK. IL-6 and the dysregulation of immune, bone, muscle, and metabolic homeostasis during spaceflight. NPJ Microgravity 4 (2018): 24.
- 132. Gómez X, et al. Key points for the development of antioxidant cocktails to prevent cellular stress and damage caused by reactive oxygen species (ROS) during manned space missions. npj Microgravity 7 (2021): 35.
- 133. Yang J, et al. Effects of Iron Overload and Oxidative Damage on the Musculoskeletal System in the Space Environment: Data from Spaceflights and Ground-Based Simulation Models. Int J Mol Sci 19 (2018).
- 134. Costa F, et al. Spaceflight Induced Disorders: Potential Nutritional Countermeasures. Frontiers in Bioengineering and Biotechnology 9 (2021).
- 135. Afshinnekoo E, et al. Fundamental Biological Features of Spaceflight: Advancing the Field to Enable Deep-Space Exploration. Cell 183 (2020): 1162-1184.
- 136. NASA. NASA Office of Inspector General Office of Audits NASA'S EFFORTS TO MANAGE HEALTH AND HUMAN PERFORMANCE RISKS FOR SPACE EXPLORATION National Aeronautics and Space Administration (2015).
- 137. ASMA. Aerospace Medical Association | Residency Programs & Related Courses. www.asma.org.
- 138. UCLA Space Medicine Fellowship program aims to prepare next generation of flight surgeons | UCLA Health. www.uclahealth.org (2022).